

# MiRNA-616 aggravates the progression of bladder cancer by regulating cell proliferation, migration and apoptosis through downregulating SOX7

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**Abstract. – OBJECTIVE:** To investigate the regulatory effect of microRNA-616 (miRNA-616) on cellular behaviors of bladder cancer and the potential mechanism.

**PATIENTS AND METHODS:** The expressions of miRNA-616 and SOX7 in bladder cancer tissues and cell lines were examined by quantitative Real Time-Polymerase Chain Reaction (qRT-PCR). The relationship between miRNA-616 and SOX7 was assessed through Dual-Luciferase Reporter Gene Assay. The regulatory effect of miRNA-616 and SOX7 on cellular behaviors of bladder cancer were evaluated through cell counting kit-8 (CCK-8), colony formation, cell migration assay, and flow cytometry.

**RESULTS:** MiRNA-616 was upregulated, whereas SOX7 was downregulated in bladder cancer tissues and cell lines. The overexpression of miRNA-616 attenuated the proliferation and migratory abilities, arrested cell cycle progression, inhibited G<sub>0</sub>/G<sub>1</sub> phase, and stimulated apoptosis in T24 cells. SOX7 was the target gene of miRNA-616, and its level was negatively regulated by miRNA-616. The knockdown of SOX7 enhanced the proliferation and migratory abilities, and attenuated apoptosis of bladder cancer cells.

**CONCLUSIONS:** MiRNA-616 accelerates bladder cancer cells to proliferate and migrate and inhibit apoptosis by downregulating SOX7. MiRNA-616/SOX7 may be potential therapeutic targets for bladder cancer.

**Keywords:** miRNA-616, SOX7, Bladder cancer.

## Introduction

Bladder cancer is the second most common urological malignancy in the United States, which ranks first in our country. In 2012, the International Agency for Research on Cancer (IARC)

reported 54,486 new cases and 26,820 death cases of bladder cancer in the United States<sup>1</sup>. In 2015, 740,000 new cases and 166,000 death cases of bladder cancer were reported in the United States<sup>2</sup>. The genetic polymorphisms, chromosomal abnormalities, and epigenetic changes are responsible for the occurrence and progression of bladder cancer<sup>3</sup>. The urothelial transitional cell carcinoma is the most prevalent subtype of bladder cancer<sup>4</sup>. For non-metastatic bladder cancer, surgery with post-operative intravesical instillation is the preferred therapeutic strategy<sup>5</sup>. Although great strides have been made in the surgical procedures and adjuvant chemotherapy, the prognosis of bladder cancer remains unsatisfactory<sup>6</sup>. Hence, it is of significance to develop new sensitive biomarkers and therapeutic targets for bladder cancer.

MicroRNAs (miRNAs) are a group of single-stranded, endogenous, non-coding short RNAs<sup>7</sup>. They are post-transcriptional regulators that degrade target genes or inhibit their translation by complementary base pairing with the 3'-untranslated region (3'-UTR) of the target genes<sup>8,9</sup>. Currently, a great number of miRNAs have been identified in human genome<sup>10,11</sup>. They widely participate in physiological and pathological processes<sup>12,13</sup>. Abnormally expressed miRNAs are involved in the development and progression of a variety of human diseases, such as atherosclerosis, diabetes mellitus, migraine, and cancer diseases<sup>14-16</sup>. Serving as oncogenes or tumor-suppressor genes, miRNAs could influence tumor progression<sup>17</sup>. It is believed that miRNAs are capable of diagnosing and treating malignant diseases.

MiRNA-616 is a newly discovered cancer-related miRNA. It is upregulated in serum samples of patients with gastric cancer<sup>18</sup>, lung cancer<sup>19</sup>, and prostate cancer<sup>20</sup>. In addition, miRNA-616

upregulates TFPI-2 in androgen-independent prostate cancer<sup>20</sup>. Our previous discovery found that miRNA-616 was upregulated in bladder cancer cells. In this paper, we mainly explored the biological role of miRNA-616 in the progression of bladder cancer.

## Patients and Methods

### Sample Collection

40 paired bladder cancer tissues and adjacent normal tissues were surgically resected, immediately placed in liquid nitrogen, and preserved at -80°C. None of the enrolled bladder cancer patients received preoperative anti-tumor therapies. Patients and their families in this study have been fully informed. This study was approved by the Ethics Committee of Shanghai Tongren Hospital.

### Cell Culture

The human bladder immortalized epithelium cells (SV-HUC-1) and bladder cancer cells (UMUC3 and T24) were provided by American Type Culture Collection (ATCC) (Manassas, VA, USA). The cells were cultured in Life Technologies' Modified Eagle's Medium (DMEM; Gibco, Rockville, MD, USA) containing 10% fetal bovine serum (FBS; Life Technologies, Gaithersburg, MD, USA), 100 UI/mL penicillin and 100 µg/mL streptomycin. The cells were maintained at 37°C, in a 5% CO<sub>2</sub> incubator. The medium was replaced every 2-3 days.

### Transfection

The transfection reagents were provided by GenePharma (Shanghai, China). The cells were pre-seeded in the 6-well plates and transfected using Jetfectamine 2000 (Polyplus, Carlsbad, CA, USA) at 70% of confluence. At 24-48 h, the cells were harvested for subsequent experiments.

### RNA Extraction and qRT-PCR

RNA extraction from cells was performed using TRIzol Reagent (Invitrogen, Carlsbad, CA, USA). RNA was purified by DNase I treatment and reversely transcribed into complementary deoxyribose nucleic acid (cDNA) using PrimeScript RT Reagent (TaKaRa, Otsu, Shiga, Japan). The obtained cDNA underwent qRT-PCR using SYBR<sup>®</sup> Premix Ex Taq<sup>™</sup> (TaKaRa, Otsu, Shiga, Japan). Glyceraldehyde 3-phosphate dehydrogenase (GAPDH) and U6 were

used as internal references. Each sample was performed in triplicate, and the relative level was calculated by the 2<sup>-ΔΔCt</sup> method. The primer sequences were as follows: miRNA-616: F: 5'-CTGCTCAAACCCTCCAATGACTT-3'; R: 5'-GTGTAACACGTCTATACGCCG-3'; SOX7: F: 5'-CAAGATGCTGGGAAAGT-3'; R: 5'-CCGTAAGTCTGAGTTGGGGT-3'; GAPDH: F: 5'-GGTGGTCTCCTCTACTTC-3'; R: 5'-GTTGCTGTAGCCAAATCGTTG-3'; U6: F: 5'-AGAGAAGATTTCATGGCCCC-3'; R: 5'-ATCCAGTGGCCTCCG-3'.

### Cell Counting Kit-8 (CCK-8)

The cells were seeded in the 96-well plate with 2×10<sup>3</sup> cells per well. At the appropriate time points, the absorbance value at 450 nm of each sample was recorded using the CCK-8 kit (Dojindo Laboratory, Kumamoto, Japan) for depicting the viability curves.

### Colony Formation Assay

The cells were inoculated in a 6-well plate with 1×10<sup>3</sup> cells per well and incubated for 2 weeks. Subsequently, the cells were fixed in 100% methanol and dyed with 0.5% crystal violet for 20 min. The colonies were finally captured and calculated using a microplate reader.

### Cell Cycle Distribution Determination

The cells were fixed in 70% ethanol overnight, incubated with RNase A, and dyed with Propidium Iodide (PI) for 25 min. Subsequently, cell cycle distribution was analyzed on the BD FACSCalibur<sup>™</sup> flow cytometry (FACSCalibur; BD Biosciences, Detroit, MI, USA).

### Cell Apoptosis Determination

The cells were collected, centrifuged at 1000 r/min for 3 min, and resuspended in 800 µL of phosphate-buffered saline (PBS). After dying with Annexin V and PI in the dark for 15 min, the distribution of normal, early-stage apoptotic, late-stage apoptotic, and necrotic cells were analyzed using flow cytometry.

### Transwell Migration Assay

The cells were inoculated in the upper side of the transwell chamber (Corning, Corning, NY, USA). In the bottom side, 700 µL of medium containing 10% FBS was applied. After 48 h of incubation, the cells migrated to the bottom side were subjected to fixation in methanol for 15 min, crystal violet staining for

20 min, and cell counting using a microscope. The number of migratory cells was counted in 5 randomly selected fields per sample (magnification 200×).

#### Dual-Luciferase Reporter Gene Assay

The potential binding sites between miRNA-616 and SOX7 were predicted by TargetScan. The cells were co-transfected with miRNA-616 mimics/NC and wt SOX7 3'UTR/mut SOX7 3'UTR using Lipofectamine 2000. After 24 h, the co-transfected cells were collected to determine the luciferase activity using a Dual-Luciferase Reporter Assay System (Promega, Madison, WI, USA).

#### Statistical Analysis

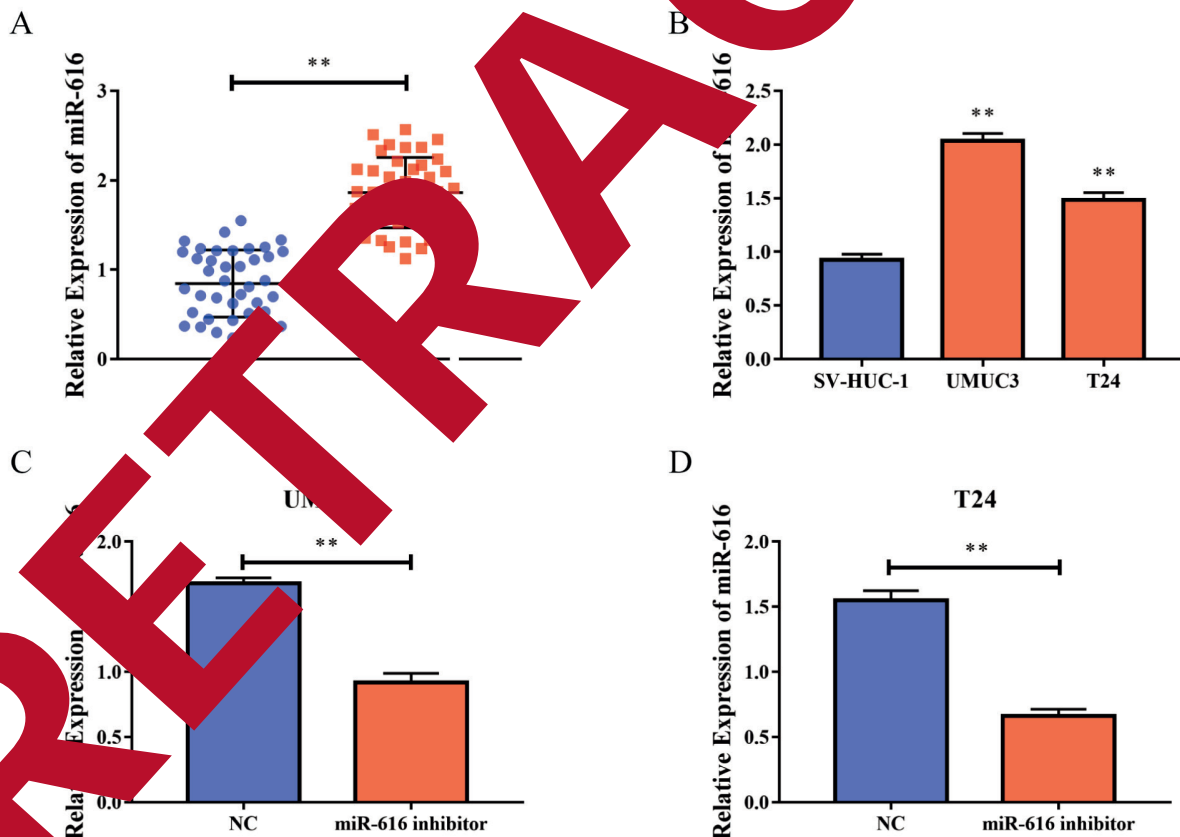
GraphPad Prism 5 (La Jolla, CA, USA) was used for the data analyses. The data were expressed as mean ± standard deviation. The intergroup differences were analyzed by the *t*-test.

The Spearman regression test was performed to evaluate the relationship between the two genes.  $p < 0.05$  was considered as statistically significant.

## Results

### MiRNA-616 Was Upregulated in Bladder Cancer

By analyzing the collected bladder cancer tissues and adjacent normal tissues, it is found that miRNA-616 was upregulated in bladder cancer (Figure 1A). Similarly, miRNA-616 was highly expressed in the bladder cancer cells than that of the bladder epithelial cells (Figure 1B). To uncover the biological function of miRNA-616 in bladder cancer, the miRNA-616 inhibitor was constructed. Transfection of miRNA-616 inhibitor in UMUC3 and T24 cells markedly downregulated miRNA-616 level (Figures 1C, 1D). It is indicated



**Figure 1.** MiR-616 was upregulated in bladder cancer. **A**, The relative level of miR-616 in bladder cancer tissues and adjacent normal tissues. **B**, The relative level of miR-616 in human bladder immortalized epithelium cells (SV-HUC-1) and bladder cancer cells (UMUC3 and T24). **C**, The transfection efficacy of miR-616 inhibitor in UMUC3 cells. **D**, The transfection efficacy of miR-616 inhibitor in T24 cells.

that miRNA-616 may exert a carcinogenic role in bladder cancer.

### **MiRNA-616 Regulated Proliferative, Migratory, and Apoptotic Abilities, and Cell Cycle Progression of Bladder Cancer**

The cellular behavior changes of bladder cancer influenced by miRNA-616 were explored through a series of functional experiments. CCK-8 assay showed a viability reduction in UMUC3 and T24 cells transfected with miRNA-616 inhibitor (Figures 2A, 2B). The silence of miRNA-616 also reduced the colony formation number in bladder cancer cells, suggesting the inhibited proliferative ability (Figures 2C, 2D). The apoptotic rate was found to be markedly elevated in UMUC3 and T24 cells transfected with miRNA-616 inhibitor (Figure 2E). The flow cytometry was conducted to assess cell cycle progression. The ratio of the cells distributed in the G2 phase significantly increased, and that in G1 phase decreased after the knockdown of miRNA-616 (Figures 2F, 2G). Moreover, the number of migratory cells was reduced after the transfection of miRNA-616 inhibitor (Figure 2H).

### **MiRNA-616 Bound to SOX7 3'UTR**

Through bioinformatics prediction, the binding sites between SOX7 and miRNA-616 were identified (Figure 3A). Based on these sites, we constructed wt SOX7 3'UTR and mutant SOX7 3'UTR vectors for performing the Dual-Luciferase Reporter Gene Assay. Luciferase activity markedly decreased after the transfection of miRNA-616 mimic and wt SOX7 3'UTR in UMUC3 and T24 cells, verifying the binding between miRNA-616 and SOX7 (Figures 3B, 3C). The transfection of miRNA-616 inhibitor remarkably upregulated the SOX7 level in bladder cancer cells (Figure 3D). The transfection efficacy of si-SOX7 was verified in both UMUC3 and T24 cells (Figure 3E). The miR-66 level was upregulated after transfection of si-SOX7, suggesting a negative relationship between the expressions of miRNA-616 and SOX7 (Figure 3F).

### **SOX7 Was Downregulated in Bladder Cancer Tissues**

Compared with the adjacent normal tissues, SOX7 was downregulated in bladder cancer tissues (Figure 4A). A negative correlation was observed between SOX7 and miRNA-616 in bladder cancer tissues (Figure 4B). Subsequently, we explored the influence of SOX7 on cellular be-

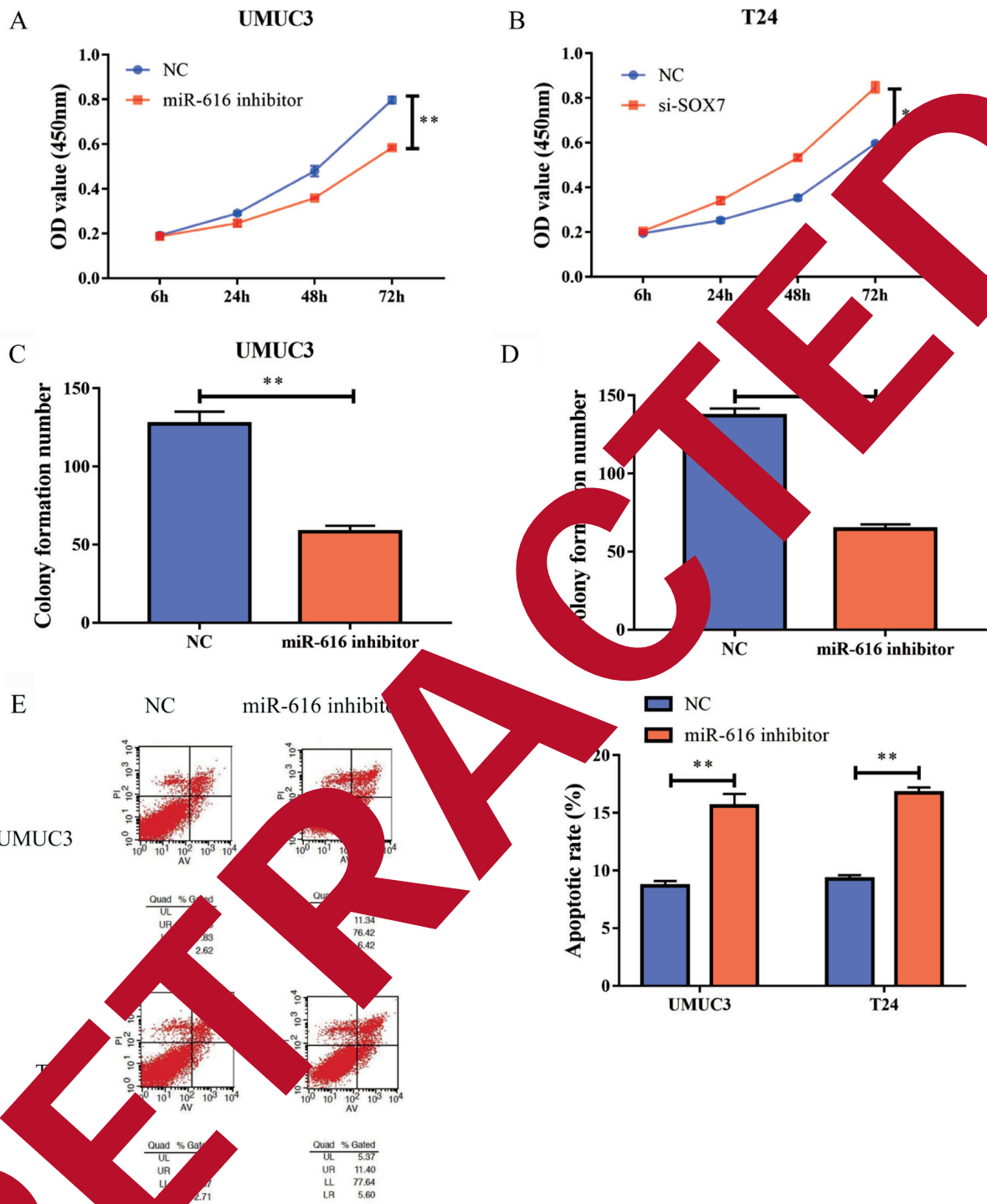
haviors of bladder cancer. As the viability curves revealed, the transfection of si-SOX7 elevated the viability in both UMUC3 and T24 cells (Figures 4C, 4D). The colony formation number was enhanced after the knockdown of SOX7, demonstrating the improved proliferative ability (Figure 4E). Meanwhile, the migratory cell number per field was markedly elevated after transfection of si-SOX7 (Figure 4G). On the contrary, the silence of SOX7 attenuated apoptosis of the bladder cancer cells (Figure 4F).

### **DISCUSSION**

Bladder cancer is one of the most common malignancies in males worldwide<sup>21</sup>. So far, radical cystectomy is the standard treatment for bladder cancer, and adjuvant chemotherapy effectively prolongs the survival in patients with muscle-invasive bladder cancer (MIBC) undergoing radical cystectomy<sup>23</sup>. However, the prognosis of bladder cancer is still poor due to high local recurrence and metastasis rates<sup>24</sup>. The identification of key molecules involved in bladder cancer progression is particularly important to improve therapeutic efficacy. Many upregulated and downregulated miRNAs in bladder cancer have been identified, which are closely related to the changes in cellular behaviors and signaling pathways in bladder cancer<sup>25</sup>.

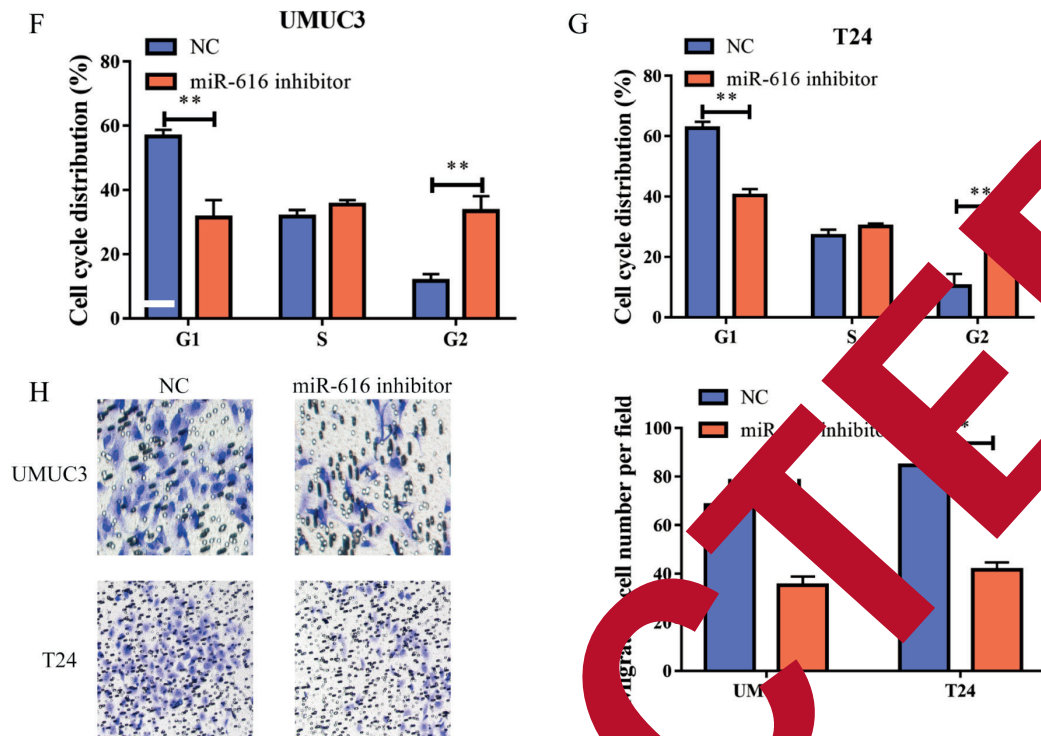
SOX genes (SOXs) exert an important role in the occurrence, migration and invasion of malignant tumors<sup>26</sup>. SOX7 locates on chromosome 8p23.1. It consists of two exons and an intron with a gene length of about 7.7 kbp. The SOX7 gene is composed of 388 amino acids. Its C-terminus is a transactivation domain that regulates the transcriptional activity and the N-terminus is an HMG-box DNA binding domain that specifically recognizes 5'-(A/T) (A/T) CAA (A/T)-3' sequence<sup>27</sup>. The transcriptional regulators encoded by the SOX gene are involved in the nervous system development, cartilage development, cardiovascular formation, and blood cell formation<sup>28</sup>. Meanwhile, these transcriptional factors are also involved in tumorigenesis and tumor development<sup>29</sup>. It is reported that SOX7 can inhibit the activation of the catenin and block catenin-induced transcription of the downstream genes, suggesting that SOX7 may be a potential checkpoint for bladder cancer<sup>30</sup>. In addition, the tumor proliferative and differentiation rates in SOX7<sup>-/-</sup> mice are significantly accelerated. The immunohistochem-



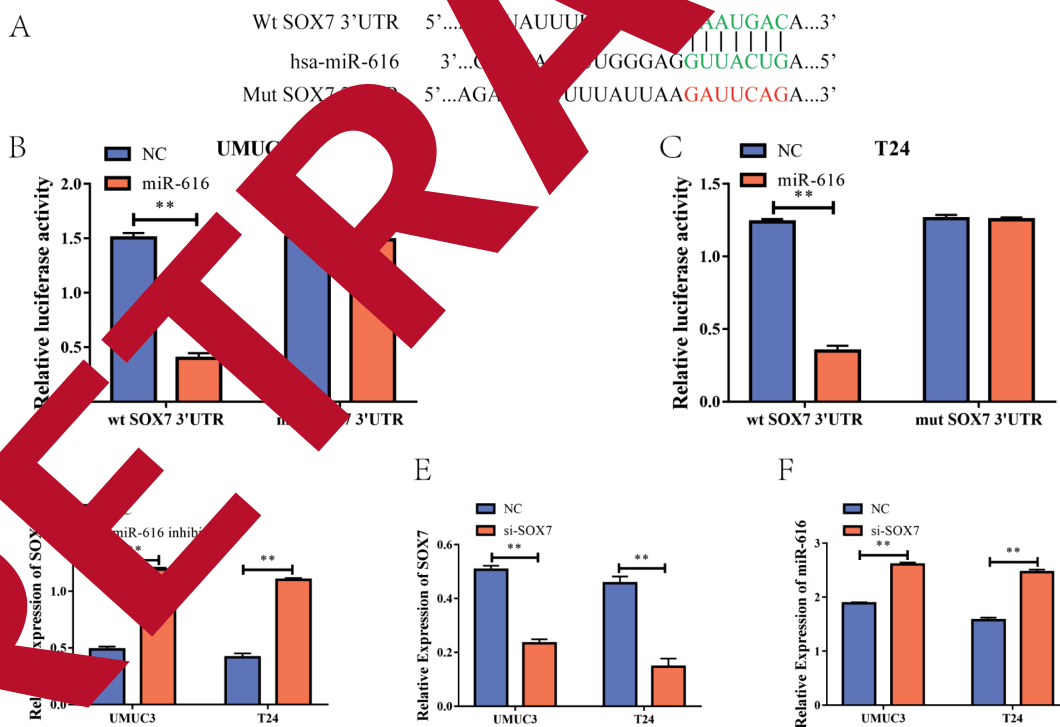


**Figure 2** MiR-616 regulated proliferative, migratory and apoptotic abilities, and cell cycle progression of bladder cancer. **A**, The viability in UMUC3 cells transfected with NC or miR-616 inhibitor. **B**, The viability in T24 cells transfected with NC or miR-616 inhibitor. **C**, The colony formation number in UMUC3 cells transfected with NC or miR-616 inhibitor. **D**, The colony formation number in T24 cells transfected with NC or miR-616 inhibitor. **E**, Apoptotic rate in UMUC3 and T24 cells transfected with NC or miR-616 inhibitor. **F**, The cell cycle distribution in UMUC3 cells transfected with NC or miR-616 inhibitor. **G**, The cell cycle distribution in T24 cells transfected with NC or miR-616 inhibitor. **H**, The migratory cell number per field in UMUC3 and T24 cells transfected with NC or miR-616 inhibitor (magnification: 40 $\times$ ).

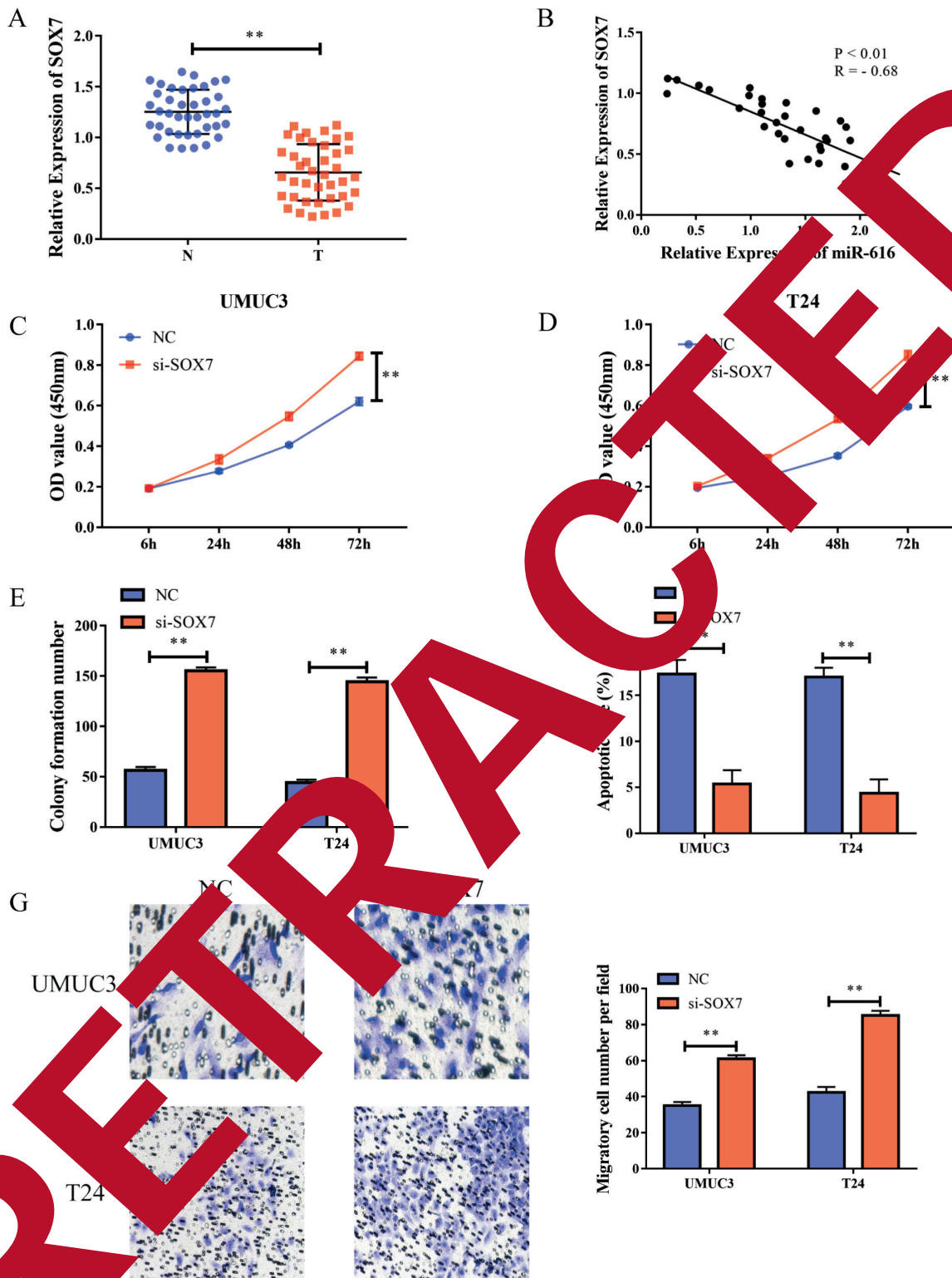
Figure continued



**Figure 2.** (Continued). **F**, The cell cycle distribution in UMUC3 cells transfected with NC or miR-616 inhibitor. **G**, The cell cycle distribution in T24 cells transfected with NC or miR-616 inhibitor. **H**, Migratory cell number per field in UMUC3 and T24 cells transfected with NC or miR-616 inhibitor (magnification: 40 $\times$ ).



**Figure 3.** MiR-616 bound to SOX7 3'UTR. **A**, The binding sites between miR-616 and SOX7. **B**, The luciferase activity in UMUC3 cells co-transfected with wt SOX7 3'UTR/mut SOX7 3'UTR and miR-616 mimics/NC. **C**, The luciferase activity in T24 cells co-transfected with wt SOX7 3'UTR/mut SOX7 3'UTR and miR-616 mimics/NC. **D**, The relative level of SOX7 in UMUC3 and T24 cells transfected with NC or miR-616 inhibitor. **E**, The transfection efficacy of si-SOX7 in UMUC3 and T24 cells. **F**, The relative level of miR-616 in UMUC3 and T24 cells transfected with NC or si-SOX7.



**Figure 4.** SOX7 was downregulated in bladder cancer. **A**, The relative level of SOX7 in bladder cancer tissues and adjacent normal tissues. **B**, A negative correlation between the expression levels of miR-616 and SOX7. **C**, The viability in UMUC3 cells transfected with NC or si-SOX7. **D**, The viability in T24 cells transfected with NC or si-SOX7. **E**, The colony formation number in UMUC3 and T24 cells transfected with NC or si-SOX7. **F**, The apoptotic rate in UMUC3 and T24 cells transfected with NC or si-SOX7. **G**, Migratory cell number per field in UMUC3 and T24 cells transfected with NC or si-SOX7 (magnification: 40 $\times$ ).

ical examination has found that SOX7 expression in prostate cancer is markedly downregulated<sup>31</sup>. SOX7 can be used as a tumor suppressor of prostate cancer. The downregulation of SOX7 in prostate cancer is associated with high methylation of promoters. It is found that SOX7 regulates the decomposition of active S33Y $\beta$ -catenin through APC-independent mechanism and thus exerts an anti-tumor effect<sup>32</sup>. In liver cancer, the upregulated SOX7 can impair the carcinogenic effect of microRNA-24<sup>33</sup>. A survival analysis showed that SOX7 is an independent prognostic factor for assessing overall survival and recurrence-free survival in lung adenocarcinoma<sup>34</sup>.

In this paper, miRNA-616 was upregulated, whereas SOX7 was downregulated in bladder cancer tissues and cell lines. The silence of miRNA-616 attenuated proliferative and migratory abilities, arrested cell cycle progression in G2 phase, and stimulated apoptosis in UMUC3 and T24 cells. SOX7 was the target gene of miRNA-616, and its level was negatively regulated by miRNA-616. The knockdown of SOX7 enhanced the proliferative and migratory abilities, and attenuated apoptosis of the bladder cancer cell lines.

## Conclusions

MiRNA-616 accelerates bladder cancer cells to proliferate and migrate and inhibits apoptosis by downregulating SOX7. MiRNA-616/SOX7 may be potential therapeutic target for bladder cancer.

## Conflict of Interest

The Authors declare that they have no conflicts of interests.

## References

- 1) CHENG Y, SHI Y, CHEN T, HU H, XIE W, QIAO Z, LIU L, LI S, WANG W, XING C, WANG Y, QIAN C. Genetic variations rs11892031 and rs40110 are associated with bladder cancer risk in a Chinese population. *Int J Mol Sci* 2014; 15: 19330-19341.
- 2) REGEL RL, MILLER KD, JEMAL A. Cancer statistics, 2015. *CA Cancer J Clin* 2015; 65: 5-29.
- 3) GAO JA. Molecular pathogenesis of bladder cancer. *Int J Clin Oncol* 2008; 13: 287-297.
- 4) SCELO G, BRENNAN P. The epidemiology of bladder and kidney cancer. *Nat Clin Pract Urol* 2007; 4: 205-217.
- 5) WU D, ZHOU Y, PAN H, ZHOU J, FAN Y, QU P. MicroRNA-99a inhibiting cell proliferation, migration, and invasion by targeting fibroblast growth

- factor receptor 3 in bladder cancer. *Oncol Lett* 2014; 7: 1219-1224.
- 6) KIM WJ, BAE SC. Molecular biomarkers in urothelial bladder cancer. *Cancer Sci* 2008; 99: 646-652.
- 7) AMBROS V. MicroRNA pathways in flies and mammals: growth, death, fat, stress, and learning. *Cell* 2003; 113: 673-676.
- 8) BARTEL DP. MicroRNAs: target recognition and regulatory functions. *Cell* 2009; 136: 215-233.
- 9) BARTEL DP. MicroRNAs: biogenesis, processing, mechanism, and function. *Cell* 2004; 119: 291-297.
- 10) WU W, SUN M, ZHANG L, CHEN Y. MicroRNA and cancer: current status and perspective. *Int J Cancer* 2007; 100: 95-100.
- 11) NELSON KM, HASS GJ. MicroRNAs and cancer: past, present, and potential. *Cancer Biol Ther* 2007; 8: 355-366.
- 12) GARZANO T, FALASCO A, CIMMINO A, CALIN GA, CROCE CM. MicroRNA expression and function in cancer. *Trends Mol Med* 2006; 12: 580-587.
- 13) GARCIA I, MISKOVIC M. MicroRNA functions in animal development and human disease. *Development* 2005; 132: 4653-4662.
- 14) TANA C, GIANNARDINO MA, CIPOLLONE F. MicroRNA profiling in atherosclerosis, diabetes, and migraine. *Appl Med* 2017; 49: 93-105.
- 15) WANG K, KOH T. MicroRNA epigenetic signature in human disease. *Arch Toxicol* 2016; 90: 2405-2419.
- 16) WANG Y, ZANG A, LI J, JIA Y, LI X, ZHANG L, HUO J, LIU J, FENG J, GE K, YANG Y, ZHANG Y, JIANG J. MicroRNA-383 is a tumor suppressor and potential prognostic biomarker in human non-small cell lung cancer. *Biomed Pharmacother* 2016; 83: 1175-1181.
- 17) CALIN GA, CROCE CM. MicroRNA signatures in human cancers. *Nat Rev Cancer* 2006; 6: 857-866.
- 18) YAO Y, SUO AL, LI ZF, LIU LY, TIAN T, NI L, ZHANG WG, NAN KJ, SONG TS, HUANG C. MicroRNA profiling of human gastric cancer. *Mol Med Rep* 2009; 2: 963-970.
- 19) RANI S, GATELY K, CROWN J, O'BYRNE K, O'DRISCOLL L. Global analysis of serum microRNAs as potential biomarkers for lung adenocarcinoma. *Cancer Biol Ther* 2013; 14: 1104-1112.
- 20) MA S, CHAN YP, KWAN PS, LEE TK, YAN M, TANG KH, LING MT, VIELKIND JR, GUAN XY, CHAN KW. MicroRNA-616 induces androgen-independent growth of prostate cancer cells by suppressing expression of tissue factor pathway inhibitor TFPI-2. *Cancer Res* 2011; 71: 583-592.
- 21) YU QF, LIU P, LI ZY, ZHANG CF, CHEN SQ, LI ZH, ZHANG GY, LI JC. MiR-103/107 induces tumorigenicity in bladder cancer cell by suppressing PTEN. *Eur Rev Med Pharmacol Sci* 2018; 22: 8616-8623.
- 22) WITJES JA. Bladder cancer in 2015: improving indication, technique and outcome of radical cystectomy. *Nat Rev Urol* 2016; 13: 74-76.
- 23) KITAMURA H, TSUKAMOTO T, SHIBATA T, MASUMORI N, FUJIMOTO H, HIRAO Y, FUJIMOTO K, KITAMURA Y, TOMITA Y, TOBISU K, NIWAKAWA M, NAITO S, ETO M, KAKEHI Y, UROLOGIC ONCOLOGY STUDY GROUP OF THE JAPAN



- CLINICAL ONCOLOGY GROUP. Randomised phase III study of neoadjuvant chemotherapy with methotrexate, doxorubicin, vinblastine and cisplatin followed by radical cystectomy compared with radical cystectomy alone for muscle-invasive bladder cancer: Japan Clinical Oncology Group Study JCOG0209. *Ann Oncol* 2014; 25: 1192-1198.
- 24) SALAMA A, ABDELMAKSOU D AM, SHAWKI A, ABDELBARY A, ABOULKASSEM H. Outcome of muscle-invasive urothelial bladder cancer after radical cystectomy. *Clin Genitourin Cancer* 2016; 14: e43-e47.
- 25) WEI Y, HE R, WU Y, GAN B, WU P, QIU X, LAN A, CHEN G, WANG Q, LIN X, CHEN Y, MO Z. Comprehensive investigation of aberrant microRNA profiling in bladder cancer tissues. *Tumour Biol* 2016; 37: 12555-12569.
- 26) CASTILLO SD, SANCHEZ-CESPEDES M. The SOX family of genes in cancer development: biological relevance and opportunities for therapy. *Expert Opin Ther Targets* 2012; 16: 903-919.
- 27) TAKASHI W, CANIZARES J, BONNEAUD N, POULAT F, MATTEI MG, JAY P, BERTA P. SOX7 transcription factor: sequence, chromosomal localisation, expression, transactivation and interference with Wnt signalling. *Nucleic Acids Res* 2001; 29: 4274-4283.
- 28) FRANCOIS M, KOOPMAN P, BELTRAME M. SoxF genes: key players in the development of the cardiovascular system. *Int J Biochem Cell Biol* 2010; 42: 445-448.
- 29) BONAZZI VF, IRWIN D, HAYWARD NK. Identification of candidate tumor suppressor genes inactivated by promoter methylation in melanoma. *Genes Chromosomes Cancer* 2009; 40: 101-110.
- 30) COSTA G, MAZAN A, GANDILLET A, PEANON S, LACAUD G, KOUSKOFF V. SOX7 regulates the expression of VE-cadherin in the haemopoietic endothelium at the onset of haematopoietic development. *Development* 2012; 139: 1597-1598.
- 31) CASTILLO SD, SANCHEZ-CESPEDES M. The SOX family of genes in cancer development: biological relevance and opportunities for therapy. *Expert Opin Ther Targets* 2012; 16: 903-919.
- 32) GUO L, ZHONG D, MAU S, ZHANG XY, SHI H, YANG VW, VERTINO P, MORENO DE WARMAN Y, ZHONG JT, ZHOU W. Sox7 is an independent checkpoint for beta-catenin activation in prostate and colon epithelial cells. *Mol Cancer Res* 2009; 6: 1421-1430.
- 33) MA Y, SHE XG, LI Y, LI YZ, WAN QQ. MiR-24 promotes the proliferation and invasion of HCC by targeting SOX7. *Tumour Biol* 2014; 35: 10731-10736.
- 34) LI B, GE Z, SONG S, ZHANG S, YAN H, HUANG B, ZHANG Y. Decreased expression of SOX7 is correlated with poor prognosis in lung adenocarcinoma patients. *Pathol Oncol Res* 2012; 18: 1039-1045.