

MiR-195 inhibits myocardial fibrosis in hypertensive rats by regulating TGF β 1-Smad3 signaling pathway

Q. XU, X.-X. LIN, P. LIU, W. ZHANG, K. TANG, Y.-S. ZHAI, L.-H. LIU, W.-Y. MEI

Department of Cardiology, East Hospital, The First Affiliated Hospital, Sun Yat-sen University, Guangzhou, China

Abstract. – **OBJECTIVE:** The aim of this study was to investigate the effect of micro-ribonucleic acid-195 (miR-195) on myocardial fibrosis in hypertensive rats through the transforming growth factor beta 1 (TGF β 1)-Smad3 signaling pathway.

MATERIALS AND METHODS: Spontaneously hypertensive rats (SHRs) were selected in this study to establish the animal model. The content of miR-195 in the model group and control group was measured, respectively. Arterial blood pressure, liver function and myocardial function in the two groups were detected and examined. Pathological changes in rat myocardial tissues were detected via hematoxylin-eosin (HE) staining. After that, myocardial fibroblasts were collected and added with miRNA inhibitor and mimics to suppress and overexpress miR-195. Thereafter, Reverse Transcription-Polymerase Chain Reaction (RT-PCR) and Western blotting were employed to detect the mRNA and protein expression levels of check point kinase 1 (Chk1) and alpha-smooth muscle actin (α -SMA) (important molecules for proliferation and migration of myocardial fibroblasts) and the related pathway TGF β 1-Smad3. Furthermore, the effects of miR-195 on myocardial fibrosis in hypertensive rats through the TGF β 1-Smad3 signaling pathway were comprehensively observed.

RESULTS: Serum alkaline phosphatase (ALP), glutamic pyruvic aminotransferase (ALT) and creatine kinase (CK) levels in the SHR group were significantly higher than those of the normal group. Cardiac function examination showed that SHR group had significantly reduced cardiac shortening (FS, %) and ejection fraction (EF, %) in comparison with the normal group. However, systolic blood pressure, diastolic blood pressure, left ventricular end-diastolic dimension (LVEDd) and left ventricular end-systolic dimension (LVESd) were markedly elevated in the SHR group. In addition, the miR-195 expression level was remarkably increased in hypertensive rats. Histopathological changes in rat myocardial tissues were detected through HE staining. The results showed that the normal group had orderly arranged myocar-

dial cells. In contrast, SHR group showed disorderly arranged myocardial cells, thickened myocardial fibers and myocardial fibrosis. RT-PCR assay results revealed that the mRNA levels of Chk1, α -SMA, TGF β 1 and Smad3 in rat myocardial fibroblasts were significantly reduced in Mimics group ($p < 0.05$) and increased in Inhibitors group ($p < 0.05$). Western blotting results demonstrated that, compared with the control group, the protein levels of α -SMA, TGF β 1 and Smad3 in rat myocardial cells decreased significantly in Mimics group ($p < 0.05$). Opposite results were observed in Inhibitors group ($p < 0.05$). The above results suggested that the overexpression of miR-195 inhibited the expression of TGF β 1-Smad3 signaling pathway and related molecules, further repressing myocardial fibrosis.

CONCLUSIONS: MiR-195 participates in the development and progression of myocardial fibrosis in hypertensive rats through the TGF β 1-Smad3 signaling pathway. Furthermore, this can inhibit the development of myocardial fibrosis in hypertensive rats and prevent myocardial diseases.

Key Words:

MiR-195, TGF β 1-Smad3 pathway, Rat, High blood pressure, Myocardial fibrosis.

Introduction

High blood pressure (HBP), the increase of arterial systolic and/or diastolic pressure(s) ($\geq 140/90$ mmHg) in resting state, is the most important cause of myocardial fibrosis. Myocardial fibrosis is pathological damage caused by various cardiovascular diseases¹⁻³. The mechanism of hypertensive myocardial fibrosis is relatively complex, which is regulated by a variety of cell growth factors. At present, a few studies have paid much attention to hypertensive myocardial

fibrosis. Myocardial fibroblasts maintain the systole and diastole of a healthy heart. However, after activation, abnormally proliferated and differentiated myocardial fibroblasts will cause myocardial fibrosis. Meanwhile, this may affect the connection and contraction of myocardial cells, leading to the occurrence and development of cardiovascular diseases, including coronary arteriosclerosis, arrhythmia and cardiomyopathy^{4,5}.

MiRNAs are a group of small non-coding RNAs with about 22 nucleotides in length, which can regulate the gene expression. Multiple studies have manifested that micro-ribonucleic acids (miRNAs) are involved in the differentiation, proliferation and apoptosis of cells^{6,7}. They participate in the specific regulation of protein-coding and -non-coding genes mainly *via* binding to the 3'-untranslated region (3'-UTR) of target gene messenger RNAs (mRNAs)^{8,9}. Previous studies have revealed that miRNAs bind to the 3'-UTR of target mRNAs through base pairing, leading to instability of mRNAs. Recent researches have discovered that miRNAs are capable of modulating and participating in various cellular processes during normal development and disease occurrence. This may eventually contribute to the progression of many diseases¹⁰. MiRNAs can modulate 1/3 human genes. Meanwhile, they play important roles in physiological homeostasis, health and many diseases, including cell cycle, development, metabolism, various immune responses¹² and other processes¹³. Roles of miRNAs in the pathogenesis of different diseases have been widely studied. Current findings have shown that they are important regulators of cell proliferation in many diseases. Moreover, miRNA regulatory networks have attracted much attention in recent years¹⁴. For example, miR-195 participates in the systole and diastole of myocardium and myocardial fibrosis through multiple signaling pathways. Another study has found that miRNAs, including miR-129, miR-21, miR-13 and miR-195, are specifically expressed in myocardial fibrosis and other diseases¹⁵. Research evidence has suggested that the transforming growth factor beta 1 (TGF β 1)/Smad signaling pathway is involved in myocardial remodeling. Additional studies have revealed that miR-195 can inhibit the TGF β 1/Smad pathway, which significantly improved myocardial hypertrophy and fibrosis, in which TGF- β 1 plays a crucial role¹⁶. However, the regulatory role of miR-195 in myocardial fibrosis and its effect on myocardial fibrosis have not been fully elucidated. Moreover, the mechanism of activating myocardial fibroblasts remains unclear so far.

MiR-195 is an important regulator of various diseases. However, a few studies have investigated its role in the pathogenesis of myocardial fibrosis in hypertensive rats. Therefore, the aim of this work was to investigate the potential role of miR-195 in myocardial fibrosis in hypertensive rats through *in-vivo* and *in-vitro* experiments. Our findings might help to improve the theoretical basis for the influences of miR-195 on myocardial fibrosis and TGF- β 1/Smad3 signaling pathway in hypertensive rats.

Materials and Methods

Experimental Animals and Grouping

A total of 20 male spontaneously hypertensive rats (SHRs) aged 6 weeks and 10 Wistar rats weighing 250 g were enrolled in this study and subjected to adaptive feeding. All rats were randomly divided into two groups, including the SHR group ($n=10$) and normal control group (Wistar rats, $n=10$). This study was approved by the Animal Ethics Committee of Sun Yat-sen University Animal Center. All operations on animals were carried out according to relevant standards in the NIH Guide for the Care and Use of Laboratory Animals.

Culture and Grouping of Myocardial Fibroblasts

SHRs were disinfected and dissected under aseptic conditions. The heart was taken out, washed, cut into pieces and fully digested with collagenase. Then, they were homogeneously digested into cell suspension using a constant temperature water bath shaker. After that, the cell suspension was repeatedly centrifuged, and the supernatant was discarded. Subsequently, the cells were inoculated into a cell culture flask. Cells in the logarithmic growth phase were collected and divided into negative control group (NC group), miR-195 inhibitors group (Inhibitors group) and miR-195 mimics group (Mimics group). After starvation treatment, the cells were transfected, followed by 36 h of continuous culture.

Examination of Liver Function and Myocardial Function

To predict myocardial fibrosis in hypertensive rats in the clinic and provide important references for early diagnosis, liver function indicators, including glutamic pyruvic aminotransferase (ALT), alkaline phosphatase (ALP), and myocardial function index

creatine kinase (CK) were examined in this study. Blood samples were routinely collected from rats in the two groups of rats, followed by centrifugation to isolate the serum. Finally, detection was performed using an automatic biochemical analyzer.

Determination of Arterial Blood Pressure and Cardiac Physiological Function Indexes in Rats

Left ventricular function, including left ventricular end-diastolic dimension (LVEDd) and left ventricular end-systolic dimension (LVESd), fractional shortening (FS, %) and ejection fraction (EF, %), was examined *via* magnetic resonance imaging (MRI) and echocardiography (ECG). Each rat was subjected to electrocardiography using with a 10 MHz probe. Systolic blood pressure and diastolic blood pressure (mmHg) of the rat tail artery were measured in accordance with relevant instructions.

Changes in MiR-195 Content in Rats of Model Group and Control Group

Blood samples were first collected from rats in each group, followed by centrifugation to collect the serum. Collected samples were then compared to pre-treatment. Reverse Transcription Polymerase Chain Reaction (RT-PCR) assay was performed to detect the content of miR-195 in each group, and the original data were recorded.

Detection of Pathological Changes in Rat Myocardial Tissues Through Hematoxylin-Eosin (HE) Staining

Rats were sacrificed and the hearts and heart samples were separated. Hearts were treated with 4% paraformaldehyde at 4°C for 24 h. After washing with distilled water, the tissues were dehydrated with different concentrations of alcohol, and embedded with paraffin (5 μm). After deparaffinization, the tissues were hydrated with 95%, 90%, 80%, 75% and 50% ethanol, respectively, followed by HE staining (Boster, Wuhan, China). Finally pathological changes in myocardial tissue structure were observed under a light microscope.

Determination of mRNA Expression Levels of Checkpoint Kinase 1 (Chk1) and Smooth Muscle Actin (α-SMA) in Myocardial Related Pathway TGFβ1-Smad3

Total RNA was extracted from collected cells and synthesized into complementary deoxyri-

bose nucleic acids (cDNAs) in strict accordance with PrimeScript™ Kit (TaKaRa, Otsu, Japan). Subsequently, single-stranded cDNAs were amplified using a conventional reaction system and stored at -20°C for Polymerase Chain Reaction (PCR) amplification. Primer sequences for target genes and glyceraldehyde 3-phosphate dehydrogenase (GAPDH) were designed based on the sequences on GenBank. The expression levels of target genes were measured by quantitative RT-PCR. Specific primer sequences were shown in Table I. The relative expression levels of related genes in myocardial cells of rats were calculated by the 2^{-ΔΔCT} method.

Determination of Protein Expression Levels of α-SMA and Related Pathway TGFβ1-Smad3 through Western Blotting

The culture solution was discarded, and the cells were washed with Phosphate-Buffered Saline (PBS; Gibco, Grand Island, NY, USA) 3 times. Lysis buffer was added, followed by centrifugation to collect the supernatant. The protein concentration was determined according to the instructions of the bicinchoninic acid (BCA) assay kit (Pierce, Waltham, MA, USA). Total protein was extracted from myocardial cells, followed by boiling in Laemmli buffer at 100°C in a water bath for 8 min and centrifugation at 1000 g for 5 min. Extracted protein was separated by SDS-PAGE electrophoresis and transferred onto a membrane using a semi-dry transfer method. After blocking, the membranes were incubated with primary antibodies overnight. On the next day, the membranes were incubated with the corresponding secondary antibody. Finally, protein band scanning and

Table I. Primer sequences.

Target gene	Primer sequence (5'-3')
GAPDH	GACATGCCGCTGGAGAAAC AGCCCAGGATGCCCTTTAGT
Collagen I	TCAGCCCAAACCCCAAGGAGA CGCAGGAAGGTCAGCTGGATAG
Collagen II	TGATGGGATCCAATGAGGGAGA GAGTCTCATGGCCTTGCGTGT
α-SMA	GTCCCAGACATCAGGGAGTAA TCGGATACTTCAGCGTCAGGA
Chk1	GCCAGTGCCTTTTGTGGAAG TCTATGGCCCGCTTCATGTC
TGF-β1	TGT GGC TCC TAG TGT TGA CG GCA GTT TGG ACA GGA TCT GG
Smad3	GCTTCTTGACGAGAGAGTCTACGG TACTAACACTGGTGGCAGCACTGG

quantification was detected by an Odyssey scanner (Lincoln, NE, USA). GAPDH was used as an internal reference.

Statistical Analysis

Statistical Product and Service Solutions (SPSS) 20.0 analysis software (IBM, Armonk, NY, USA) was used for all statistical analysis. GraphPad Prism 5.0 (La Jolla, CA, USA) was applied for image plotting. Experimental results were expressed as mean ± standard deviation ($\bar{x} \pm SD$). $p < 0.05$ was considered statistically significant.

Results

Liver Function and Myocardial Function

To provide important references for early diagnosis in clinical practice, liver function indexes (ALT and ALP) and myocardial function index CK were examined. The results (Table II) showed that the levels of serum ALP, ALT and CK in the SHR group were overtly higher than those of the normal group ($p < 0.05$). These results implied that liver function and myocardial function indexes increased markedly in the development and progression of myocardial fibrosis in hypertensive rats.

Arterial Blood Pressure and Cardiac Function Indexes in Rats of Model Group and Control Group

Arterial blood pressure and cardiac function indexes in rats of the two groups are shown in Table III. The results indicated that the SHR group had significantly increased FS, elevated systolic blood pressure, diastolic blood pressure, LVEDd and LVEDs in comparison with normal group ($p < 0.05$). This suggests successful establishment of SHR model in rats.

Changes in MiR-195 Content in Model Group and Control Group

The content of serum miR-195 in rats of each group was detected through RT-PCR. The results

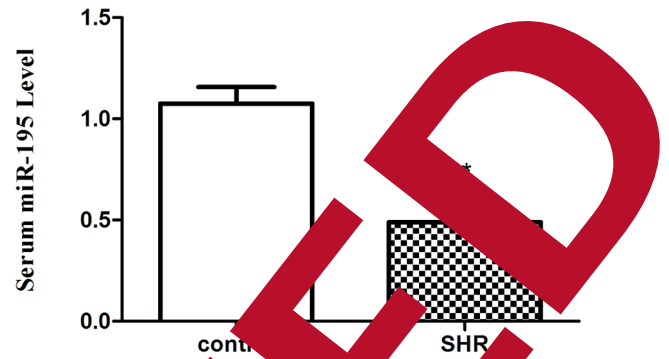


Figure 1. MiR-195 content in SHR group and control group. The content of miR-195 in SHR group was markedly lower than that in the control group ($p < 0.05$).

revealed that miR-195 content in the SHR group was significantly lower than that of the control group ($p < 0.05$). This implied that the expression level of miR-195 decreased in hypertensive rats (Figure 1).

Morphological Changes in Rat Myocardial Tissue Staining

The morphology and damage of rat myocardial tissues in each group were detected through HE staining. The results (Figure 2) showed that myocardial cells were arranged in order in the normal group (Figure 2A). However, they were disorderly arranged in the SHR group, with thickened muscle fibers and significant myocardial fibrosis (Figure 2B).

MRNA Expression Levels of Collagen, Chek1 and α-SMA and Related Pathway TGFβ1-Smad3 via RT-PCR

The results of RT-PCR assay showed that the mRNA expression levels of Collagen, Chek1, α-SMA, TGFβ1 and Smad3 decreased significantly in Mimics group when compared with the control group ($p < 0.05$). However, their expressions were all remarkably up-regulated in Inhibitors group ($p < 0.05$). The above findings suggested that miR-195 overexpression suppressed the expressions of TGFβ1-Smad3 signaling pathway and related molecules, further inhibiting myocardial fibrosis (Figure 3).

Protein Expression Levels of α-SMA and Related Pathway TGFβ1-Smad3 through Western Blotting

To further determine the key molecules in the fibrosis of myocardial fibroblasts and the effect of miR-195 on the TGFβ1-Smad3 signaling

Table II. Results of serum biochemical examination.

Group	ALP (U/L)	ALT (U/L)	CK (U/L)
Normal group	102.5±0.4	55.2±0.5	80.5±0.6
SHR group	217.6±0.1 ^a	125.5±0.3 ^a	198.9±0.4 ^a

Results: Serum ALP, ALT and CK levels in SHR group are significantly increased compared with those in the normal group ($p < 0.05$), indicating that liver function and myocardial function indexes are abnormal ($p < 0.05$).

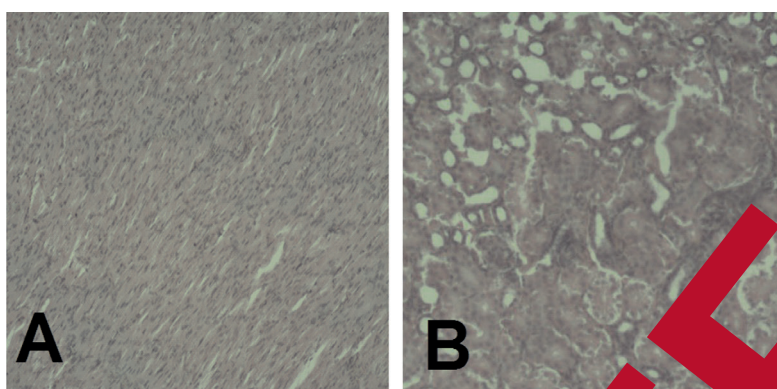


Figure 2. Pathological changes in rat myocardial tissues. Myocardial cells were arranged in order in the normal group (A), while was disorderly arranged in the SHR group (B), with thickened muscle fibers and myocardial fibrosis (magnification: 100×).

pathway, Western blotting was performed. The results (Figure 4) revealed that compared with the control group, the protein expression levels of α -SMA, TGF β 1 and Smad3 were significantly down-regulated in Mimics group ($p<0.05$). However, they increased markedly in Inhibitors group ($p<0.05$). These results indicated that overexpression of miR-195 inhibited the expressions of the TGF β 1-Smad3 signaling pathway and related molecules, further repressing myocardial fibrosis.

Discussion

MiRNAs are able to regulate the differentiation, proliferation and apoptosis of cells. Therefore, they play important roles in physiological homeostasis and health as well as various diseases¹⁸. The roles of miRNAs in the pathogenesis of different diseases have been widely investigated. MiRNAs have been considered as important gene expression regulators in various diseases. For example, miR-195 plays a key role in many processes of myocardial fibrosis in hypertensive rats. In

this study, we revealed that the content of miR-195 was significantly reduced in the SHR group. This indicates that the expression of miR-195 was inhibited. Moreover, the results of HE staining showed that myocardial cells were arranged orderly in the normal group. However, myocardial cells were arranged disorderly, muscle fibers were thickened, and myocardial fibrosis was detected in the SHR group. These results suggested that overexpression of miR-195 promoted the development and progression of myocardial fibrosis. To provide important references for early diagnosis in clinic, liver function indexes (ALP and GPT) and myocardial function index (CK) were examined as well. The results showed that serum levels of ALP, GPT and CK in the SHR group were overtly higher than those of the normal group ($p<0.05$). These results implied that liver function and myocardial function indexes increased significantly in the development and progression of myocardial fibrosis in hypertensive rats, which might provide an important reference for early diagnosis. Cardiac hemodynamic indexes and arterial blood pressure were also observed in this study. It was found that the SHR group

Table 1. Arterial blood pressure and cardiac function indexes determined via MRI & ECG ($\bar{x}\pm s$).

Group	LVEDd (mm)	LVESd (mm)	EF (%)	FS (%)	Systolic blood pressure (mmHg)	Diastolic blood pressure (mmHg)
Normal group	5.29±0.46	3.21±0.33	59±5.2	52.8±2.0	105.4±2.6	79.9±3.9
SHR group	8.12±0.51 ^a	6.28±0.52 ^a	48±2.6 ^a	38.4±2.8 ^a	151.3±4.9 ^a	115±5.8 ^a

Arterial blood pressure and cardiac function indexes. Compared with those in the normal group, the FS and EF in SHR group are evidently reduced, while the systolic blood pressure, diastolic blood pressure, LVEDd and LVESd are remarkably increased ($p<0.05$) ($p<0.05$).

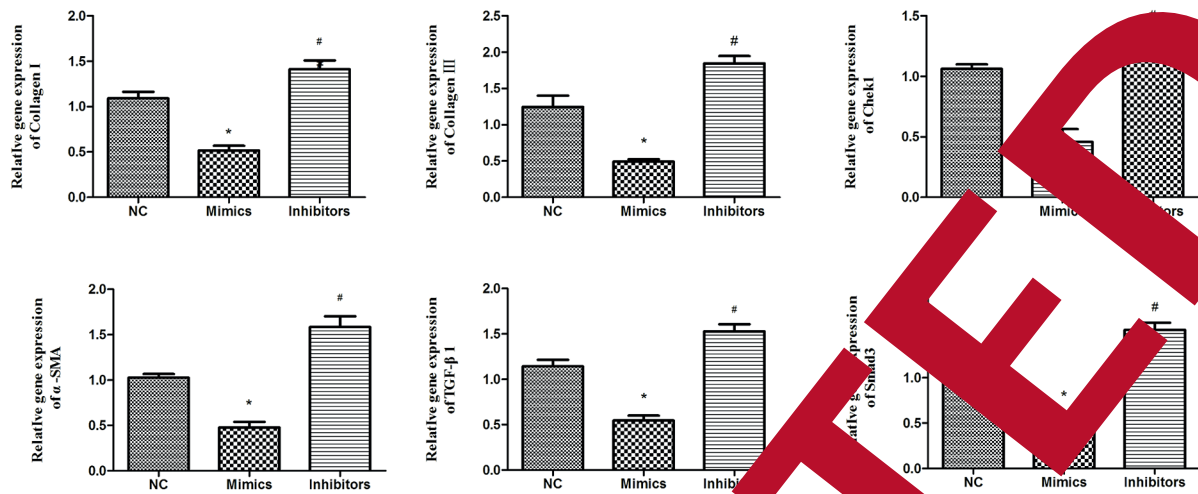


Figure 3. mRNA expression levels of Collagen, Chek1 and α -SMA and related pathway genes TGF- β 1 and Smad3 via RT-PCR. The expression levels of Collagen, Chek1, α -SMA, TGF- β 1 and Smad3 decreased significantly in Mimics group when compared with the control group ($p < 0.05$). However, they were remarkably elevated in Inhibitors group compared with the control group ($p < 0.05$) (* $p < 0.05$, # $p < 0.05$).

showed evidently reduced FS and EF and markedly increased systolic blood pressure, diastolic blood pressure, LVEDd and LVESd in compared with normal group ($p < 0.05$). This indicated that the selection of SHR model was a successful. Our results met the requirements for experimental animals, and further studies could be performed.

In summary, *in-vivo* results showed that inhibition of miR-195 expression promoted development of myocardial fibrosis, which was consistent with the findings of previous studies^{19,20}.

In addition, miR-195 targeted genes and proteins important for myocardial fibrosis, including Collagen, Chek1 and α -SMA. Chek1 is a key molecule regulating the mitosis and proliferation of myocardial cells. Meanwhile, its increased expression facilitates myocardial fibrosis. α -SMA, Collagen I and Collagen III are crucial components of myocardial fibroblast extracellular matrix. Their expression levels have been found to be markedly increased once myocardial fibrosis developed²¹⁻²³. This study discovered that Collagen, Chek1 and α -SMA were lowly expressed in Mimics group. The results suggested that overexpression of miR-195 inhibited the expression of genes related to the proliferation and migration of myocardial fibroblasts, thereby suppressing myocardial fibrosis development. Moreover, our findings indicated that miR-195 regulated multiple steps of myocardial fibrosis, affecting its development and progression. To

further investigate its effect, the protein expression level of α -SMA was detected by Western blotting. The results were consistent with those of mRNA expression levels, further confirming the regulatory role of miR-195 in myocardial fibrosis. In addition, these data were similar to the findings of predecessors^{24,25}.

TGF- β 1 acts as a regulator in the differentiation, proliferation and apoptosis of fibroblasts. The activation and binding of TGF- β 1 to its receptor can further trigger the phosphorylation of its downstream protein Smad²⁶. Researches have found that the activation of the TGF- β 1/Smad signaling pathway promotes the proliferation and migration of myocardial cells. Ultimately, this may increase cardiovascular resistance and induce cardiovascular remodeling²⁷. This study revealed that the expression levels of TGF- β 1 and Smad3 in rat tissues in Mimics group decreased significantly. Increased expression of miR-195 in myocardial cells reduced myocardial fibrosis in hypertensive rats. Meanwhile, it markedly lowered the expression levels of TGF- β 1 and Smad3 in myocardial cells. These results suggested that the activation of the TGF- β 1/Smad3 signaling pathway might be a leading cause of myocardial fibrosis in hypertensive rats. Furthermore, miR-195 might regulate myocardial fibrosis in rats by inhibiting the TGF- β 1/Smad3 signaling pathway, which was consistent with the findings reported by Lal H and Melchior-Becker A^{28,29}.

Conclusions

Overexpression of miR-195 suppresses the progression of myocardial fibrosis in hypertensive rats by inhibiting the TGF- β 1/Smad3 signaling pathway. Meanwhile, the miR-195/TGF- β 1/Smad3 axis regulates myocardial cells, playing an important role in the pathogenesis of myocardial fibrosis in hypertensive rats. Therefore, the miR-195/TGF- β 1/Smad3 axis can be used for the treatment of patients and the evaluation of therapeutic effect and prognosis.

Conflict of Interests

The Authors declare that they have no conflict of interests.

References

- 1) HOBBS R, KORUTLA V, SUZUKI Y, ACKERMAN S, VALLABHAPURU S, YULA P. Mechanical circulatory support as a bridge to definitive surgical repair after post-myocardial infarct ventricular septal defect. *Perf Surg* 2015; 30: 535-540.
- 2) TISMINETZKY M, ERSKINE N, CHEN HY, CHEN Y, GURWITZ J, YARZEBSKI J, JOFFE S, SHAPIRO P, GOLDBERG RJ. Long-term trends in, and characteristics associated with, patients not undergoing cardiac catheterization in elderly adults hospitalized with ST-segment elevation acute myocardial infarction. *Am Geriatr Soc* 2015; 63: 925-931.
- 3) CHEN HY, GURWITZ J, LAPANE JL, YARZEBSKI J, ERSKINE N, PERSON SD, GURWITZ J, JOFFE S, GOLDBERG RJ. A long-term perspective (1975-2009) into the long-term prognosis and hospital mortality of patients discharged from the hospital after ST-segment acute myocardial infarction. *Am J Cardiol* 2015; 115: 14-29.

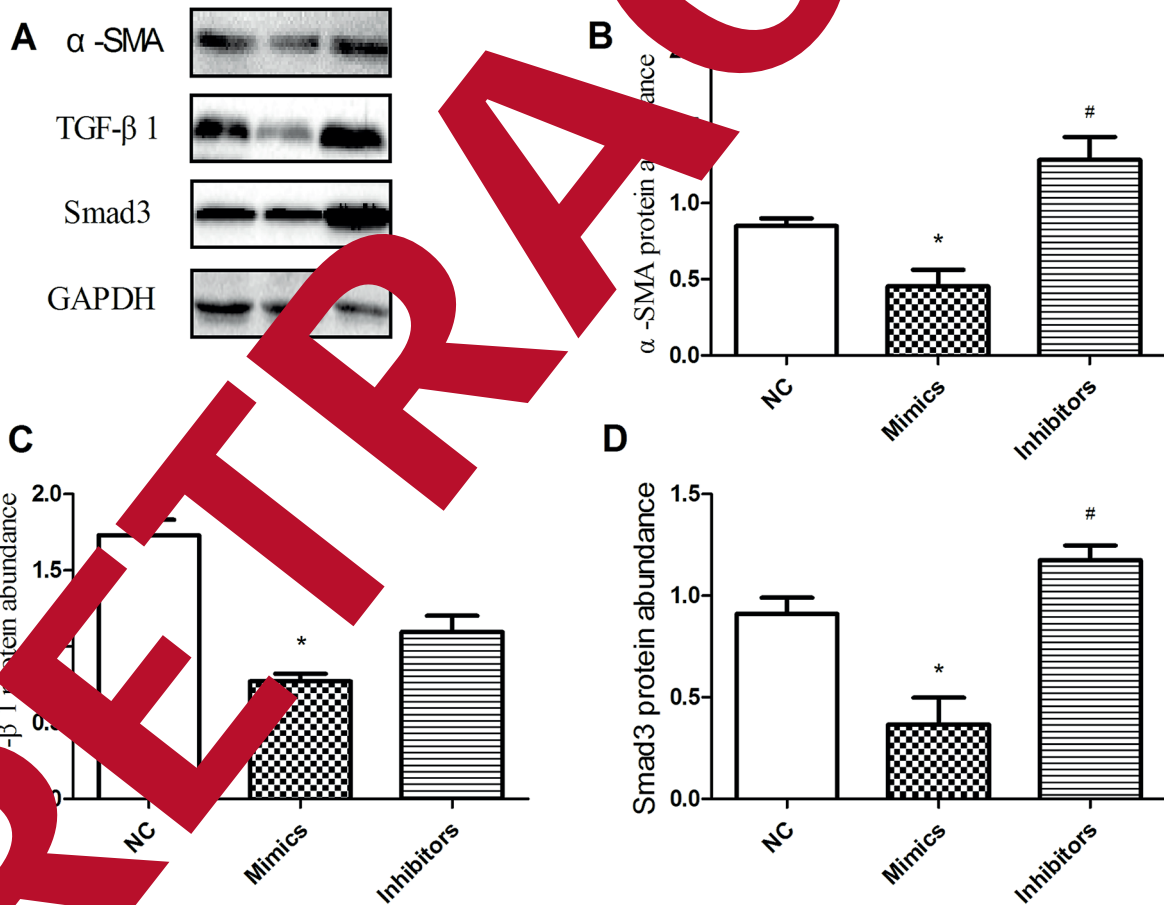


Figure 4. Protein expression levels of α -SMA and related pathway TGF β 1-Smad3 through Western blotting. The protein expression levels of α -SMA, TGF β 1 and Smad3 decreased significantly in Mimics group ($p < 0.05$), whereas increased markedly in Inhibitors group ($p < 0.05$). * $p < 0.05$, # $p < 0.05$.

- 4) WANG CC, SHANG BB, YANG CW, LIU YF, LI XD, WANG SY. MicroRNA-325 alleviates myocardial fibrosis after myocardial infarction via downregulating GLI1. *Eur Rev Med Pharmacol Sci* 2018; 22: 5339-5346.
- 5) HERSCOVICI R, KUTYIFA V, BARSHESHET A, SOLOMON S, McNITT S, POLONSKY B, LEE AY, ZAREBA W, MOSS AJ, GOLDENBERG I. Early intervention and long-term outcome with cardiac resynchronization therapy in patients without a history of advanced heart failure symptoms. *Eur J Heart Fail* 2015; 17: 964-970.
- 6) ZHU G, ZHANG W, LIU Y, WANG S. miR371b5p inhibits endothelial cell apoptosis in monocrotaline-induced pulmonary arterial hypertension via PTEN/PI3K/Akt signaling pathways. *Mol Med Rep* 2018; 18: 5489-5501.
- 7) MIAO C, CHANG J, ZHANG G. Recent research progress of microRNAs in hypertension pathogenesis, with a focus on the roles of miRNAs in pulmonary arterial hypertension. *Mol Biol Rep* 2018; 45: 2883-2896.
- 8) BARTEL DP. MicroRNAs: genomics, biogenesis, mechanism, and function. *Cell* 2004; 116: 281-297.
- 9) NAGAO Y, HISAOKA M, MATSUYAMA A, KANEMITSU S, HAMADA T, FUKUYAMA T, NAKANO R, UCHIYAMA A, KAWAMOTO M, YAMAGUCHI K, HASHIMOTO H. Association of microRNA-21 expression with its targets, PDCD4 and TIMP3, in pancreatic ductal adenocarcinoma. *Mod Pathol* 2012; 25: 112-121.
- 10) YANG W, WANG A, ZHAO C, LI Q, PAN Z, HAN Y, WANG C, WANG G, JI C, WANG G, JIA G, JU J, GAO W, LIU W, LIU X, CHEN X, FENG W, GAO Z, LI J, REN C. miR-195-497 enhances IL-8 production in early-onset severe preeclampsia by targeting Sphingosine-1-Phosphate Lyase 1. *PLoS One* 2015; 10: e016940.
- 11) CARLETON M, CLEARY MA, LIN J, WANG S. MicroRNAs and cell cycle regulation. *Cell Cycle* 2007; 6: 2127-2132.
- 12) GANTIER MP, SADLER AJ, VAN DER MADE I, VAN DEN HOOGENHOF MM, VAN DEN ENKEL WJ, VAN DEEL ED, DE GROOT NE, ALEKSEEV S, FLUITER K, SCHROEN B, GOUMANS MJ, VAN DER VELDEN J, DUNCKER DJ, PINTO YM, CREEMERS EE. The miR-15 family inhibits the TGFbeta1 signaling pathway in the heart. *Cardiovasc Res* 2014; 104: 670-671.
- 17) BAI YW, YE MJ, YANG DL, YU MF, LIU CF, SHEN T. Hydrogen sulfide attenuates angiotensin-induced epithelial-mesenchymal transition in human alveolar epithelial cells through regulating transforming growth factor-beta1/Smad2/3 signaling pathway. *J Appl Toxicol* 2018; 39: 432-440.
- 18) HARFE BD. MicroRNAs in vertebrate development. *Curr Opin Genet Dev* 2005; 15: 400-415.
- 19) NGUYEN BL, CAPOTE M, PERSI L, LACANICA A, RAFIGUE A, PICCIRILLO G, GAUDINO M, CES, SIEGEL M, VITARELLI A. Global and regional left ventricular strain indices in post-myocardial infarction patients with ventricular arrhythmias and moderately reduced left ventricular fractional shortening. *Int J Cardiol* 2013; 141: 407-417.
- 20) CARLSON MG, VAN DER WOUDE F, KOZIK TM, PELTER MM. Post-myocardial infarction arrhythmias. *Am J Crit Care* 2015; 24: 269-274.
- 21) DE-SILVA-CHAVES P, CERQUEIRA R, PINTALHAO M, LEITE-MOREIRA AF. New pathways of the renin-angiotensin system: the role of ACE2 in cardiovascular pathophysiology and therapy. *Expert Opin Ther Targets* 2010; 14: 485-496.
- 22) DOMASEK JJ, GAMBINI G, HINZ B, CHAPONNIER C, BROWN JH. Myofibroblasts and mechano-regulation of tissue remodelling. *Nat Rev Mol Cell Biol* 2002; 3: 349-363.
- 23) PORRELLO ER, JOHNSON BA, AURORA AB, SIMPSON E, NAMIKOVICH SJ, DORN GN, VAN ROOIJ E, OLSON EN. miR-15 family regulates postnatal mitotic arrest of cardiomyocytes. *Circ Res* 2011; 109: 670-679.
- 24) SATO T, YAMAMOTO T, SEHARA-FUJISAWA A. miR-195/497 induce postnatal quiescence of skeletal muscle stem cells. *Nat Commun* 2014; 5: 4597.
- 25) GABBIANI G. The myofibroblast in wound healing and fibrocontractive diseases. *J Pathol* 2003; 200: 500-503.
- 26) LIN L, LI R, CAI M, HUANG J, HUANG W, GUO Y, YANG L, YANG G, LAN T, ZHU K. Andrographolide ameliorates liver fibrosis in mice: involvement of TLR4/NF-kappaB and TGF-beta1/Smad2 signaling pathways. *Oxid Med Cell Longev* 2018; 2018: 7808656.
- 27) LIAO K, YONG CW, HUA K. SB431542 inhibited cigarette smoke extract induced invasiveness of A549 cells via the TGF-beta1/Smad2/MMP3 pathway. *Oncol Lett* 2018; 15: 9681-9686.
- 28) LAL H, AHMAD F, ZHOU J, YU JE, VAGNOZZI RJ, GUO Y, YU D, TSAI EJ, WOODGETT J, GAO E, FORCE T. Cardiac fibroblast glycogen synthase kinase-3beta regulates ventricular remodeling and dysfunction in ischemic heart. *Circulation* 2014; 130: 419-430.
- 29) MELCHIOR-BECKER A, DAI G, DING Z, SCHAFFER L, SCHRADER J, YOUNG MF, FISCHER JW. Deficiency of biglycan causes cardiac fibroblasts to differentiate into a myofibroblast phenotype. *J Biol Chem* 2011; 286: 17365-17375.