# Long non-coding RNA DLEU7-AS1 promotes the occurrence and development of colorectal cancer via Wnt/β-catenin pathway

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**Abstract.** – OBJECTIVE: To investigate the expression features of long non-coding RNA (IncRNA) DLEU7-AS1 in colorectal cancer (CRC), so as to further study its role in the occurrence and development of CRC and its potential regulatory mechanism.

**PATIENTS AND METHODS:** The expression levels of IncRNA DLEU7-AS1 in 82 pairs of tissues and para-carcinoma normal tissu detected via quantitative Real-time poly se chain reaction (qRT-PCR), and the corre of DLEU7-AS1 expression with pathological dexes of CRC and patients' prognosis was alyzed. Besides, the expressi EU7-A in CRC cells was further de RT-PCF ed expres The DLEU7-AS1 knockdo n model **BNA** in was established using a CRC cell lines HT-29 a d H ons or one of DLEU7-AS1 on ogical it-8 (CCKcells was analyze a Cell Cou 8) and transwell on/migration y. Finally, its potentia lec gated via m was inves

Western blotting. RESUL The results RT-PCR showed pression level of U7-AS1 in CRC that the ificantly higher than that in normal tiswas nd the fference was statistically sig-SU nific ared with those in patients with low D 51 expr on, the tumor stage in EU7-AS1 expression was igh ents the e rates of lymph node mes and di r metastasis were higher, and tas verall survival rate was lower. Compared the he negative control group, the cell invasion, and migration capaci-JIII were decreased significantly in DLEU7-AS1 down expression group. Moreover, the reof Western blotting revealed that the expressions of key proteins in Wnt/β-catenin pathway, including  $\beta$ -catenin, c-myc, and cyclinD1, were decreased in si-DLEU7-AS1.

SIONS: The consistent of DLEU7-AS1 animoantly increased to CRC, which is markassociated with CRC staging, lymph node astasis, distribution metastasis and poor progno-DLEU7-AS1 to promote the proliferation, inand migrouph capacities of CRC through the to  $/\beta$ -catenin pathway.

Words: pon-coding RNA, DLEU7-AS1, Colorectal can-

# Introduction

Colorectal cancer (CRC) is a kind of common malignant tumor seriously threatening the human life and health. Like other tumors, its pathogenesis remains unclear so far<sup>1</sup>. Heredity, diet, unhealthy lifestyle and precancerous lesions are closely associated with its occurrence<sup>2,3</sup>. More than half of CRC patients have suffered from micro-metastasis in clinic before radical surgery, which is the direct cause of postoperative metastasis and recurrence of CRC<sup>4</sup>. Its pathogenesis has not been fully clarified yet, so the difficulty in diagnosis and treatment is one of the important reasons for its high morbidity and mortality rate<sup>5</sup>. Therefore, clarifying the molecular mechanism of CRC metastasis, predicting and diagnosing CRC metastasis and predicting the prognosis, are important contents in the CRC research.

The occurrence and development of CRC is a long-term, multi-gene and multi-stage complex process. However, the exact molecular pathogenesis of CRC is still not clear. With the rapid development of molecular biology and gene diagnosis technique, etc., it has been found that its possible mechanism is the vicious cell transformation and irreversible gene modification caused by the long-term interaction between genetic and environmental factors, which is mainly manifested as the activation of oncogenes and inactivation of tumor suppressor gene<sup>6</sup>. These changes eventually make the cell-related key physiological functions out of control, including proliferation, apoptosis, differentiation and other signal transduction<sup>7</sup>. Despite great achievements, whether there are other genes or epigenetic regulatory mechanisms involved in its occurrence and development still need further study.

With the continuous deepening of research, researchers have found that non-coding RNA is closely related to the tumor occurrence and development<sup>8</sup>. 98% RNAs in human genome are the non-coding RNAs, while only 2% transcripts are the coding RNAs. Non-coding RNA is the functional RNA molecule that cannot be translated into protein, including long non-coding RNA (IncRNA) and short non-coding RNA (piRNA, siRNA, and microR) LncRNA is a kind of RNA fragment with de than 200 bp in length, which does not protein. The abnormal expression of Inc molecules can be detected in almost all tu tissues, and more and more st have co firmed that the lncRNA mg s an e le occu tremely important role i hce and RNA development of tumor<sup>19</sup> regulate the relevant prot **1-CO** many ways at diff nt level. play an extremely importa le in the p enesis of tumor<sup>11,12</sup>. The RNA in C s poorly understood<sup>13</sup>; some In and CCA<sup>7</sup> can promote s, such as CRNDE <sup>14,15</sup>, while some, ET, can inhibit AS5 and IncRN such a in the pethogenesis of CRC, lncRNA is CRC in t<sup>1</sup> activation of Wnt pathway and inv wth face receptor (EGFR) paepide ansforming growth facon of ay, n Autation and epithelial-me-TGF on, etc<sup>17,18</sup>., but the research sei mal tra. area is still at the initial stage. in

y, the expression levels of lncRNA EU7-AS1 in 82 pairs of CRC tissues and pareinoma normal tissues were analyzed, and the ect of DLEU7-AS1 on biological functions of CRC cells was investigated. Our results suggested that DLEU7-AS1 may serve as a new target in the treatment of CRC.

# **Patients and Methods**

#### Patients and CRC Samples

A total of 82 pairs of surgically rese mor tissue and para-carcinoma tiss of CRC patients were collected. ording to the 7<sup>th</sup> version of Union for Inter onal Cancer Control/American Joint Commit Cancer ph nod (UICC/AJCC) CRC tumor ly stasis (TNM) staging criteria, a<sup>1</sup> dients enro ere postoperative pa diagnosed as CRC via d not gical analysis, and the ive the pr perative anti-tumor the as radio herapy or chemotherapy is stu appro by the Aospital, Ethical Comp e of Rizha and patient ir families h en fully informed the the sa would be used for scientific research and sign informed consent.

## Lines and Reagen

our CRC cell lines (HCT-8, HT-29, HCT-116 SW-620) an he fetal human colonic epithe-1 ll line (FH were purchased from ATCC s VA SA); high-glucose Dulbecco's (M. modific medium (DMEM) and fetal bovine rum (FBS) were purchased from Life Technoloskville, MD, USA); cells were cultured in ator containing 5% CO<sub>2</sub> at 37°C, and the medium used was the high-glucose DMEM containing 10% fetal bovine serum (FBS).

## Transfection

The negative control (siRNA) and siRNA containing DLEU7-AS1 interference sequence (si-D-LEU7-AS1) were purchased from GenePharma (Shanghai, China). The cells were paved onto the 6-well plate and cultured until the cell density reached 70%. SiRNA transfection was performed using Lipofectamine 2000 (Invitrogen, Carlsbad, CA, USA) according to the instructions of manufacturer. Cells were collected after 48 h for quantitative Real-time polymerase chain reaction (qRT-PCR) and cell function test.

#### Cell Proliferation Assay

Cells were collected at 48 h after transfection and paved onto the 96-well plate (2000 cells/well). Cells were incubated for 6 h, 24 h, 48 h and 72 h, respectively, and added with Cell Counting Kit-8 (CCK-8) (Dojindo Laboratories, Kumamoto, Japan) reagent for incubation for another 2 h. Then the optical density (OD) of each well at the absorption wavelength of 490 nm was measured using the microplate reader and the data were analyzed.

#### Transwell Migration/Invasion Assay

At 48 h after transfection, the cells were digested with trypsin and resuspended using the serum-free medium. After cell counting, the cell density was adjusted to  $2.0 \times 10^5$ /mL; the transwell chambers containing and not containing matrix gel were placed in the 24-well plate; 200 µL cell suspension was added into the upper chamber, while 500 µL medium containing 10% FBS was added into the lower chamber. The plate was placed in an incubator at 37°C for incubation. After 48 h, the chamber was taken out, fixed with 4% paraformaldehyde for 30 min, stained with crystal violet for 15 min and washed with PBS. The inner face of basement membrane was carefully cleaned, and the inner-layer cells were removed. Cells passing through the membrane stained on the outer layer of basement membrane were observed under the microscope, and five visual fields were randomly selected for counting.

#### qRT-PCR

Total RNA was extracted from CRC cell lines and tissues using TRIzol reagent (Invitrogen, Carlsbad, CA, USA), and then RNA was sely transcribed into cDNA using Primes 2T-Reagent (TaKaRa, Otsu, Shiga, Japan). Th PCR was performed using SYBR<sup>®</sup>Premi Taq<sup>™</sup> (TaKaRa, Otsu, Shiga, Japan) and Step Plus Real-time PCR system (Apr iosysten Foster City, CA, USA). The primei LU7-AS were used for qRT-PCR: orward: 5'-GAGGGAGACACTT everse: AAC 5'-CACGTTGTTGG TT 'AAT-3', reforward: 5'-CCTG ACCCA 5'-GCT CACATCI GAA-3'. verse: lep One Data were ap ng the Ab. expression level of software and the rel  $e^{2^{-\Delta\Delta Ct}}$  method. mRNA w calculated us

# We n Blotting

d cells were lysed with cell lysis nsfe on ice **1**30 min and centrifuged buffer nd 4° 4000 or 15 min. The total proas calculated using the bioncen CA) protein assay kit (Pierce, ninic ack cin ord, IL, USA). The extracted protein was Ro 10% SDS-PAGE and then transferd onto the polyvinylidene difluoride (PVDF) brane (Millipore, Billerica, MA, USA). n blotting analysis was performed according to standard procedures.  $\hat{\beta}$ -catenin, c-myc, cyclinD1, Tublin primary antibodies, anti-mouse and anti-rabbit secondary antibodies were purchased from Cell Signaling Technology (Danvers, MA, USA).

#### Statistical Analysis

Statistical Product and Service (SPSS, Armonk, NY, USA) 22.0 was d for data processing, and data were prese as mean ± used for standard deviation ( $x \pm s$ ). The *t*-te continuous variables, and  $x^2$ st or F exact test was used for classif variables. an Meier method was use assess the pronts, a and survival time of Log-rank st es amor diffewas used to compare the at the rent curves. p < 0sugge erence was statistical gnificant.

# lts

# 207-AS1 was High, Expressed in CRC sues and Cell Lines

of DLEU7-AS1 in 82 pairs he expression C tissues a the corresponding para-car-0 issue nd CRC cell lines were decin PCR. The results showed that tected mpared with that in para-carcinoma tissues, ession level of DLEU7-AS1 was signiincreased in CRC tissues, and the difference was statistically significant (Figure 1A). Compared with that in intestinal mucosal epithelial cells (FHC), the expression of DLEU7-AS1 was significantly high in CRC cells, and the difference was statistically significant (Figure 2A). Moreover, the expression levels of DLEU7-AS1 in HT-29 and HCT-116 were the highest, so we selected these two cell lines for subsequent experiments.

## DLEU7-AS1 Expression was Correlated with Clinical Staging, Lymph Node Metastasis, Distant Metastasis and Overall Survival of CRC Patients

The expression of DLEU7-AS1 was divided into high expression group and low expression group according to the qRT-PCR results of DLEU7-AS1 expressions in 82 pairs of CRC tissues and para-carcinoma tissues. The correlation of DLEU7-AS1 expression with the patient's age, gender, tumor site, clinical staging, lymph node metastasis, and distant metastasis was analyzed *via x*<sup>2</sup>-test. The high-expression DLEU7-AS1 was positively correlated with CRC clinical staging, lymph node metastasis, but not correlated with age, gender and tumor site



(Table I). In addition, in order to investigate the relationship between the expression of DLEU7-AS1 and the prognosis of CRC patients, relevant follow-up data were collected. Kaplan-Mark survival curve revealed that the high-express. DLEU7-AS1 was associated with poor processis of CRC, and the higher the expression level of DLEU7-AS1 was, the worse the prognosis we be (p<0.001; Figure 1B). This was suggest that HOTTIP may serve as a procession of a line for predicting the prognosis (RCC.)

ΑD

Table I. Association of lr

# ckdown CDLEU7-AS1 Inhibited

In the probability of the effect of DLEU7-AS1 on the probability of CRC cells, the DLEU7-AS1 interfering expression model was fully established (Figure 2B) and its prolirate was detected *via* CCK-8 in control group and DLEU7-AS1 interfering expression group. The cell proliferation rate in si-DLEU7-AS1 group was significantly decreased compared with that in si-NC group (Figure 2C-D).

with clinicopathologic characteristics of CRC.

		DLEU7-AS1 expression		
Parameters	f cases	Low (%)	High (%)	<i>p</i> -value
Age (year				0.381
<60		22	13	
≥60	47	25	22	
Ge				0.170
Male	40	26	14	
Female	42	21	21	
or loca				0.598
n	56	31	25	
	26	16	10	
				0.011
T	46	32	14	
	36	15	21	
uph node metastasis				0.042
	48	32	16	
	34	15	19	
Distance metastasis	Distance metastasis			0.040
No	63	40	23	
Yes	19	7	12	



**Figure 2.** (*A*) qRT-PCR analysis qRT-PCR were used to verify to of HT-29 and HCT-116 cells with LEU7-, expressio, RC ciency of EU7-AS1 k, ckdo

17-A

Knockdown D. 17-AS1 Inh. 20 the cell Migration . Invasion

The eff of DLEU7-A the migration and apacities of CRC invasio was investigated swell migration/invasion assay. The resulvia 7 ay (Figure 3A-B) showed that the ts d atior cells par g through the membrane numb mbe er knockdown of DLEU7-Fransv as sig reduced compared with that gesting that the migration capain C group, inhibited. The results of invasion assay were cit those above (Figure 3C-D).

# ckdown of DLEU7-AS1 Inhibited the E. Assion of Wnt/ $\beta$ -catenin Signaling Patnway

In order to analyze the potential mechanism of DLEU7-AS1 in promoting the cell proliferation,

RC cell lines and intestinal mucosal epithelial cells FHC; (B) k, ockdown; (C-D) Growth curve analysis showing the cell growth

invasion and migration capacities, the changes in expressions of key proteins ( $\beta$ -catenin, c-myc, and cyclinD1) in Wnt/ $\beta$ -catenin pathway after knockdown of DLEU7-AS1 were detected *via* Western blotting. The results showed that the expression levels of the above-mentioned proteins were decreased significantly after knockdown of DLEU7-AS1 (Figure 4).

## Discussion

CRC is one of the common malignant tumors in the world. In recent years, the morbidity and mortality rates of CRC have been gradually increasing; besides, the early diagnosis rate of CRC patients in China is very low, and most of them have been in the middle and late stages when treated,



**Figure 3.** (A, B) (A, B) (CT-116 cells), we exceed with si-DLEU7-AS1 displayed significantly lower migration capacity; (C,D) HT-29 and CT-110 capacity is observed with si-DLEU7-AS1 displayed significantly lower invasion capacity.

avanced tumor accounts for the majority<sup>4</sup>. so th Th is, metastasis, and recurrence of diar CRC djuvant rapy after operation of have in the emphases in current ancea ch. Re es have shown that IncRNA role in a variety of diseases, in impor pla ing tumors. Many IncRNAs are abnormalin A CRC, which may play important es in the diagnosis, treatment, and prognosis of <sup>19-21</sup>. Therefore, searching for the abnormalessed lncRNA in CRC and analyzing its correlation with clinical prognosis will help improve the diagnosis and treatment levels of CRC, and improve the clinical prognosis of patients.

In this study, the expression of lncRNA DLEU7-AS1 in CRC and its role in the occurrence and development of CRC were investigated. First, the expressions of DLEU7-AS1 in 82 pairs of CRC tissues and para-carcinoma tissues were verified. The results showed that the DLEU7-AS1 expression was significantly up-regulated and positively correlated with CRC staging, lymph node metastasis, distant metastasis and poor prognosis. Thus, we believe that DLEU7-AS1 may play a tumor-promoting role in CRC. In order to further investigate the effect of DLEU7-AS1 on biological functions of CRC, the DLEU7-AS1 knockdown expression model was established using



**Figure 4.** Knockdown of DLEU7-AS1 expression significantly decreased the expression of  $\beta$ -catenin, c-myc and cyclin D1.

small-interfering RNA. The results of CCK-8 and invasion/migration assay revealed that DLEU7-AS1 could promote the occurrence and development of CRC and exert important functions in CRC. However, its specific molecular mechanismus unclear.

Wnt/ $\beta$ -catenin signaling pathway is an ortant signaling pathway related to tumor sis<sup>22</sup>. The abnormal activation of Wnt path in nasopharyngeal carcinoma sophag squamous carcinoma promo ation tumor<sup>23</sup>. Wnt/β-catenin sig ing path is activated in most gastric ca and r tes the proliferation of gastri and sion of Wnt-1 is as ated with voliferation, progression and mall cell prognosis o lung cancer<sup>25</sup> n 90% CR n occur upon the activation of sal Wnt/β-catenin siway<sup>26</sup>. When Wnt signaling pagnaling p ctivated,  $\beta$ -caten. thway. Il gather in the leading to the loss of epithelial structure, nucl a ph menon is significantly associated and asion appretastasis<sup>27</sup>. with t hether DLEU7-AS1 proxplor n ora

the other and development of CRC this is regulated Wnt/ $\beta$ -catenin, the changes in expressions of key proteins ( $\beta$ -catenin, c-myc, and Vnt/ $\beta$ -catenin pathway after knockwn of DLEU7-AS1 were detected *via* Western ing. The results revealed that the expression less of the above proteins were significantly decreased after knockdown of DLEU7-AS1, indicating that DLEU7-AS1 has a positive regulatory relation with Wnt/ $\beta$ -catenin pathway.

#### Conclusions The expression of DLEU7-AS1 is significantly increased in CRC, which is markedly as with CRC staging, lymph node metast metastasis and poor prognosis. DL -AS1 may promote the proliferation, invasio d migration capacities of CRC through regul the Wn $t/\beta$ -catenin pathway. Conflict of interest The authors declare no cont ferences 1) COLORECTAL CANCER ION (PDQ(R)): Health proal version. 20 C, RAMAPURAM JB, RI AK. The epigenomics of embryonic pathway signaling in colorectal cancer. Front armacol 2017; 8: 267. Renner AT, L HERTY M, REULAND DS. Colorectal g in average risk patients. Med cer scree arth 017; 101: 755-767. 4) MARI TROIANI T, CARDONE C, VITIELLO P, SFORZA V, Ciardiello D, Napolitano S, Della CC, Morgillo F, A, Cuomo A, Selvaggi F, Ciardiello F, Marti-. Present and future of metastatic colorectal cancer treatment: a review of new candidate targets. World J Gastroenterol 2017; 23: 4675-4688. 5) SHETIWY M, FADY T, SHAHATTO F, SETIT A. Standardizing the protocols for enhanced recovery from colorectal cancer surgery: are we a step closer to ideal recovery? Ann Coloproctol 2017; 33: 86-92. WRIGHT M, BEATY JS, TERNENT CA. Molecular markers for colorectal cancer. Surg Clin North Am 2017; 97: 683-701. 7) MAHASNEH A, AL-SHAHERI F, JAMAL E. Molecular biomarkers for an early diagnosis, effective treatment and prognosis of colorectal cancer: Current updates. Exp Mol Pathol 2017; 102: 475-483. 8) BHAN A, SOLEIMANI M, MANDAL SS. Long noncoding RNA and cancer: a new paradigm. Cancer Res 2017; 77: 3965-3981. KONDO Y, SHINJO K, KATSUSHIMA K. Long non-coding 9) RNAs as an epigenetic regulator in human cancers. Cancer Sci 2017; 108: 1927-1933. 10) BOLHA L, RAVNIK-GLAVAC M, GLAVAC D. Long noncoding RNAs as biomarkers in cancer. Dis Markers 2017: 2017: 7243968. 11) HEERY R, FINN SP, CUFFE S, GRAY SG. Long non-coding RNAs: key regulators of epithelial-mesenchymal transition, tumour drug resistance and cancer stem cells. Cancers (Basel) 2017; 9: pii: E38. 12) RAO A, RAJKUMAR T, MANI S. Perspectives of long non-coding RNAs in cancer. Mol Biol Rep 2017; 44: 203-218.

24

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