MicroRNA-1269a promotes the occurrence and progression of osteosarcoma by inhibiting TGF-β1 expression

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> n OS cells while, mic 69a knocko ted the expressions of markedly own TGF-β1, p-Smad2, ad3, N-cad, Vimentin and M Furthermo F-β1 knockdown reecreased mig y and invasive abilm of OS cells. CONCLUSION MicroRNA-1269a is highly exich is remarkably correlated sed in OS, umor stag distant metastasis and poor w is of O In addition, microRNA-1269a pro prom alignant progression of OS by

regulating reaF-β1 expression.

Mis. RNA-1269a, TGF- β 1, OS, Malignant progression.

Introduction

Osteosarcoma (OS) is the most common primary malignancy. In recent years, the incidence of OS has gradually increased, accounting for 20% of all primary malignancies¹⁻³. OS is particularly common among adolescents and children, especially in those with 15-19 years old, with a prevalence of 5% of childhood tumors⁴. OS originates from immature tissues, such as osteoid tissue and mesenchymal tissue. Due to the rapid differentiation, proliferation, growth and invasion of OS cells, normal skeletal tissues are often affected and destroyed by malignant OS tissues⁵. As a bone tumor, OS frequently occurs in the proximal end of the long bone and the active end of the epiphysis. Meanwhile, it seriously affects the life quality of OS patients^{1,6}. Although improved surgical resection, adjuvant radiotherapy and chemotherapy techniques can achieve extensive removal of OS tissues and affected extremities,

genesis of osteosarcoma (OS) has not been reported. The aim of this work was to investigate the expression characteristics of microR-NA-1269a in OS and to further study its regulatory effects on the malignant progression of **PATIENTS AND METHODS:** The expres an microRNA-1269a in 61 pairs of OS tiss para-cancerous tissues was detected by ntitative Real Time-polymerase Chain Rea (qRT-PCR). Chi-square test was used to ana the relationship between micro -1269a pression and the characteris patient including age, sex, clinic tage distan icroRN metastasis. Subsequent 269a expression in OS cell lines dete After knockdown of icro structing relevant all intern RNA, biological performa of MG63 OS cells counting were accessed CCK-8). nswell assay. Meancolony forma n a rotein expl ns of key genes in while, the ad pathway w etected by Westthe EMT ern blg inally, si-TGF-β1 (t. forming growth) was transfected into OS cells, and cell fact vasion were detected by tranmi an

Abstract. - OBJECTIVE: MicroRNAs are en-

dogenous, non-coding small RNAs that are ca-

pable of regulating biological and pathological

processes. Previous studies have shown that

microRNA-1269a serves as an oncogene. How-

ever, the role of microRNA-1269a in the patho-

swell MicroP RESU 1269a was highly exed in compared with para-can-22 gh expression of microRtiss sitively correlated with young 69a was NA tients and high rate of distant metastasis, OS not correlated with age, sex and neking stage. Kaplan-Meier survival curves wed that high expression of microRNA-1269a gnificantly associated with poor progno-OS. The knockdown of microRNA-1269a sis in MG63 and U2OS cells significantly inhibited cell proliferation, migration and invasion. Mean-

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the recurrence rate and mortality of OS remain high^{2,6}. High malignancy and metastatic rate result in the poor prognosis of OS patients^{6,7}. It is estimated that the 5-year survival of postoperative OS patients is only 10%. About 40% of OS patients die from lung metastasis. Hence, it is of great significance to search for more efficient therapeutic targets for OS, thereby improving its clinical outcomes⁸⁻¹⁰.

The incidence of OS results from a complex network instead of a single mutation or deletion. The interaction of multiple unstable genome and proteins at the genetic level altogether leads to the pathogenesis of OS¹¹⁻¹³. So far, drugs targeting the immune system, intracellular and extracellular signaling for OS therapy have been well explored^{14,15}. In recent years, microRNAs, a small non-coding ribonucleic acid containing about 22 nucleotides, have been found to be crucial in the process of tumor pathophysiology¹⁶.

MicroRNAs are a class of newly discovered non-coding small RNAs with approximately 19-22 nucleotides in length, which are widely found in eukaryotes^{16,17}. Most microRNAs originate from gene spacer sequence, and are distri in single-copy, multi-copy and other for completely or incompletely binding to the TR of target genes, microRNAs degrade or i the translation of the corresponding target nes. They may eventually regu ll proli ration, apoptosis and differ Unlik stiple ta t genes. siRNA, microRNAs have foration. Meanwhile, their regulati cell apoptosis and cell cyc are involvement conditions¹⁹. Studie ve show incidence a of microRNAs in elopment ially in tume of human dise seases²⁰. ons of different types It is suggested that the of micropers vary great nd they may serve nes or tumor-sup as onc or genes²¹. Mi--1269a is highly expressed in a variety of croB s as an oncogene²². In this study, t tur ma he expression of microRNA-1269a we ex lines. Moreover, we also QS th and g ed its role in the development of led to provide new directions 05 ur study treatment of OS. fo

dg growth factor-β (TGF-β1) is a ember or the TGF superfamily, and its potenole in tumors has been well concerned²³. It is parted that TGF-β1 is highly expressed in multiple malignant tumors, such as colon cancer, bladder cancer, ovarian cancer, kidney cancer, lymphoma, etc. TGF-β1 also exerts an essential

role in tumor invasion and metastasis²⁴⁻²⁶. The effects of TGF-B1 on tumors are not only complicated, but also contradictory. In the early stage of tumorigenesis, TGF-β1 can inhibit cell feration, initiate cell differentiation apoptosis. However, TGF-β1 prom the invasion and metastasis of tumor cells ing the malignant transformation²⁷. Therefore speculated whether microRNA-126 regula $GF-\beta1$ expression, thereafter inf icing the performances of OS.

Patie is and thods

Patients a

underwein surgical re-61 OS atien section in our hospit. e enrolled, including 39 d 6-48 (mean age of mal 2 female cas 4±9.56 years). All paties is were diagnosed and 2 firmed as OS. No significant differences in age gender were nd in these patients. OS tissues ara-cancer tissues were harvested and a 1 in -8 . Informed consent was obtaipre. d patients and their families. This ned fro. udy was approved by the Ethics Committee.

Samples

OS cell lines (HOS, U2OS, SOSP-9607, MG63, 143B and SaOS-2) and human osteoblast cell line (hFOB) were obtained from the American Type Culture Collection (ATCC) (Manassas, VA, USA). Cells were cultured in Roswell Park Memorial Institute-1640 (RPMI -1640) (HyClone, South Logan, UT, USA) containing 10% fetal bovine serum (FBS) (Gibco, Grand Island, NY, USA) and maintained in a 5% CO₂ incubator at 37°C. Culture medium was replaced every 2-3 days. Cell passage was performed until 90% of confluence.

Cell Transfection

es

Negative control (si-NC) and si-microR-NA-1269a were provided by GenePharma Biotechnology Co., Ltd. (Shanghai, China). Cells were first seeded into 6-well plates. When the confluency was up to 70%, cell transfection was performed according to the manufacturer's instructions of Lipofectamine 2000 (Invitrogen, Carlsbad, CA, USA). Cells were harvested 48 h later for the following experiments.

Cell Counting Kit-8 (CCK-8)

48 h after transfection, OS cells were seeded into 96-well plates with 2000 cells per well. After

culturing for 6 h, 24 h, 48 h and 72 h, respectively, CCK-8 (Dojindo Laboratories, Kumamoto, Japan) reagent was added in each well, followed by incubation for 2 h in the dark. Finally, the optical density (OD) value of each well was measured using a microplate reader.

Colony Formation Assay

48 h after transfection, OS cells were seeded into 6-well plates with 200 cells per well. OS cells were cultured in complete medium for 2 weeks. Culture medium should not be replaced in the first week to prevent discarding un-adherent cells. Meanwhile, the medium was replaced twice in the second week. Until colony formation, the cells were washed with phosphate-buffered saline (PBS) twice and fixed with 2 ml of methanol for 20 min. Subsequently, the cells were washed with PBS and stained with 0.1% crystal violet staining solution for 20 min. Finally, formed colonies were observed and captured using a microscope.

Transwell Assay

48 h after transfection, OS cells were digested and re-suspended in serum-free medium density was adjusted to 5.0×10^{5} /mL. Tr chamber containing Matrigel (BD Biosc es. Franklin Lakes, NJ, USA) or not was place 24-well plates. 200 µL of cell suspension col ning 1.0×10⁵ cells was added to per cha ber. Meanwhile, 700 µl of ntainin .1U 20% FBS was added to t lower d hber. 48 hours later, the chamber and the remo cells were fixed with par min. After washing e cells, vere stamed with 0.2% cryst let for 20 The inner layer cells wer removed. 1 ids were ample. The cells were randomly selected for a captured the number petrating cells was calcula

O. tative ceal Time-polymerase Chain Reac. T-PCR

acted from OS cell lines Total vas zol reagent (Invitrogen, Carsues t extracted RNA was reversely CA, US lsb ribed into cDNA using Primescript RT Reatra , Otsu, Shiga, Japan). QRT-PCR was rformed using SYBR® Premix Ex TaqTM (Ta-, Otsu, Shiga, Japan), and StepOne Plus Re-PCR System (Applied Biosystems, Foster a City, CA, USA). Primers used in this study were as follows: microRNA-1269a: 5'-CCCTCACA-GCAATTTTATAGCATCT-3'; U6: 5'-TGCGG-

GTGCTCGCTTCGGCAGC-3'; TGF- β 1: forward: 5'-CTCTCCGACCTGCCACAGA-3', reverse: 5'-AACCTAGATGGGCGCGCGATCT-3'; β -actin: forward: 5'-CCTGGCACCCAGCACAAT verse: 5'-TGCCGTAGGTGTCCCTTT relative expression levels of mRNA per calculated by the 2^{- $\Delta\Delta$ Ct} method and approved by ABI Step One software.

Western Blot

OS cells were lysed ken on ice for . 1000 g and centrifuged at 4 n for 15 n. a. was determined The concentration of to. 'A) pro using the bicinck mic ac deterrce, Rockie A). The mination kit. % sodium extracted p as separated crylamide gel electrophododecyl s nateresis (SDS-PAGE) transferred onto polydifluoride 🚺) membranes (Milvin e, Bulerica, MA, US,). Western blot was li ducted according to the standard procedures. (TGF-β1, p-Smad2, p-Smad3, ary antibod MP9 and GAPDH) and se-Vimentir N s (anti-mouse and anti-rabbit) antibe con by Cell Signaling Technology were Danvers, MA, USA).

cal Analysis

GraphPad Prism 6 V6.01 software (La Jolla, CA, USA) was used for all statistical analyses. Data were expressed as mean \pm standard deviation. *t*-test was used to compare the difference between the two groups followed by Least Significant Difference as the Post-hoc test. Kaplan-Meier method was used to calculate the survival time of OS patients, and the difference between the curves was compared by Log-rank test. *p*<0.05 was considered statistically significant.

Results

MicroRNA-1269a was Highly Expressed in OS Tissues and Cell Lines

We first examined the expression of microR-NA-1269a in 61 pairs of OS tissues and para-cancerous tissues by qRT-PCR. The results showed that microRNA-1269a was highly expressed in OS tissues when compared with the para-cancerous ones, and the difference was statistically significant (Figure 1A, 1B). Similarly, microR-NA-1269a expression in OS cell lines was determined as well. Compared with human osteoblast cell line (hFOB), microRNA-1269a was highly



expr

Figure 1. MicroRNA-1269a was highly express tissues and para-cancerous tissues detected by qR1 of microRNA-1269a expression in OS patients. Dat

expressed in OS cells, and the state was statistically significant (Figurat C). An g them, MG63 and U2OS cell line pressed bighest level of microRNA-1/59a, d for subsequent expressed and statistically significant (Figurate C). An g them, was statistically significant (Figurate C) and (Figurate C). An g them, was statistically significant (Figurate C) and (Figurate C) and (Figurate C). An g them, was statistically significant (Figurate C) and (Fig

MicroRNA-1 A pression we Correlated with Ch. Stage, Lymph Node Marstasis, Disc. Metastasis, and Carall Survival in Catients

a on the expression of microRNA-1269a B s patients were divided into the in sues high n grour nd the low expression ıр. Т nber. atients in each group was test was used to analyze the led. C en microRNA-1269a expreshship be rel nd the clinical characteristics of OS patiensic age, sex, clinical stage and distant tastasis. As shown in Table I, high expression icroRNA-1269a was positively correlated oung OS patients and high rate of distant metastasis, whereas was not correlated with age, sex and Enneking stage. Subsequently, follow-up data were collected to explore the relationship

ues and ceremon *A-B*, MicroRNA-1269a expression in OS NA-1269a expression in OS cell lines. *D*, Survival curves $p \pm SD$, *p < 0.05, **p < 0.01, ***p < 0.001.

between microRNA-1269a expression and the prognosis of OS patients. Kaplan-Meier survival curves showed that high expression of microR-NA-1269a was significantly associated with poor prognosis of OS patients. The higher expression level of microRNA-1269a indicated remarkably worse prognosis of OS (p<0.05) (Figure 1D). These results indicated that microRNA-1269a might serve as a new biological indicator for predicting OS prognosis.

Knockdown of MicroRNA-1269a Inhibited Cell Proliferation

To explore the effect of microRNA-1269a on the proliferation of OS cells, we first constructed the microRNA-1269a interference model. Meanwhile, the transfection efficiency of si-microR-NA-1269a was determined (Figure 2A, 2B). The CCK-8 results found that OS cells transfected with si-microRNA-1269a present a significantly decreased proliferative rate than those transfected with si-NC (Figure 2C, 2D). Similarly, microRNA-1269a knockdown remarkably decreased the colony formation ability of the OS cells (Figure 2E).

Knockdown of MicroRNA-1269a Inhibited Cell Migration and Invasion

Transwell assay was conducted to explore the regulatory effect of microRNA-1269a on the mised, suggesting the inhibited migratory and invasive abilities (Figure 3A, 3B). Similar results were obtained in U2OS cells (Figure 3C, 3D).

TGF-β1 Modulated MicroRNA-12



Figure 2. Knockdown of microRNA-1269a inhibited cell proliferation. A-B, Transfection efficacy of si-microRNA-1269a. C-D, Proliferative rate of MG63 and U2OS cells detected by CCK-8 assay after transfection of si-microRNA-1269a. E-F, Colony formation of MG63 and U2OS cells after transfection of si-microRNA-1269a. Data were expressed as mean \pm SD, *p<0.05.

Parameters		miR-1269a expression		
	Number of cases	Low (%)	High (%)	<i>p</i> -value
Age (years)				
<21	27	10	17	
≥21	34	22	12	
Gender				36
Male	39	19	20	
Female	22	10	12	
Enneking stage				0.796
IA	8	4	4	
IIA	15	5	10	
IIB	29	9	20	
III	9	3		
Distance metastasis	5			0
No	34	20	14	
Yes	27	9		



characteristics.of osteosarcoma.

matics analysis. QRT-PCR and Western blot results detected the mRNA and protein expressions of TGF- β 1 in 61 pairs of OS tissues and para-cancerous tissues. Compared with para-cancerous tissues, TGF- β 1 was lowly expressed in Office sues at both mRNA and protein levels 250 Additionally TGF- β 1 was also lowly expressed in OS cell to as than that of hFOB cell line (hence 4B). Sub-quently, we selected 16 pairs from pairs and DS tissues and para-cancerous tissues of the correlation between miroRNA-1269a and TGF- β 1. QRT-PCR data indi-



Figure 3. Knockdown of microRNA-1269a inhibited cell migration and invasion. *A-B*, Invasive and migratory abilities in MG63 cells after transfection of si-microRNA-1269a. *C-D*, Invasive and migratory abilities in U2OS cells after transfection of si-microRNA-1269a. Data were expressed as mean \pm SD, **p<0.01.



Figure 4. TGF-\beta1 modulated microR M 1269a expr sues and para-cancerous tissues. *B*, The second existing the with TGF- β 1 expression in OS cells of Known on of m p-Smad3, N-cad, Vimentin and the P9 in OS s. Data

ssion in alls n of mich s. Data wel xy

cated that the expression of microl 269a was negatively corrected with TGF- β 1 e. ession in OS tissues (Fig.are 4C)

Further re, small h rence plasmid of as constructed. TGF-B1 transfection eff si-TCF-β1 was verified by qRT-PCR ficaç (Figure 5A, 5B). Further resulern and knockd n of TGF-β1 markedly ts sho and invasive abilities of ease nigra 4G63 s cells (Figure 5C, 5D).

Kr kdown of MicroRNA-1269a Ated Expressions of Key enes in £MT/Smad Pathway

analyze the potential mechanism of mich A-1269a in regulating the biological performances of OS cells, we detected the expressions of relative genes in the EMT/Smad pathway. Western blot results revealed that mi**a human osteosarcoma cells.** *A*, TGF- β 1 expression in OS tislls. *C*, MicroRNA-1269a expression was negatively correlated -1269a downregulated the expressions of TGF- β 1, p-Smad2, xpressed as mean \pm SD, *p<0.05, **p<0.01, ***p<0.001.

croRNA-1269a knockdown in OS cells markedly downregulated the protein expression levels of TGF-β1, p-Smad2, p-Smad3, N-cad, Vimentin and MMP9 (Figure 4D).

Discussion

OS is a primary malignancy with high heterogeneous, which mainly affects adolescents and children^{1,4}. Long bones, such as femur, tibia, fibula and ilium, are frequently involved in OS. The prognosis and survival rate of OS patients remain poor even after surgery^{4,6}. Currently, surgery combined with multiple chemotherapy drugs is the preferred treatment for OS. However, drug resistance and adverse events resulted from chemotherapy drugs, including methotrexate, doxorubicin, cisplatin and ifosfamide, have greatly



For effect provide and invasion of OS. *A-B*, Transfection efficacy of TGF- β 1. *C-D*, Migration and MG63 and U2 after knockdown of TGF- β 1. Data were expressed as mean \pm SD, *p<0.05, **p<0.01,

efficiency^{8,9}. Therefotherape restric o ful' Aucidate the pathogenesis t is u ovel therapeutic targets^{11,12}. to se e molecular mechanism, inte-Ex ations of between genome and proteome, as well rag r signaling pathways lay the solid undation for developing a new treatment for Furthermore, ideal treatment should reduce of lung metastasis and tumor recurrence, th thereby improving the survival rate of OS^{12,15}.

invasion of

***p<0.00

Site-specific gaps or cleavage regions that occur on the metaphase chromosomes of microR- NA-1269a are regions of genomic instability²². Fragile sites are usually stable in somatic cells, but are deleted or rearranged in many tumor cells. MicroRNAs near the fragile sites are closely related to tumorigenesis, often accompanied by chromosome breakage^{19,20}. Previous studies have found that under pathological conditions, microRNA-1269a is closely related to the malignant progression of tumor cells. MicroRNA-1269a can regulate cell cycle, tumor invasion, hematopoietic differentiation, immune response and other biological functions¹⁶⁻¹⁸. Multiple target

genes of microRNA-1269a form a complex network regulation system, including c-Myc, BNIP3, IRS-1 and other cell cycle-related genes. This may eventually affect the incidence and development of tumors^{19,21,22}. However, the effects of microRNA-1269a on OS cells and its specific mechanism have not been reported yet. Previous studies have mostly focused on the role of microRNA-1269a in regulating the proliferation and apoptosis of tumor cells. Few studies have reported the regulatory effects of microR-NA-1269a on cell invasion and metastasis. The strong invasive and metastatic abilities of OS are the main reasons for its poor prognosis. In this work, we explored the role of microRNA-1269a in the proliferation, invasion and metastasis of OS cells. The results showed that microRNA-1269a was highly expressed in OS tissues, which was correlated with tumor stage, lymph node metastasis, distant metastasis and poor prognosis. We observed that microRNA-1269a might be an oncogene in the OS development. Furthermore, the knockdown of microRNA-1269a significantly inhibited the proliferation, migration and invasion of OS cells. However, the specific me sm remains unclear.

The TGF- β superfamily is a type of cyt es with multiple biological effects. Mature T includes 5 subtypes, namely TGF-β1-5. An them, TGF- β 1 is currently kp be as ciated with malignant tum nwhild on tun TGF-β1 exerts a dual eff In the hibit the early stage of tumorige it car proliferation of tumo rells hthe advantes the proliferation tumor ion, besides ced stage^{25,26}. In β1 itself. TGF-β1 recep and Smad also im-In this experiment, portant in tuniorigen. rescue ex iments found there was a muation between m. tual re NA-1269a and . In-depth researches on the regulatory TGE rok GF-In the incidence and progresmay co ibute to improving the sion o tmer nd prognosis of tumors. nosi rings good news to tumor undou amilies. ts and the pal verify whether microRNA-1269a promoted

al progression of OS by regulating $\beta F - \beta I$, we detected the TGF- βI expression in ells after microRNA-1269a knockdown. Our ndicated that the TGF- β 1 expression was regulated by microRNA-1269a, showing that microRNA-1269a promoted the malignancy of OS probably by regulating TGF- β 1.

Conclusions

We showed that microRNA-1269a is highly expressed in OS, which is remarkably co with tumor stage, distant metastasis ap gnosis of OS. In addition, microRN 269a promotes the malignant progression S by regulating TGF-β1 expression.

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