

# The burden of calcific aortic stenosis: what's behind?

S. ROMITI, M. VINCIGUERRA, M. D'ABRAMO, N. BRUNO, F. MIRALDI, C. GAUDIO, E. GRECO

Department of Clinical, Internal Medicine, Anaesthesiology and Cardiovascular Sciences, Sapienza University of Rome, Rome, Italy

**Abstract.** – In Western countries, calcific aortic valve stenosis (CAS) is widely common, representing the third cause of death among cardiovascular diseases (CVD). The burden of CAS is high, with an increasing prevalence rate related to age. An efficient medical treatment, according to guidelines, lacks to prevent the development and to reduce the progression of CAS. In this context, due to the aging population and the lack of effective medical management, the prevalence is expected to double-triple within the next decades. In our review, we aim to provide an overview of the underlying mechanisms of pathogenesis and the current state of the art regarding pathophysiological insights and novel potential therapeutic targets.

*Key Words:*

Calcific aortic stenosis, Atherosclerosis, Aortic valve interstitial cell, Lipid-lowering agents, Antiresorptive agents, Inflammation.

## Introduction

The global burden of calcific aortic valve disease (CAVD) is increasing, representing the most common valvular heart disease in the aging population. It was estimated that in 2017 more than 12 million people were affected, and it was responsible for more than 100,000 deaths<sup>1</sup>. The prevalence rate of calcific aortic stenosis (CAS) ranges from 1.7% in people over 65 years to more than 6% in people aged 85<sup>2-4</sup>, having an increasing impact both for the public health and healthcare resources<sup>4</sup>. CAS is a chronic progressive disease strictly age-related and characterized by fibro-calcific remodeling of aortic leaflets, which results in an obstruction of the left ventricular outflow, myocardial hypertrophy, and, once symptoms appear, risk of heart failure and sudden death. No medical treatments to prevent the development or to

reduce the progression of CAS are currently recommended by both American Heart Association/American College of Cardiology (AHA/ACC) and European Society of Cardiology and the European Association for Cardio-Thoracic Surgery (ESC/EACTS) guidelines<sup>5,6</sup>; the only option available for the treatment of severe and symptomatic CAS still remains aortic valve replacement, even with minimal invasive approach, or trans-catheter implantation<sup>7-11</sup>. Patients with severe CAS in the absence of surgical or percutaneous intervention have a poor prognosis with a high rate of re-hospitalization and risk of mortality<sup>4</sup>. The onset of symptoms changes dramatically the natural history of the disease, with a mortality rate of approximately 50% after two years without any intervention<sup>12</sup>. Risk factors such as age, male sex, smoking, hypertension, dyslipidemia, high levels of Lipoprotein (a) (Lp(a)), diabetes mellitus, and obesity are evidenced in CAS and atherosclerosis leading to consider them as a diversified expression of the same disease<sup>13-15</sup>. In addition, the bicuspid aortic valve (BAV), a congenital malformation of the aortic valve, with related abnormal hemodynamic stress and genetic factors, is a powerful risk factor<sup>16</sup>.

## Aortic Valve Anatomy and Features

To better understand the pathophysiological insights of CAS is essential to describe the anatomic and histological features of the aortic valve (AV). The AV is an avascular structure, which modulates the unidirectional blood flow from the left ventricle to the aorta, opening during the systole and closing during the diastolic phase of the cardiac cycle. The AV is normally composed of three cusps named according to their relationship with the coronary artery ostia (the left coronary, the

right, and the non-coronary cusp) and a fibrous annulus in continuity with the anterior leaflet of the mitral valve and the membranous septum. The AV consists of connective tissue stratified into three layers: the fibrosa, the spongiosa, and the ventricularis layer. The connective tissue is composed of extracellular matrix components (fibronectin, lamin, collagen, and elastin fibers, etc.) and predominantly two cellular populations: the aortic valve interstitial cells (AVICs) and the aortic valve endothelial cells (AVECs)<sup>17,18</sup>. The AVICs in healthy patients are described as “quiescent” sharing phenotypic similarities with fibroblast and can be activated by different stimuli in the osteogenic and myofibroblastic phenotype. The AVECs are located over the surface of the leaflets as a physical barrier. They have been shown to possess mechanosensitive properties and contribute to extracellular matrix homeostasis by communicating with the underlying AVICs<sup>18</sup>. Additionally, extra valvular cells may be involved in the homeostasis of the extracellular matrix, especially in the elderly, as suggested by experimental data based on animal models<sup>19</sup>.

### Pathophysiological Insights

CAS is a complex age-related multifactorial disease still incompletely understood. Predisposing risk factors such as age, male sex, smoking, hypertension, dyslipidemia, high levels Lp(a), diabetes mellitus, and obesity are determinants to increase susceptibility and speed up the progression<sup>20</sup>. Moreover, in younger patients, BAV represents the most common etiology of CAS<sup>21</sup>.

BAV is the commonest congenital heart disease, with a worldwide incidence ranging up to 2%. Its natural history is characterized by valvular (insufficiency, stenosis, endocarditis) and vascular complications (dilatation, aneurysm, dissection)<sup>22</sup>. In BAV disease AV forms two instead of three leaflets and, according to the number of raphe, can be classified into three types (Figure 1)<sup>23</sup>. It is genetically determined in most cases. The inheritance appears to be autosomal dominant with incomplete penetrance. However, some studies<sup>24</sup> suggested the existence of an X-linked form, as indicated by the high prevalence of this pathology in patients with Turner syndrome. Although a single gene defect has not yet been identified, NOTCH1, GATA gene mutations, and endothelial nitric oxide synthase abnormalities were found<sup>22</sup>. Genetic factors in association with

abnormal shear stress in these patients lead to earlier leaflets degeneration and consequent earlier need for surgery compared to tricuspid aortic valve patients<sup>16,22</sup>.

It is, therefore, evident that CAS and vascular atherosclerosis share the same risk factors and, consequently, the early molecular pathogenic mechanisms. Different research groups have already investigated the association between atherosclerosis and CAS, finding that about 50% of patients with CAS have a concomitant coronary artery disease or vascular disease<sup>25,26</sup>. Both are chronic inflammatory diseases, whereas common risk factors activate and promote a self-maintenance inflammatory state leading respectively to the formation and progression of the valvular fibro-calcific remodeling and the atheromatous plaques<sup>13</sup>. As in atherosclerosis, mechanical stress is a key determinant of endothelial damage, leading to the infiltration and accumulation of lipids beneath the endothelium of the valve. Progressive endothelial injury and lipid oxidization activate the inflammatory response and exacerbate oxidative stress. This pro-inflammatory state promotes the activation of AVICs in CAS and the phenotypic switch of vascular smooth muscle cells in atherosclerosis, leading to extracellular matrix remodelling<sup>27</sup>. However, discrepancies exist, as recently reported by Lee et al<sup>28</sup> who showed that AV calcification progression was associated only with the progression of calcified atherosclerotic plaque but not with non-calcified plaque. Additionally, ultrastructural differences were found between leaflet calcification and atherosclerotic plaque showing in atherosclerosis the unique massive accumulation of lipids and the pronounced neovascularization<sup>27</sup>.

For many years the pathogenesis of CAS was considered only a passive process characterized by dystrophic calcification and remodeling of valve leaflets due to cells aging and death with consecutive calcification of their degradation products<sup>29</sup>. New evidence suggests that CAS is a complex active process still incompletely understood in which several pathways drive the progression of the disease<sup>15,30</sup>. Based on this thesis, two different phases of pathogenesis can be identified: the initiation and the propagation phase (Figure 2). In the initiation phase, CAS pathogenesis is like atherosclerosis: mechanical/shear stress is at the basis of endothelial damage, allowing the infiltration and accumulation of lipids in the endothelium of the valve. The oxidation of lipoproteins activates the inflammatory response, exacerbating oxidative stress, which promotes the

phenotypic change in AVICs from a quiescent to an osteogenic and myofibroblastic one<sup>14</sup>.

The AVICs activation starts the propagation phase leading to leaflets calcification and fibro remodeling due to activated and self-maintained calcific and inflammatory signaling pathways<sup>15</sup>.

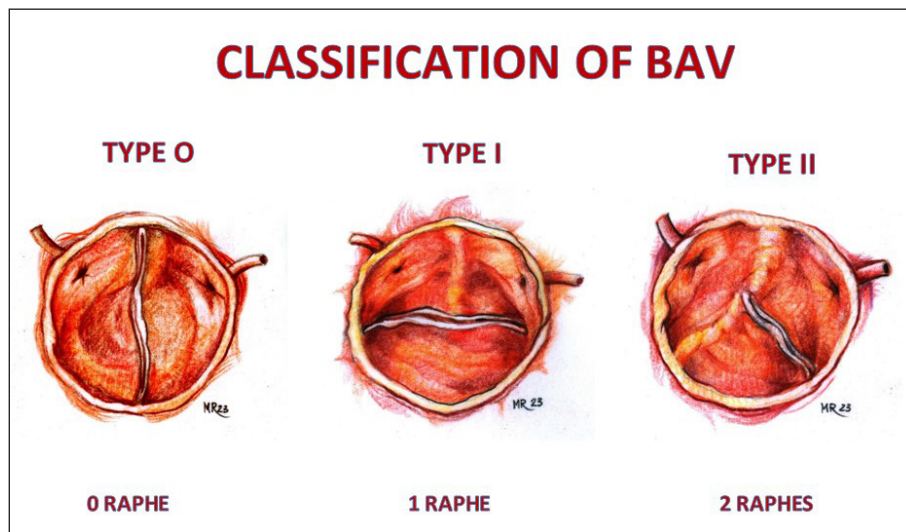
### The Initiation Phase

Endothelial damage represents the *primum movens* of CAS pathogenesis, allowing cell infiltration and lipid accumulation through mechanosensitive signaling pathways. Dysfunctional endothelium by altered paracrine signaling leads to up-regulation of cell adhesion molecules such as intercellular adhesion molecule 1 (ICAM-1) and vascular cell adhesion molecule 1 (V-CAM-1), which promote the invasion of macrophages and T cells in the valvular fibrosa. The immune cells infiltration results in pro-fibrotic and pro-inflammatory markers activation, such as extracellular protease, cytokines, and growth factors<sup>27,31</sup>. Interestingly macrophages demonstrated a crucial role in extracellular matrix remodeling and degradation<sup>13</sup>. Histopathologic studies<sup>32,33</sup> on early calcific valve lesions showed lipoproteins deposition and diffuse distribution of T cells both in the bicuspid and tricuspid aortic valves. On the contrary, on normal aortic valves, the absence of these cells has been reported. Moreover, neo-vascularization and inflammatory infiltrates are reported as histological features of the CAS valve, in addition to intra-leaflet hemorrhages and iron accumulation<sup>27,34-36</sup>. Laguna-Fernandez et al<sup>36</sup> found that the iron accumulated can be uptake by AVICs and potentially contributes to their activation and extracellular matrix remodeling and calcification. Emerging evidence highlighted the involvement of the NF- $\kappa$ B pathway through toll-like receptors and NOD-like receptor signaling pathways<sup>37,38</sup>. The immune system activation leads to the production and release of several cytokines. Interestingly Urban et al<sup>39</sup> found in their work that patients with CAS had higher levels of pro-inflammatory cytokines than both controls and patients with aortic regurgitation. Transforming growth factor  $\beta$ 1 (TGF- $\beta$ 1), interleukin- $1\beta$  (IL- $1\beta$ ), and tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) are reported as the predominant inflammatory cytokines involved in vascular calcification<sup>40</sup>. They contribute to a self-maintained inflammatory state by increasing the local production of matrix metalloproteinases, modulating apoptosis, cell proliferation, mi-

gration, and differentiation. Additionally, they can promote the endothelial-mesenchymal transition of AVECs into AVICs and, subsequently, their activation in the osteogenic phenotype<sup>41</sup>. AVICs (myo)fibroblast phenotype differentiation can be stimulated directly by TGF- $\beta$ 1 and leads to extracellular matrix rearrangement, collagen deposition, etc<sup>42</sup>. Interestingly, Chakrabarti et al<sup>43</sup>, in a study on animal model, found that the inhibition of the TGF $\beta$ 1-dependent SMAD3 signaling pathway reduces significantly the AV calcification. Inflammatory cytokines from different stimuli increase oxidative stress, which promotes the formation of oxidized low-density lipoproteins (Ox-LDLs) and phospholipids (Ox-PL). Oxidized lipids trigger and add further stimuli to endothelial dysfunction and inflammation by up-regulating cell adhesion molecules and inducing the activation of the Toll-like receptors and NF- $\kappa$ B (nuclear factor  $\kappa$ B) pathway<sup>44</sup>. In addition to inflammatory stimuli, biomechanical stress, and valve injury can activate an autocrine signal which leads to AVICs activation and differentiation into the osteoblast-like and myofibroblast-like phenotype<sup>45</sup>. In a recent pre-clinical study, Rogers et al<sup>46</sup> found that the retinoid acid may be involved in AVICs osteogenic differentiation and potentially represent a novel therapeutic target.

### The Propagation Phase

When the inflammatory state is established, and the AVICs are activated from prolonged different stimuli, CAS disease progresses into the propagation phase. The AVICs switch phenotype into myofibroblast- and osteoblast-like cells may represent the critical step in the pathogenesis and progression of CAS, leading to valve calcification and remodeling. At the beginning of pathogenesis, the inflammatory pathways seem to drive the progression of the disease. On the contrary, in the later stages, the calcific pathways such as the RANK (receptor activator of nuclear factor kappa B)/RANKL (RANK ligand)/OPG (osteoprotegerin), the Wnt (wingless and Int-1)/ $\beta$ -catenin, and the NOTCH signaling pathways seem to become predominant<sup>44</sup>. The NOTCH1 signaling pathway is involved in the repression of Runx2 a transcriptional regulator of osteoblast cell fate. Inactivating NOTCH1 mutations was reported as a predisposing risk factor and faster progression for CAS and also may contribute to the develop-



**Figure 1.** Representation of BAV morphologies, according to Sievers classification<sup>23</sup>. BAV type 0 or “true” BAV is characterized by the absence of raphe; on the contrary, type I and type II are characterized by the presence, respectively, of one and two raphes. BAV: bicuspid aortic valve.

ment of BAV<sup>47,48</sup>. Moreover, NOTCH1 appears to be involved in the activation of bone morphogenetic protein (BMP)-2, essential for osteoblastic differentiation<sup>49</sup>. The RANK/RANKL/OPG pathway can promote AVICs switch phenotype and subsequently matrix calcification and calcific nodules deposition. Not only the differentiation but also the apoptosis of AVICs can enhance and perpetuate the disease through dystrophic calcification<sup>30</sup>.

### Dyslipidemia and Lipid-Lowering Therapy

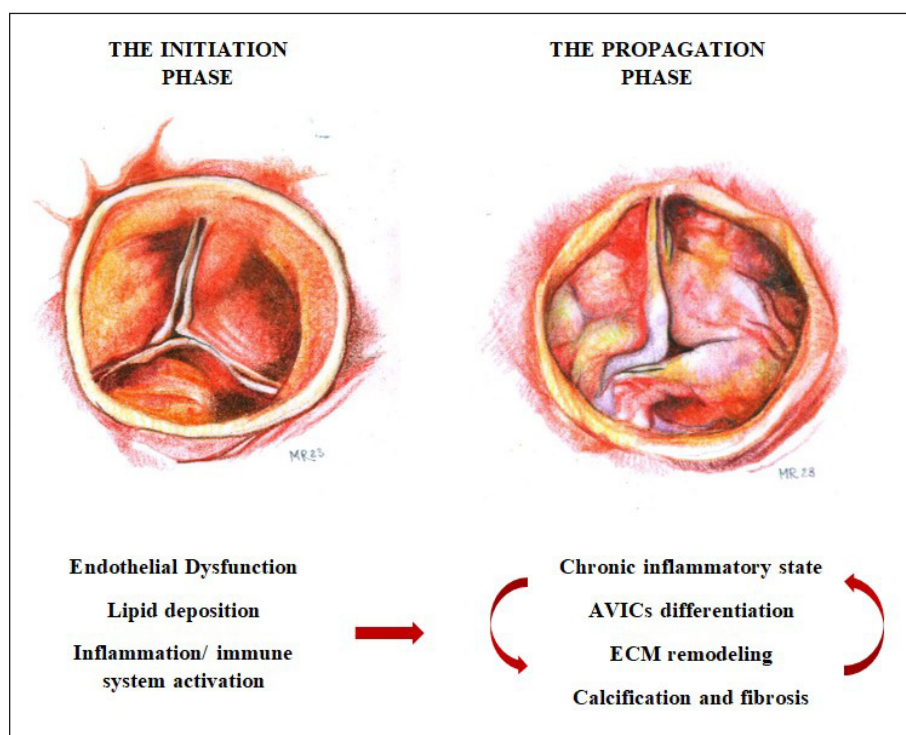
The pivotal role of dyslipidemia in triggering and promoting AV calcification is well recognized by several studies<sup>50,51</sup>. In patients with homozygous familial hypercholesterolemia is described an increased incidence and progression of valve degeneration. Elevated low-density lipoproteins (LDL) and reduced high-density lipoproteins (HDL) levels were associated with a higher risk of incidence of CAS<sup>14,52</sup>. Several immunohistochemical studies<sup>33,52,53</sup> demonstrated the presence of various apolipoproteins, including apoE, apoA1, apo(a), and ApoB in association with high levels of oxidized phospholipid (OxPL)<sup>33</sup>, which concentration directly correlated with the degree of inflammation and fibro-calcific remodeling<sup>52,53</sup>. Visceral obesity and metabolic syndrome are important risk factors for CAS leading to an inflam-

matory state and enhancing oxidative stress<sup>54,55</sup>. Interestingly, these conditions increased the risk of CAS not only due to hypercholesterolemia and the consequent increase in LDL levels but also to the reduction in adiponectin and HDL serum levels<sup>20,55</sup>. In particular, adiponectin is a peptide hormone with anti-inflammatory and anti-atheromatous properties, produced by the adipocyte, which is greatly reduced in obese patients (BMI  $\geq 30$ ) and in patients with metabolic syndrome<sup>56</sup>. Serum levels of adiponectin have been recognized as a risk factor for atherosclerosis and a potential novel therapeutic target for CAS and its progression<sup>57,58</sup>.

The potential of Statin therapy in CAS disease was largely described in literature<sup>14,59-62</sup>. Statins are competitive inhibitors of HMG-Coa (3-hydroxy-3methylglutaryl-coenzyme) reductase, a key enzyme in sterol biosynthesis, and are a milestone of cardiovascular disease prevention. They have been shown to possess notable pleiotropic effects. In addition to lowering lipid levels, statins reduce the expression of inflammatory cytokines decreasing oxidative stress and inflammation as well as improving endothelial function<sup>63</sup>. Despite the strong correlation of CAS with altered lipid metabolism, chronic inflammation, and essential factors of atherosclerosis, none of the randomized clinical trials showed significant benefits of statin therapy<sup>64-66</sup> regarding both clinical presentation and CAS disease progression, with the exception of a non-randomized study<sup>67</sup>.

Nevertheless, a limitation of all these studies<sup>63-67</sup> may be the introduction of statin therapy in the late phases of the disease when fibrosis and structural changes of the leaflets are already in an advanced step and when the pro-inflammatory state was established and AVICs activated in a self-feeding circuit. Differences age-related in patients with CAS were shown by Owens et al<sup>68</sup>; they found that elevated LDL was a risk factor for CAS only in participants younger than 65 years. In addition, results from the PROGRESSA study<sup>69</sup> and the EPIC-Norfolk prospective study<sup>70</sup> evidenced that, especially in younger patients, there was a significant association between high apoB/apoA-I ratio (ApoB is the main component of LDL and ApoA-I of HDL) and the hemodynamic progression rate of CAS; conversely, in elderly patients, this association was less evident, probably due to the predominant role of age-related factors such as osteoporosis, disorders of the calcium-phosphorus metabolism and the other side statins effects such as osteogenic properties, increased levels of Lp(a) and increased resistance to insulin. Likewise, dyslipidemia may have a pivotal role in the early stage (initiation phase) and has a minor impact when the activation of AVICs and the pro-inflammatory state are established

(propagation phase). Lp(a) is a lipoprotein composed of apoB100 of LDL covalently attached to apo(a). It is genetically determined and represents a random and independent risk factor for CAS<sup>70,71</sup> and atherosclerotic cardiovascular disease<sup>72,73</sup>. As reported in recent reviews<sup>73,74</sup>, elevated Lp(a), as a major carrier of OXlp, promote a pro-inflammatory state by stimulating activation of monocytes and macrophages, pro-inflammatory cytokines (e.g., IL1 $\beta$ , IL-6, IL-8, and TNF- $\alpha$ ) and mediators both involved in the development and progression of CAS and atherosclerosis. Elevated Lp(a) levels and the corresponding associated genotypes (rs10455872, rs3798220, kringle IV type 2 repeat polymorphism) were also correlated with an increased risk of aortic stenosis in the general population and a tripled risk for Lp(a) > 90 mg/dl<sup>75</sup>. Due to its potential as a novel therapeutic target, great interest emerged in the scientific community regarding agents that can potentially act to decrease Lp(a) levels. Lp(a) levels are not significantly modified by statin treatment, but the use of new therapeutic agents such as Proprotein Convertase Subtilisin/Kexin Type 9 (PCSK9) inhibitors and antisense oligonucleotides (ASOs) gain a huge interest (**Supplementary Table I**). Emerging evidence showed that patients with a loss-of-



**Figure 2.** Schematic representation of CAS pathogenesis and its related two phases: the initiation and the propagation phase. CAS: calcific aortic stenosis, ECM: extracellular matrix components.

function mutation of PCSK9 have reduced levels of Lp(a), LDL cholesterol and lower risk of CAS and cardiovascular diseases<sup>76,77</sup>. Indeed this protein reduces the hepatocyte receptors that remove LDL cholesterol (LDL-C) from the blood; its inhibition leads to a reduction in the degradation of the receptor with a consequent reduction in blood concentrations of LDL-C<sup>78</sup>. Newsworthy, the use of monoclonal antibodies against PCSK9, Evolocumab, and Alirocumab, seems to significantly reduce Lp(a) values and potentially the incidence of cardiovascular disease and adverse events<sup>72,79,80</sup>. Promising results from clinical trials<sup>81,82</sup> showed the effectiveness of PCSK9 inhibitors as lipid-lowering therapy, especially in association with statin. The FOURIER (Further Cardiovascular Outcomes Research With PCSK9 Inhibition in Patients With Elevated Risk) trial<sup>83</sup> evaluated the effectiveness of Evolocumab, a monoclonal antibody against PCSK9, compared with placebo in patients with dyslipidemia who were receiving statin therapy to reduce the cardiovascular death, myocardial infarction, and stroke (ClinicalTrials.gov Identifier: NCT01764633). A significant reduction in all cardiovascular events was found. The combination of Evolocumab with statin therapy lowered the LDL-C levels by 60% compared to statin therapy alone. A secondary analysis of the FOURIER trial highlighted the potential of Evolocumab to reduce the risk of CAS progression and its adverse events<sup>84</sup>. In the ODYSSEY Outcomes trial<sup>82</sup> (Evaluation of Cardiovascular Outcomes After an Acute Coronary Syndrome During Treatment With Alirocumab) in patients with previous acute coronary syndrome, the combination of Alirocumab with statin therapy significantly reduce the risk of cardiovascular events (ClinicalTrials.gov Identifier: NCT01663402). An interesting ongoing clinical trial<sup>85</sup> is investigating the effectiveness of monoclonal antibodies against PCSK9 in association with statins in preventing and delaying the progression of CAS (ClinicalTrials.gov Identifier: NCT04968509). Emerging evidence highlights the potential of synthetic ASOs as a pharmaceutical intervention to directly decrease Lp(a) levels in patients with cardiovascular disease or CAS<sup>86,87</sup>. Indeed the antisense oligonucleotides IONIS-APO(a)Rx and the IONIS-APO(a)LRx, completed, respectively, the trial phase 2 and phase 1<sup>86</sup>. Randomized, double-blind, placebo-controlled trials (ClinicalTrials.gov Identifier: NCT02160899; ClinicalTrials.gov Identifier: NCT02414594, ClinicalTrials.gov Identifier: NCT03070782; EudraCT Number:

2012-004909-27)<sup>86-89</sup> were conducted to evaluate their efficacy and safety to lower Lp(a) levels. Both the two ASOs resulted tolerable and highly effective to reduce Lp(a) concentrations<sup>88,89</sup> (Supplementary Table I). Additionally, they seem to reduce the pro-inflammatory activation of circulating monocytes in patients with elevated Lp(a)<sup>90</sup>. Promising findings resulted from a clinical trial<sup>91</sup> investigating the effectiveness of Mipomersen, an ASOs inhibitor of apo(b) synthesis, in the management of patients at higher cardiovascular risk with severe hypercholesterolemia. Interestingly, Thomas et al<sup>91</sup> found that Mipomersen as an add-on therapy significantly modified LDL-C and lipoproteins levels (Supplementary Table I).

Newsworthy, some studies<sup>92</sup> suggest that Autotaxin, an enzyme involved in the production of extracellular lysophosphatidic acid, may promote inflammation and osteogenic transition in the AVICs resulting in a potential novel biomarker of CAS progression.

### **Dysregulated Mineral Metabolism and Antiresorptive Agents**

Recent evidence<sup>93-95</sup> suggests the association between CAS and dysregulated mineral metabolism and/or osteoclast deficiency, although the underline mechanisms remain still unclear. A correlation has been observed between the incidence of CAS and disorders of bone turnover, such as low bone mineral density, as well as chronic kidney disease and Paget's disease<sup>93,94</sup>. In the pathophysiology of CAS, a critical step is represented by the AVICs differentiation in the osteogenic phenotype leading to increase expression of osteoblast-specific proteins such as bone sialoprotein and osteopontin<sup>30</sup>. Calcific signaling pathways seem to have a dominant role in the later phases of the disease, when the differentiation of AVICs drives the disease progression. The AVICs activation enables positive feedback in which calcium deposition on leaflets increases mechanical stress and consequently injury-induced activation of the Wnt/b-catenin pathway, with further osteoblast differentiation<sup>30,95</sup>. Interestingly it has been reported that the factors, such as inflammatory cytokines and modified lipoproteins, that in skeletal bone cells induce bone resorption in vascular mineralization appear to have the opposite effect<sup>30,96</sup>. Therefore, over the last few years, there has been growing interest in studying the potential of antiresorptive agents (e.g., denosumab and alendronate, etc.) on AV calcification (Supplementary

Table I). Denosumab is a human monoclonal antibody RANKL inhibitor used for the medical treatment of osteoporosis<sup>97</sup>. *In vitro* and observational studies<sup>98,99</sup> have demonstrated the potential of Denosumab as an inhibitor of AVICs to delay CAS progression. Additionally, Alendronate, a bisphosphonate that inhibits bone resorption by suppressing the activity of osteoclast, appears to slow down the progression of CAS, especially in patients with concomitant osteoporosis<sup>100</sup>. Despite the promising findings of *in vitro* and observational studies<sup>98,99</sup>, in a recent double-blind, randomized controlled trial<sup>101</sup>, neither Denosumab nor Alendronate affected the progression of valve calcification in patients with CAS (Supplementary Table I).

### Novel Therapeutic Targets

Both innate and adaptive immunities seem to be independently related to the leaflet remodeling process and the CAS progression. *In vitro* studies<sup>102,103</sup> have investigated the potential of anti-inflammatory agents to suppress AVICs activation and calcium deposition, such as IL-38 and IL-37, underlying the potential of anti-inflammatory therapies in the treatment and prevention of CAS as a chronic inflammatory disease. Interestingly, two randomized clinical trials are evaluating the effectiveness of colchicine in CAS progression (ClinicalTrials.gov Identifier: NCT05162742; EudraCT Number: 2021-005586-40)<sup>104,105</sup> (Supplementary Table I). Additionally, some authors proposed natural antioxidant agents as a potential novel therapeutic option. In the field of vascular calcification, vitamin K2 as an inhibitor of arterial calcification has been suggested to potentially slow valve calcification, showing significant changes in observational studies in the levels of calcification agents, although no effect has been proved in elderly men treated with vitamin K2 and vitamin D supplementation<sup>106</sup>. Currently, another ongoing trial<sup>107</sup> is evaluating its efficacy on AV calcification (Supplementary Table I). News-worthy, pre-clinical studies<sup>108</sup> showed that non-vitamin K antagonist oral anticoagulants (NOACs) inhibit AVICs activation and subsequently may potentially reduce aortic valve calcification.

### Conclusions

CAS is a complex multifactorial disease where different molecular agents are involved and dom-

inate the progression of the disease according to the stage of pathogenesis. In the initiation phase, like atherosclerosis, endothelial injury, dyslipidemia, and inflammation trigger the activation of the AVICs in the osteogenic and fibroblastic phenotype. In the propagation phase, when the inflammatory state is established, and the AVICs are activated, a self-feeding mechanism is triggered: the AVICs activated induce calcific deposition and remodeling of the matrix through the activation of calcium pathways and the immune system, on the other hand, the immune system adds further stimulus both directly and indirectly by stimulating the activity of AVICs and enabling the rearrangement of the extracellular matrix components.

The complexity of the molecular mechanisms underlying the onset and progression of valve remodeling reveals the difficulty of identifying a therapeutic target that is effective in the various stages of the disease and, therefore, an effective medical treatment. A different therapeutic management for the prevention and treatment of CAS may be considered according to the different stages of the disease. In support of this thesis, the promising results of retrospective studies in delaying the onset and progression of CAS in patients treated for long-term with medical therapy recommended for atherosclerosis (statin therapy, etc.) didn't show effectiveness when introduced in the late phases of the disease, when the propagation phase speed out the progression and a self-maintained vicious circle was established.

To date, CAS still remains a complex, still not fully understood disease, as demonstrated by the controversial results of potential medical therapies in animal model studies and clinical trials.

Further methodological studies are needed to extend our knowledge on the pathogenesis of age-related CAS and on the effectiveness of the treatments used to prevent and control this pathology; maybe a tailored therapy characterized by drugs association could be suggested since several different pathways have been shown to be implicated in the progression of valve degeneration.

---

### Conflict of Interest

The Authors declare that they have no conflict of interests.

---

### Acknowledgments

We thank Marzio Romiti for drawing the figures.

### Funding

The authors received no financial support for the research, authorship, and/or publication of this article.

### Authors' Contributions

All authors contributed significantly to this work and read and approved the final version of the manuscript. Ernesto Greco performed the conception and design of the study and revised it critically, Silvia Romiti performed the design of the study and wrote the manuscript, Mattia Vinciguerra, Mizar D'Abramo, Noemi Bruno, Carlo Gaudio and Fabio Miraldi revised it critically for important intellectual content.

### ORCID ID

Silvia Romiti: 0000-0002-9594-4033  
Mattia Vinciguerra: 0000-0001-5237-7342  
Mizar D'Abramo: 0000-0002-7310-7410  
Noemi Bruno: 0000-0003-4847-9885  
Fabio Miraldi: 0000-0002-2404-3369  
Ernesto Greco: 0000-0003-3177-5303.

### Availability of Data and Materials

All data generated or analyzed during this study are included in this published article [and/or its supplementary material].

### Informed Consent

Informed consent is not applicable to this study.

### Ethics Approval

Ethics approval is not applicable to this study.

## References

- 1) Yadgir S, Johnson CO, Aboyans V, Adebayo OM, Adedoyin RA, Afarideh M, Alahdab F, Alashi A, Ali-pour V, Arabloo J, Azari S, Barthelemy CM, Benzinger CP, Berman AE, Bijani A, Carrero JJ, Carvalho F, Daryani A, Durães AR, Esteghamati A, Farid TA, Farzadfar F, Fernandes E, Filip I, Gad MM, Hamidi S, Hay SI, Ilesanmi OS, Naghibi Irvani SS, Jürisson M, Kasaeian A, Kengne AP, Khan AR, Kisa A, Kisa S, Kolte D, Manafi N, Manafi A, Mensah GA, Mirrakhimov EM, Mohammad Y, Mokdad AH, Negoji RI, Thi Nguyen HL, Nguyen TH, Nixon MR, Otto CM, Patel S, Pilgrim T, Radfar A, Rawaf DL, Rawaf S, Rawasia WF, Rezapour A, Roever L, Saad AM, Saadatagah S, Senthilkumaran S, Sliwa K, Tesfay BE, Tran BX, Ullah I, Vaduganathan M, Vasankari TJ, Wolfe CDA, Yonemoto N, Roth GA. Global, Regional, and National Burden of Calcific Aortic Valve and Degenerative Mitral Valve Diseases, 1990-2017. *Circulation* 2020; 141: 1670-1680.
- 2) Osnabrugge RL, Mylotte D, Head SJ, Van Mieghem NM, Nkomo VT, LeReun CM, Bogers AJ, Piazza N, Kappetein AP. Aortic stenosis in the elderly: Disease prevalence and number of candidates for transcatheter aortic valve replacement: A meta-analysis and modeling study. *J Am Coll Cardiol* 2013; 62: 1002-1012.
- 3) Kanwar A, Thaden JJ, Nkomo VT. Management of Patients With Aortic Valve Stenosis. *Mayo Clin Proc* 2018; 93: 488-508.
- 4) Clark MA, Arnold SV, Duhay FG, Thompson AK, Keyes MJ, Svensson LG, Bonow RO, Stockwell BT, Cohen DJ. Five-year clinical and economic outcomes among patients with medically managed severe aortic stenosis: Results from a medicare claims analysis. *Circ Cardiovasc Qual Outcomes* 2012; 5: 697-704.
- 5) Nishimura RA, Otto CM, Bonow RO, Carabello BA, Erwin JP 3rd, Fleisher LA, Jneid H, Mack MJ, McLeod CJ, O'Gara PT, Rigolin VH, Sundt TM 3rd, Thompson A. 2017 AHA/ACC Focused Update of the 2014 AHA/ACC Guideline for the Management of Patients With Valvular Heart Disease: A Report of the American College of Cardiology/American Heart Association Task Force on Clinical Practice Guidelines. *J Am Coll Cardiol* 2017; 70: e1159-e1195.
- 6) Vahanian A, Beyersdorf F, Praz F, Milojevic M, Baldus S, Bauersachs J, Capodanno D, Conradi L, De Bonis M, De Paulis R, Delgado V, Freemantle N, Gilard M, Haugaa KH, Jeppsson A, Jüni P, Pierrard L, Prendergast BD, Sádaba JR, Tribouilloy C, Wojakowski W; ESC/EACTS Scientific Document Group. 2021 ESC/EACTS Guidelines for the management of valvular heart disease. *Eur Heart J* 2022; 43: 561-632.
- 7) Greco E, Zaballos JM, Alvarez L, Urso S, Pulitani I, Sádaba R, Juaristi A, Goiti JJ. Video-assisted mitral surgery through a micro-access: A safe and reliable reality in the current era. *J Heart Valve Dis* 2008; 17: 48-53.
- 8) Greco E, Barriuso C, Castro MA, Fita G, Pomar JL. Port-access™ cardiac surgery: From a learning process to the standard. *Heart Surg Forum* 2002; 5: 145-149.
- 9) Urso S, Sadaba R, Greco E, Pulitani I, Alvarez L, Juaristi A, Goiti JJ. One-hundred aortic valve replacements in octogenarians: Outcomes and risk factors for early mortality. *J Heart Valve Dis* 2007; 16: 139-144.
- 10) Chirichilli I, D'Ascoli R, Rose D, Frati G, Greco E. Port Access (Thru-Port System) video-assisted mitral valve surgery. *J Thorac Dis* 2013; 5: S680-S685.
- 11) Marullo AG, Irace FG, Vitulli P, Peruzzi M, Rose D, D'Ascoli R, Iaccarino A, Pisani A, De Carlo C, Mazzesi G, Barretta A, Greco E. Recent Develop-



- ments in Minimally Invasive Cardiac Surgery: Evolution or Revolution? *Biomed Res Int* 2015; 2015: 483025.
- 12) Nishimura RA, Otto CM, Bonow RO, Carabello BA, Erwin JP 3rd, Guyton RA, O'Gara PT, Ruiz CE, Skubas NJ, Sorajja P, Sundt TM 3rd, Thomas JD; American College of Cardiology/American Heart Association Task Force on Practice Guidelines. 2014 AHA/ACC guideline for the management of patients with valvular heart disease: A report of the American college of cardiology/American heart association task force on practice guidelines. *J Am Coll Cardiol* 2014; 63: e71-e81.
  - 13) Lindman BR, Clavel MA, Mathieu P, lung B, Lancellotti P, Otto CM, Pibarot P. Calcific aortic stenosis. *Nat Rev Dis Prim* 2016; 2: 16006.
  - 14) Parisi V, Leosco D, Ferro G, Bevilacqua A, Pagano G, de Lucia C, Perrone Filardi P, Caruso A, Rengo G, Ferrara N. The lipid theory in the pathogenesis of calcific aortic stenosis. *Nutr Metab Cardiovasc Dis* 2015; 25: 519-525.
  - 15) Dweck MR, Khaw HJ, Sng GK, Luo EL, Baird A, Williams MC, Makiello P, Mirsadraee S, Joshi NV, van Beek EJ, Boon NA, Rudd JH, Newby DE. Aortic stenosis, atherosclerosis, and skeletal bone: Is there a common link with calcification and inflammation? *Eur Heart J* 2013; 34: 1567-1574.
  - 16) Balistreri CR, Forte M, Greco E, Paneni F, Cavarretta E, Frati G, Sciarretta S. An overview of the molecular mechanisms underlying development and progression of bicuspid aortic valve disease. *J Mol Cell Cardiol* 2019; 132: 146-153.
  - 17) Towler DA. Molecular and cellular aspects of calcific aortic valve disease. *Circ Res* 2013; 113: 198-208.
  - 18) Dutta P, James JF, Kazik H, Lincoln J. Genetic and Developmental Contributors to Aortic Stenosis. *Circulation Research* 2021; 128: 1330-1343.
  - 19) Anstine LJ, Horne TE, Horwitz EM, Lincoln J. Contribution of extra-cardiac cells in murine heart valves is age-dependent. *J Am Heart Assoc* 2017; 6: e007097.
  - 20) Kleinauskienė R, Jonkaitienė R. Degenerative aortic stenosis, dyslipidemia and possibilities of medical treatment. *Med* 2018; 54: 24-42.
  - 21) Ward C. Clinical significance of the bicuspid aortic valve. *Heart* 2000; 83: 81-85.
  - 22) Rajamannan NM. Bicuspid aortic valve disease: The role of oxidative stress in Lrp5 bone formation. *Cardiovasc Pathol* 2011; 20: 168-176.
  - 23) Sievers HH, Schmidtke C. A classification system for the bicuspid aortic valve from 304 surgical specimens. *J Thorac Cardiovasc Surg* 2007; 133: 1226-1233.
  - 24) Klásková E, Zapletalová J, Kaprálová S, Šnajderová M, Lebl J, Tüdös Z, Pavlíček J, Černá J, Mihál V, Stará V, Procházka M. Increased prevalence of bicuspid aortic valve in Turner syndrome links with karyotype: The crucial importance of detailed cardiovascular screening. *J Pediatr Endocrinol Metab* 2017; 30: 319-325.
  - 25) Boudoulas KD, Wolfe B, Ravi Y, Lilly S, Nagaraja HN, Sai-Sudhakar CB. The aortic stenosis complex: Aortic valve, atherosclerosis, aortopathy. *J Cardiol* 2015; 65: 377-382.
  - 26) Steinvil A, Leshem-Rubinow E, Abramowitz Y, Shacham Y, Arbel Y, Banai S, Bornstein NM, Finckelstein A, Halkin A. Prevalence and predictors of carotid artery stenosis in patients with severe aortic stenosis undergoing transcatheter aortic valve implantation. *Catheter Cardiovasc Interv* 2014; 84: 1007-1012.
  - 27) Kostyunin A, Mukhamadiyarov R, Glushkova T, Bogdanov L, Shishkova D, Osyayev N, Ovcharenko E, Kutikhin A. Ultrastructural pathology of atherosclerosis, calcific aortic valve disease, and bioprosthetic heart valve degeneration: Commonalities and differences. *Int J Mol Sci* 2020; 21: 7434.
  - 28) Lee SE, Sung JM, Andreini D, Al-Mallah MH, Budoff MJ, Cademartiri F, Chinnaiyan K, Choi JH, Chun EJ, Conte E, Gottlieb I, Hadamitzky M, Kim YJ, Lee BK, Leipsic JA, Maffei E, Marques H, de Araújo Gonçalves P, Pontone G, Shin S, Stone PH, Samady H, Virmani R, Narula J, Berman DS, Shaw LJ, Bax JJ, Lin FY, Min JK, Chang HJ. Association between Aortic Valve Calcification Progression and Coronary Atherosclerotic Plaque Volume Progression in the PARADIGM Registry. *Radiology* 2021; 300: 79-86.
  - 29) Mohler ER. Are atherosclerotic processes involved in aortic-valve calcification? *Lancet* 2000; 356: 524-525.
  - 30) Pawade TA, Newby DE, Dweck MR. Calcification in aortic stenosis: The skeleton key. *J Am Coll Cardiol* 2015; 66: 561-77.
  - 31) Lee SH, Choi JH. Involvement of immune cell network in aortic valve stenosis: Communication between valvular interstitial cells and immune cells. *Immune Netw* 2016; 16: 26-32.
  - 32) Wallby L, Janerot-Sjöberg B, Steffensen T, Broqvist M. T lymphocyte infiltration in non-rheumatic aortic stenosis: A comparative descriptive study between tricuspid and bicuspid aortic valves. *Heart* 2002; 88: 348-351.
  - 33) O'Brien KD, Reichenbach DD, Marcovina SM, Kuusisto J, Alpers CE, Otto CM. Apolipoproteins B<sub>100</sub>, (a)<sub>1</sub>, and E accumulate in the morphologically early lesion of 'degenerative' valvular aortic stenosis. *Arterioscler Thromb Vasc Biol* 1996; 16: 523-532.
  - 34) Chalajour F, Treede H, Ebrahimnejad A, Lauke H, Reichenspurner H, Ergun S. Angiogenic activation of valvular endothelial cells in aortic valve stenosis. *Exp Cell Res* 2004; 298: 455-464.
  - 35) Moreno PR, Astudillo L, Elmariah S, Purushothaman KR, Purushothaman M, Lento PA, Sharma SK, Fuster V, Adams DH. Increased macrophage infiltration and neovascularization in congenital bicuspid aortic valve stenosis. *J Thorac Cardiovasc Surg* 2011; 142: 895-901.
  - 36) Laguna-Fernandez A, Carracedo M, Jeanson G, Nagy E, Eriksson P, Caligiuri G, Franco-Cereceda

- A, Bäck M. Iron alters valvular interstitial cell function and is associated with calcification in aortic stenosis. *Eur Heart J* 2016; 37: 3532-3535.
- 37) Conte M, Petraglia L, Campana P, Gerundo G, Caruso A, Grimaldi MG, Russo V, Attena E, Leosco D, Parisi V. The role of inflammation and metabolic risk factors in the pathogenesis of calcific aortic valve stenosis. *Aging Clin Exp Res* 2021; 33: 1765-1770.
  - 38) Zeng Q, Song R, Ao L, Xu D, Venardos N, Fullerton DA, Meng X. Augmented osteogenic responses in human aortic valve cells exposed to oxLDL and TLR4 agonist: A mechanistic role of Notch1 and NF- $\kappa$ B interaction. *PLoS One* 2014; 9: e95400.
  - 39) Urban P, Rabajdová M, Špaková I, Sabol F, Mičková H, Lakatosová K, Zavacká M. Molecular recognition of aortic valve stenosis and regurgitation. *Eur Rev Med Pharmacol Sci* 2019; 23: 10996-11003.
  - 40) Yi YS. Role of inflammasomes in inflammatory autoimmune rheumatic diseases. *Korean J Physiol Pharmacol* 2018; 22: 1-15.
  - 41) Bischoff J, Aikawa E. Progenitor cells confer plasticity to cardiac valve endothelium. *J Cardiovasc Transl Res* 2011; 4: 710-719.
  - 42) Cushing MC, Mariner PD, Liao JT, Sims EA, Anseth KS. Fibroblast growth factor represses Smad-mediated myofibroblast activation in aortic valvular interstitial cells. *FASEB J* 2008; 22: 1769-1777.
  - 43) Chakrabarti M, Bhattacharya A. Increased TGF $\beta$ 1 and SMAD3 Contribute to Age-Related Aortic Valve Calcification. *Front Cardiovasc Med* 2022; 9: 770065.
  - 44) Goody PR, Hosen MR, Christmann D, Niepmann ST, Zietzer A, Adam M, Bönner F, Zimmer S, Nickenig G, Jansen F. Aortic valve stenosis: From basic mechanisms to novel therapeutic targets. *Arterioscler Thromb Vasc Biol* 2020; 40: 885-900.
  - 45) Rutkovskiy A, Malashicheva A, Sullivan G, Bogdanova M, Kostareva A, Stensløkken KO, Fiane A, Vaage J. Valve interstitial cells: The key to understanding the pathophysiology of heart valve calcification. *J Am Heart Assoc* 2017; 6: e006339.
  - 46) Rogers MA, Chen J, Nallamshetty S, Pham T, Goto S, Muehlschlegel JD, Libby P, Aikawa M, Aikawa E, Plutzky J. Retinoids repress human cardiovascular cell calcification with evidence for distinct selective retinoid modulator effects. *Arterioscler Thromb Vasc Biol* 2020; 40: 656-669.
  - 47) Balistreri CR, Crapanzano F, Schirone L, Allegra A, Pisano C, Ruvolo G, Forte M, Greco E, Cavarretta E, Marullo AGM, Sciarretta S, Frati G. Dereglulation of Notch1 pathway and circulating endothelial progenitor cell (EPC) number in patients with bicuspid aortic valve with and without ascending aorta aneurysm. *Sci Rep* 2018; 8: 13834.
  - 48) Wang Y, Fang Y, Lu P, Wu B, Zhou B. NOTCH Signaling in Aortic Valve Development and Calcific Aortic Valve Disease. *Front Cardiovasc Med* 2021; 8: 682298.
  - 49) Shimizu T, Tanaka T, Iso T, Matsui H, Ooyama Y, Kawai-Kowase K, Arai M, Kurabayashi M. Notch signaling pathway enhances bone morphogenetic protein 2 (BMP2) responsiveness of Msx2 gene to induce osteogenic differentiation and mineralization of vascular smooth muscle cells. *J Biol Chem* 2011; 286: 19138-19148.
  - 50) Kawaguchi A, Miyatake K, Yutani C, Beppu S, Tsumishima M, Yamamura T, Yamamoto A. Characteristic cardiovascular manifestation in homozygous and heterozygous familial hypercholesterolemia. *Am Heart J* 1999; 137: 410-418.
  - 51) Kamath AR, Pai RG. Risk factors for progression of calcific aortic stenosis and potential therapeutic targets. *Int J Angiol* 2008; 17: 63-70.
  - 52) Mohty D, Pibarot P, Després JP, Côté C, Arsenault B, Cartier A, Cosnay P, Couture C, Mathieu P. Association between plasma LDL particle size, valvular accumulation of oxidized LDL, and inflammation in patients with aortic stenosis. *Arterioscler Thromb Vasc Biol* 2008; 28: 187-193.
  - 53) Olsson M, Thyberg J, Nilsson J. Presence of oxidized low-density lipoprotein in nonrheumatic stenotic aortic valves. *Arterioscler Thromb Vasc Biol* 1999; 19: 1218-1222.
  - 54) Pagé A, Dumesnil JG, Clavel MA, Chan KL, Teo KK, Tam JW, Mathieu P, Després JP, Pibarot P. Metabolic Syndrome Is Associated With More Pronounced Impairment of Left Ventricle Geometry and Function in Patients With Calcific Aortic Stenosis. A Substudy of the ASTRONOMER (Aortic Stenosis Progression Observation Measuring Effects of Rosuvastatin). *J Am Coll Cardiol* 2010; 55: 1867-1874.
  - 55) Larsson SC, Wolk A, Håkansson N, Bäck M. Overall and abdominal obesity and incident aortic valve stenosis: Two prospective cohort studies. *Eur Heart J* 2017; 38: 2192-2197.
  - 56) Zamorano JL, Gonçalves A. Adiponectin: A Novel Target for Aortic Stenosis Medical Treatment? *Cardiology* 2011; 118: 248-250.
  - 57) Mohty D, Pibarot P, Côté N, Cartier A, Audet A, Després JP, Mathieu P. Hypoadiponectinemia is associated with valvular inflammation and faster disease progression in patients with aortic stenosis. *Cardiology* 2011; 118: 140-146.
  - 58) Carnevale R, Pastori D, Peruzzi M, De Falco E, Chimenti I, Biondi-Zoccai G, Greco E, Marullo AG, Nocella C, Violi F, Pignatelli P, Calvieri C, Frati G. Total adiponectin is inversely associated with platelet activation and CHA2DS2-VASc score in anticoagulated patients with atrial fibrillation. *Mediators Inflamm* 2014; 2014: 908901.
  - 59) Rosenhek R, Baumgartner H. Aortic sclerosis, aortic stenosis and lipid-lowering therapy. *Expert Rev Cardiovasc Ther* 2008; 6: 385-390.
  - 60) De Vecchis R, Di Biase G, Esposito C, Ciccarelli A, Cioppa C, Giasi A, Ariano C, Cantatrione S. Statin use for nonrheumatic calcific aortic valve stenosis: A review with meta-analysis. *J Cardiovasc Med* 2013; 14: 559-567.
  - 61) Venardos N, Deng XS, Yao Q, Weyant MJ, Reece TB, Meng X, Fullerton DA. Simvastatin reduces the TLR4-induced inflammatory response in hu-

- man aortic valve interstitial cells. *J Surg Res* 2018; 230: 101-109.
- 62) Rajamannan NM, Subramaniam M, Caira F, Stock SR, Spelsberg TC. Atorvastatin inhibits hypercholesterolemia-induced calcification in the aortic valves via the Lrp5 receptor pathway. *Circulation* 2005; 112: I-229-I-239.
- 63) Peruzzi M, De Luca L, Thomsen HS, Romagnoli E, D'Ascenzo F, Mancone M, Sardella G, Lucisano L, Abbate A, Frati G, Biondi-Zoccai G. A Network Meta-Analysis on Randomized Trials Focusing on the Preventive Effect of Statins on Contrast-Induced Nephropathy. *Biomed Res Int* 2014; 2014: 213239.
- 64) Cowell SJ, Newby DE, Prescott RJ, Bloomfield P, Reid J, Northridge DB, Boon NA; Scottish Aortic Stenosis and Lipid Lowering Trial, Impact on Regression (SALTIRE) Investigators. A randomized trial of intensive lipid-lowering therapy in calcific aortic stenosis. *N Engl J Med* 2005; 352: 2389-2397.
- 65) Chan KL, Teo K, Dumesnil JG, Ni A, Tam J. ASTRONOMER Investigators. Effect of lipid lowering with rosuvastatin on progression of aortic stenosis: Results of the aortic stenosis progression observation: Measuring effects of rosuvastatin (Astronomer) trial. *Circulation* 2010; 121: 306-314.
- 66) Rossebø AB, Pedersen TR, Boman K, Brudi P, Chambers JB, Egstrup K, Gerdts E, Gohlke-Bärwolf C, Holme I, Kesäniemi YA, Malbecq W, Nienaber CA, Ray S, Skjaerpe T, Wachtell K, Wilenheimer R; SEAS Investigators. Intensive lipid lowering with simvastatin and ezetimibe in aortic stenosis. *N Engl J Med* 2008; 359: 1343-1356.
- 67) Moura LM, Ramos SF, Zamorano JL, Barros IM, Azevedo LF, Rocha-Gonçalves F, Rajamannan NM. Rosuvastatin Affecting Aortic Valve Endothelium to Slow the Progression of Aortic Stenosis. *J Am Coll Cardiol* 2007; 49: 554-561.
- 68) Owens DS, Katz R, Johnson E, Shavelle DM, Probstfield JL, Takasu J, Crouse JR, Carr JJ, Kronmal R, Budoff MJ, O'Brien KD. Interaction of age with lipoproteins as predictors of aortic valve calcification in the multi-ethnic study of atherosclerosis. *Arch Intern Med* 2008; 168: 1200-1207.
- 69) Tastet L, Capoulade R, Shen M, Clavel MA, Côté N, Mathieu P, Arsenault M, Bédard É, Tremblay A, Samson M, Bossé Y, Dumesnil JG, Arsenault BJ, Beaudoin J, Bernier M, Després JP, Pibarot P. ApoB/ApoA-I ratio is associated with faster hemodynamic progression of aortic stenosis: Results from the progressa (metabolic determinants of the progression of aortic stenosis) study. *J Am Heart Assoc* 2018; 7: e007980.
- 70) Zheng KH, Arsenault BJ, Kaiser Y, Khaw KT, Wareham NJ, Stroes ESG, Boekholdt SM. apoB/apoA-I Ratio and Lp(a) Associations With Aortic Valve Stenosis Incidence: Insights From the EPIC-Norfolk Prospective Population Study. *J Am Heart Assoc* 2019; 8: e013020.
- 71) Schnitzler JG, Ali L, Groenen AG, Kaiser Y, Kroon J. Lipoprotein(A) as orchestrator of calcific aortic valve stenosis. *Biomolecules* 2019; 9: 760.
- 72) O'Donoghue ML, Fazio S, Giugliano RP, Stroes ESG, Kanevsky E, Gouni-Berthold I, Im K, Lira-Pineda A, Wasserman SM, Češka R, Ezhov MV, Jukema JW, Jensen HK, Tokgözoğlu SL, Mach F, Huber K, Sever PS, Keech AC, Pedersen TR, Sabatine MS. Lipoprotein(a), PCSK9 Inhibition, and Cardiovascular Risk. *Circulation* 2019; 139: 1483-1492.
- 73) Orsó E, Schmitz G. Lipoprotein(a) and its role in inflammation, atherosclerosis and malignancies. *Clin Res Cardiol Suppl* 2017; 12: 31-37.
- 74) Tsimikas S. Potential Causality and Emerging Medical Therapies for Lipoprotein(a) and Its Associated Oxidized Phospholipids in Calcific Aortic Valve Stenosis. *Circ Res* 2019; 124: 405-415.
- 75) Kamstrup PR, Tybjaerg-Hansen A, Nordestgaard BG. Elevated lipoprotein(a) and risk of aortic valve stenosis in the general population. *J Am Coll Cardiol* 2014; 63: 470-477.
- 76) Langsted A, Nordestgaard BG, Benn M, Tybjaerg-Hansen A, Kamstrup PR. PCSK9 R46L loss-of-function mutation reduces lipoprotein(a), LDL cholesterol, and risk of aortic valve stenosis. *J Clin Endocrinol Metab* 2016; 101: 3281-3287.
- 77) Benn M, Nordestgaard BG, Grande P, Schnohr P, Tybjaerg-Hansen A. PCSK9 R46L, Low-Density Lipoprotein Cholesterol Levels, and Risk of Ischemic Heart Disease. 3 Independent Studies and Meta-Analyses. *J Am Coll Cardiol* 2010; 55: 2833-2842.
- 78) Lagace TA, Curtis DE, Garuti R, McNutt MC, Park SW, Prather HB, Anderson NN, Ho YK, Hammer RE, Horton JD. Secreted PCSK9 decreases the number of LDL receptors in hepatocytes and in livers of parabiotic mice. *J Clin Invest* 2006; 116: 2995-3005.
- 79) Murphy SA, Pedersen TR, Gaciong ZA, Ceska R, Ezhov MV, Connolly DL, Jukema JW, Toth K, Tikkanen MJ, Im K, Wiviott SD, Kurtz CE, Honarpour N, Giugliano RP, Keech AC, Sever PS, Sabatine MS. Effect of the PCSK9 Inhibitor Evolocumab on Total Cardiovascular Events in Patients with Cardiovascular Disease: A Prespecified Analysis from the FOURIER Trial. *JAMA Cardiol* 2019; 4: 613-619.
- 80) Cao YX, Liu HH, Li S, Li JJ. A Meta-Analysis of the Effect of PCSK9-Monoclonal Antibodies on Circulating Lipoprotein (a) Levels. *Am J Cardiovasc Drugs* 2019; 19: 87-97.
- 81) Gallego-Colon E, Daum A, Yosefy C. Statins and PCSK9 inhibitors: A new lipid-lowering therapy. *Eur J Pharmacol* 2020; 878: 173114.
- 82) Schwartz GG, Steg PG, Szarek M, Bhatt DL, Bittner VA, Diaz R, Edelberg JM, Goodman SG, Hanotin C, Harrington RA, Jukema JW, Lecorps G, Mahaffey KW, Moryusef A, Pordy R, Quintero K, Roe MT, Sasiela WJ, Tamby JF, Tricoci P, White HD, Zeiher AM; ODYSSEY OUTCOMES Committees and Investigators. Alirocumab and Cardiovascular Outcomes after Acute Coronary Syndrome. *N Engl J Med* 2018; 379: 2097-2107.
- 83) Sabatine MS, Giugliano RP, Keech AC, Honarpour N, Wiviott SD, Murphy SA, Kuder JF, Wang H, Liu

- T, Wasserman SM, Sever PS, Pedersen TR; FOURIER Steering Committee and Investigators. Evolocumab and Clinical Outcomes in Patients with Cardiovascular Disease. *N Engl J Med* 2017; 376: 1713-1722.
- 84) Bergmark BA, O'Donoghue ML, Murphy SA, Kuder JF, Ezhov MV, Ceška R, Gouni-Berthold I, Jensen HK, Tokgozoglu SL, Mach F, Huber K, Gaciong Z, Lewis BS, Schiele F, Jukema JW, Pedersen TR, Giugliano RP, Sabatine MS. An Exploratory Analysis of Proprotein Convertase Subtilisin/Kexin Type 9 Inhibition and Aortic Stenosis in the FOURIER Trial. *JAMA Cardiol* 2020; 5: 709-713.
  - 85) Effect of PCSK9 InhibitorS On Calcific Aortic Valve Disease (EPISODE), <https://classic.clinicaltrials.gov/ct2/show/NCT04968509>.
  - 86) Viney NJ, van Capelleveen JC, Geary RS, Xia S, Tami JA, Yu RZ, Marcovina SM, Hughes SG, Graham MJ, Crooke RM, Crooke ST, Witztum JL, Stroes ES, Tsimikas S. Antisense oligonucleotides targeting apolipoprotein(a) in people with raised lipoprotein(a): two randomised, double-blind, placebo-controlled, dose-ranging trials. *Lancet* 2016; 388: 2239-2253.
  - 87) Tsimikas S, Viney NJ, Hughes SG, Singleton W, Graham MJ, Baker BF, Burkey JL, Yang Q, Marcovina SM, Geary RS, Crooke RM, Witztum JL. Antisense therapy targeting apolipoprotein(a): a randomised, double-blind, placebo-controlled phase 1 study. *Lancet* 2015; 386: 1472-83.
  - 88) Gencer B, Kronenberg F, Stroes ES, Mach F. Lipoprotein(a): The revenant. *Eur Heart J* 2017; 38: 1553-1560.
  - 89) Tsimikas S, Karwatowska-Prokopczuk E, Gouni-Berthold I, Tardif JC, Baum SJ, Steinhagen-Thiessen E, Shapiro MD, Stroes ES, Moriarty PM, Nordestgaard BG, Xia S, Guerriero J, Viney NJ, O'Dea L, Witztum JL; AKCEA-APO(a)-LRx Study Investigators. Lipoprotein(a) Reduction in Persons with Cardiovascular Disease. *N Engl J Med* 2020; 382: 244-255.
  - 90) Stiekema LCA, Prange KHM, Hoogeveen RM, Verweij SL, Kroon J, Schnitzler JG, Dzobo KE, Cupido AJ, Tsimikas S, Stroes ESG, de Winther MPJ, Bahjat M. Potent lipoprotein(a) lowering following apolipoprotein(a) antisense treatment reduces the pro-inflammatory activation of circulating monocytes in patients with elevated lipoprotein(a). *Eur Heart J* 2020; 41: 2262-2271.
  - 91) Thomas GS, Cromwell WC, Ali S, Chin W, Flaim JD, Davidson M. Mipomersen, an apolipoprotein b synthesis inhibitor, reduces atherogenic lipoproteins in patients with severe hypercholesterolemia at high cardiovascular risk: A randomized, double-blind, placebo-controlled trial. *J Am Coll Cardiol* 2013; 62: 2178-2184.
  - 92) Mathieu P, Boulanger MC. Autotaxin and Lipoprotein Metabolism in Calcific Aortic Valve Disease. *Front Cardiovasc Med* 2019; 6: 18.
  - 93) Shroff GR, Bangalore S, Bhawe NM, Chang TI, Garcia S, Mathew RO, Rangaswami J, Ternacle J, Thourani VH, Pibarot P; American Heart Association Council on the Kidney in Cardiovascular Disease and Stroke Council. Evaluation and Management of Aortic Stenosis in Chronic Kidney Disease: A Scientific Statement From the American Heart Association. *Circulation* 2021; 143: e1088-e1114.
  - 94) Massera D, Buzkova P, Bortnick AE, Owens DS, Mao S, Li D, De Boer IH, Kestenbaum BR, Budoff MJ, Kizer JR. Bone mineral density and long-term progression of aortic valve and mitral annular calcification: The Multi-Ethnic Study of Atherosclerosis. *Atherosclerosis* 2021; 335: 126-134.
  - 95) Pawade T, Sheth T, Guzzetti E, Dweck MR, Clavel MA. Why and How to Measure Aortic Valve Calcification in Patients With Aortic Stenosis. *JACC Cardiovasc Imaging* 2019; 12: 1835-1848.
  - 96) Akahori H, Tsujino T, Masuyama T, Ishihara M. Mechanisms of aortic stenosis. *J Cardiol* 2018; 71: P215-P220.
  - 97) Diédhiou D, Cuny T, Sarr A, Norou Diop S, Klein M, Weryha G. Efficacy and safety of denosumab for the treatment of osteoporosis: A systematic review. *Ann Endocrinol (Paris)* 2015; 76: 650-657.
  - 98) Skolnick AH, Osranek M, Formica P, Kronzon I. Osteoporosis Treatment and Progression of Aortic Stenosis. *Am J Cardiol* 2009; 104: 122-124.
  - 99) Lerman DA, Prasad S, Alotti N. Denosumab could be a Potential Inhibitor of Valvular Interstitial Cells Calcification in vitro. *Int J Cardiovasc Res* 2016; 5: 10.4172/2324-8602.1000249.
  - 100) Alishiri G, Heshmat-Gahdarijani K, Hashemi M, Zavar R, Farahani MM. Alendronate slows down aortic stenosis progression in osteoporotic patients: An observational prospective study. *J Res Med Sci* 2020; 25: 65.
  - 101) Pawade TA, Doris MK, Bing R, White AC, Forsyth L, Evans E, Graham C, Williams MC, van Beek EJR, Fletcher A, Adamson PD, Andrews JPM, Carlidge TRG, Jenkins WSA, Syed M, Fujisawa T, Lucatelli C, Fraser W, Ralston SH, Boon N, Prendergast B, Newby DE, Dweck MR. Effect of Denosumab or Alendronic Acid on the Progression of Aortic Stenosis: A Double-Blind Randomized Controlled Trial. *Circulation* 2021; 143: 2418-2427.
  - 102) Zeng Q, Song R, Fullerton DA, Ao L, Zhai Y, Li S, Ballak DB, Cleveland JC Jr, Reece TB, McKinsey TA, Xu D, Dinarello CA, Meng X. Interleukin-37 suppresses the osteogenic responses of human aortic valve interstitial cells in vitro and alleviates valve lesions in mice. *Proc Natl Acad Sci U S A* 2017; 114: 1631-1636.
  - 103) The E, de Graaf DM, Zhai Y. Interleukin 38 alleviates aortic valve calcification by inhibition of NLRP3. *Proc Natl Acad Sci U S A* 2022; 119: e2202577119.
  - 104) Colchicine and Inflammation in Aortic Stenosis (CHIANTI). Available at: <https://classic.clinicaltrials.gov/ct2/show/NCT05162742>.
  - 105) Colchicine and Inflammation in Aortic Stenosis. Available at: <https://www.clinicaltrialsregister.eu/ctr-search/trial/2021-005586-40/NL>.

- 106) Diederichsen ACP, Lindholt JS, Möller S, Øvrehus KA, Auscher S, Lambrechtsen J, Hobond SE, Alan DH, Urbonaviciene G, Becker SW, Fredgart MH, Hasific S, Folkestad L, Gerke O, Rasmussen LM, Møller JE, Mickley H, Dahl JS. Vitamin K2 and D in Patients With Aortic Valve Calcification: A Randomized Double-Blinded Clinical Trial. *Circulation* 2022; 145: 1387-1397
- 107) SLOW-Slower Progress of caLcificatiOn With Vitamin K2. Available at: <https://classic.clinicaltrials.gov/ct2/show/NCT04429035>.
- 108) Wypasek E, Natorska J, Mazur P, Kopytek M, Gawęda B, Kapusta P, Madeja J, Iwaniec T, Kapelak B, Undas A. Effects of rivaroxaban and dabigatran on local expression of coagulation and inflammatory factors within human aortic stenotic valves. *Vascul Pharmacol* 2020; 130: 106679.