

LncRNA BX357664 inhibits the proliferation and invasion of non-small cell lung cancer cells

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Abstract. – OBJECTIVE: To explore the level of long non-coding RNA (lncRNA) BX357664 in non-small cell lung cancer (NSCLC) and its role in the development of NSCLC. Meanwhile, the potential regulatory mechanism of BX357664 was also what we were interested in.

PATIENTS AND METHODS: Real-time quantitative polymerase chain reaction (RT-qPCR) was performed to examine the level of BX357664 in 82 pairs of cancer tissues and adjacent normal tissues collected from patients with NSCLC, and the relationship between BX357664 level and pathological parameters or prognosis of NSCLC patients was analyzed. Further verification by RT-qPCR was to examine BX357664 expression in NSCLC cell lines, and BX357664 overexpression model was constructed using lentivirus in NSCLC cell lines including SPC and H1299. In addition, cell cycle, cell apoptosis, cell clone formation assay and tube formation assay were performed to analyze the influence of BX357664 on the biological functions of NSCLC cells. Western Blot was conducted to explore its underlying mechanisms.

RESULTS: RT-qPCR results indicated that BX357664 in NSCLC tissues remarkably higher than that in normal tissues compared with patients with highly expressed BX357664, patients with lowly-expressed had worse tumor stage, higher incidence of lymph node metastasis or distant metastasis and lower overall survival rate. In addition, compared with NC group, the proliferation, migration and invasion ability of cells in BX357664 overexpression group was attenuated significantly. The key proteins in TGF- β 1/Smad pathway, including transforming growth factor- β 1 (TGF- β 1), p-Smad2, p-Smad3, N-cad, Vimentin and MMP-9 were also remarkably reduced.

CONCLUSIONS: BX357664 level was significantly reduced in tumor tissues of NSCLC patients, resulting in advanced tumor staging, lymph node metastasis, distant metastasis, and poor prognosis. Additionally, BX357664 may inhibit the proliferation as well as invasion and

migration of NSCLC cells. TGF- β 1/Smad pathway

Key Words:

BX357664, TGF- β 1 signaling pathway, Non-small cell lung cancer, Metastasis, Metastasis,

Introduction

Lung cancer refers to malignant tumors originating from the bronchial or bronchiole mucosal epithelium. The incidence and mortality rate of lung carcinoma are so high that it has become one of the most common malignant tumors in the world, and the number of patients with carcinoma of lung is increasing year by year. Hence, this disease has become a huge threat to human life and the first cause of cancer-related death¹⁻³. Lung carcinoma is mainly divided into two categories, which are small cell lung cancer (SCLC) and non-small cell lung cancer (NSCLC)^{4,5}. Among them, the incidence of the latter accounts for more than 85% of this cancer, and advanced patients account for 40-50%; what's worse, current radiotherapy and chemotherapy are not sensitive enough to these advanced patients⁵. Non-small cell lung cancer is basically divided into squamous cell carcinoma and non-squamous cell carcinoma (mainly with large cell carcinoma, adenocarcinoma, and other cell types), of which squamous cell carcinoma accounts for about 30-35% of all lung cancers and central type is the most common^{6,7}. Adenocarcinoma accounts for about 35-40% of all lung cancers, which has a higher incidence in non-smokers and the surrounding type is more common⁸. Based on the high recurrence rate in early stage of non-small cell lung cancer and the lack of effective treatment in the advanced stage of the disease, the 5-year survival rate of patients with NSCLC ranges from about 65% of patients in tumor node metastasis (TNM) stage I to less than

1% of patients in stage IV, and the overall 5-year survival rate is less than 16%⁸⁻¹⁰. Patients in stage III or higher without treatment has an average survival of less than 7 months, so the therapeutic method of NSCLC has become a clinical problem^{9,10}. In recent years, as the advancement of science and technology, certain progress has been made in diagnosis and treatment of NSCLC. With the discovery of a series of tumor-targeting genes, a number of drugs targeting NSCLC-involved genes have emerged¹¹. With more and more in-depth research on the pathogenesis of tumor development and development, it is now basically determined that the body's response to the tumor are caused by changes in certain gene activities and expression in tumor cells. Long non-coding RNA (LncRNA) is one kind of non-coding RNA with the length greater than 200 nucleotides^{12,13}. Although LncRNA does not encode proteins, its involvement can constitute a complicated and very important network of gene expression regulation^{14,15}. Recent studies¹⁵ have shown that LncRNAs play a vital role in the development of normal tissues and the regulation of cellular pluripotency as well as cell differentiation. In addition, LncRNAs are involved in the control of multiple molecular pathways, such as changes in gene expression that ultimately regulate cell proliferation, apoptosis, and cell migration^{16,17}. Therefore, the expression disorder of LncRNA is closely related to various diseases in human, such as tumor formation^{15,18}. A number of LncRNAs, LncRNA BX357664 can participate in lots of pathophysiological processes, such as cell adhesion, migration and proliferation^{19,20}. In the discovery, many studies have shown that BX357664 is dysregulated in many malignant tumors, such as kidney cancer, colon cancer, etc. Meanwhile, its expression level is closely associated with the pathological grade, clinical stage and prognosis of some tumors, but its expression in NSCLC has not been reported^{19,20}.

The transforming growth factor- β 1 (TGF- β 1) signaling pathway regulates downstream transcription factors-mediated epithelial mesenchymal transition (EMT) to promote tumor invasion and metastasis²¹. TGF- β 1 has been shown to induce EMT in a variety of epithelial cells, including fibroblasts, hepatocytes and mammary epithelial cells²¹⁻²³. Experimental studies²¹⁻²³ of cell culture *in vitro* and metastatic tumors *in vivo* have both confirmed that TGF- β 1 can regulate EMT. Therefore, TGF- β 1 plays a crucial role in EMT no matter *in vitro* or *in vivo*²¹. The TGF- β 1 signaling network is mainly comprised of two

transduction pathways including the classical Smad-dependent pathway and the Smad-independent pathway²⁴⁻²⁶. Therefore, this work aims to explore whether BX357664 can mediate the molecular mechanism of invasion and metastasis of NSCLC through TGF- β 1/Smad signaling pathway and thus provide experimental evidence for its clinical application.

In our research, we examined the expression of LncRNA BX357664 in 82 pairs of NSCLC tissues and adjacent tissues and analyzed the relationship between BX357664 level and clinicopathological factors to explore how BX357664 affects the occurrence and progression of NSCLC via TGF- β 1/Smad signaling pathway.

Patients and Methods

Patients and NSCLC Samples

82 cases of primary NSCLC lesions and 82 non-cancerous tissues (parallel tissues were more than 5 cm away from cancerous tissues) were resected, and personal information and detailed clinical data of each patient were collected. All patients were diagnosed as NSCLC by postoperative pathological analysis, and no anti-tumor treatment such as radiotherapy or chemotherapy was performed before surgery. The study was approved by the Ethics Committee of the First Affiliated Hospital of Jinzhou Medical University and relevant informed consent forms were signed by patients and their families.

Cell Lines and Reagents

The human lung cancer cell lines (SKMES1, SPCA1, H358, H1299, A549) and normal lung cell line, 16HBE, were provided by American Type Culture Collection (ATCC, Manassas, VA, USA). High Glucose Dulbecco's Modified Eagle Medium (DMEM) medium and fetal bovine serum (FBS) were purchased from Life Technologies (Gaithersburg, MD, USA). The cells were cultured with DMEM medium containing 10% fetal calf serum (FCS) in a 37°C, 5% CO₂ incubator.

Cell Transfection

Sequences of knockdown or overexpression and the corresponding control were designed by Shanghai Jima Co., Ltd. (Shanghai, China) according to the gene sequence of BX357664 to be loaded in the lentiviral vector. The cells in logarithmic growth phase were seeded into 6-well plates, and the appropriate amount of virus solu-

tion (calculated according to viral MOI) was added and incubated in a cell culture incubator for 48 hours. The fluorescence intensity was evaluated under microscope to evaluate the transfection efficiency for RT-qPCR analysis and cell function experiments. Finally, the stably transfected cell line was screened with puromycin.

Cell Proliferation Assay

After 48 h of transfection, cells were digested and seeded into 96-well plates at 2000 cells per well. After 6 h, 24 h, 48 h and 72 h, the cells were added with cell counting kit-8 (CCK-8) (Dojindo Laboratories, Kumamoto, Japan) reagent, respectively. After incubation for 2 hours, the OD value of each well was measured using a microplate reader.

Cell Scratch

Marker pen was first used to draw a horizontal line with a ruler on the back of the 6-well plate. About 5×10^5 cells were added in each well, and the specific amount was different depending on the cells. The next day, tip of gun was used to draw another line perpendicular to the horizontal one, and the tip of the gun should be vertical. The cells were washed 3 times with phosphate-buffered saline (PBS), the scratched cells were removed and serum-free medium was added. Cell samples were taken out at 0, 12, 24, 48 hours for taking pictures.

Transwell assay

After transfection for 48 hours, the cells were trypsinized and resuspended in serum-free medium. After cell counting, cells were diluted to $2.0 \times 10^5/\text{mL}$, and the transwell chambers with matrigel or without matrigel were placed in a 24-well plate. $500 \mu\text{L}$ of the cell suspension was added to the upper chamber, and $500 \mu\text{L}$ medium containing 10% fetal bovine serum (FBS) were added to the lower chamber. After 48 hours of incubation, the chamber was taken out, fixed with 4% paraformaldehyde for 30 minutes, and stained by crystal violet for 15 minutes. After washed with PBS, the inner surface of the chamber was carefully cleaned to remove the cells in inner layer. The stained migrated cells in the outer layer of the chamber were observed under the microscope, and 50 cells per view were randomly selected.

Real-Time Quantitative Polymerase Chain Reaction (RT-qPCR)

Total RNA was extracted using TRIzol (Invitrogen, Carlsbad, CA, USA), and reverse transcription was performed using Quant script RT kit (China

Tianguen, Beijing, China). The BX357664 mRNA was quantified by Real-time fluorescent quantitative PCR using the apparatus of ABI's 7900HT. The experimental procedure was performed according to the Invitrogen specification (Carlsbad, CA, USA), and $2^{-\Delta\Delta Ct}$ was used to indicate the relative level of mRNA in tissues. The following primers were used for RT-qPCR reactions: BX357664: forward, 5'-GGCGTGGTTTTGATGGAGTC-3' and reverse, 5'-AGGCTGCAGAGTTGATTCG-3'; MMP-9: forward, 5'-GTGGACAATCGCAAAGAC-3' and reverse, 5'-AAAGGGGTAACGGCACTA-3'.

Western Blot Assay

Cells were lysed using cell lysis buffer, shaken on ice for 30 minutes, and centrifuged at $14,000 \times g$ for 15 min at 4°C . Total protein concentration was then calculated using the Pierce and Warriner NLSCLCA Protein Assay Kit. Extracted proteins were separated using a 10% sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) gel and subsequently transferred to a polyvinylidene difluoride (PVDF) membrane. Western blot analysis was performed according to standard procedures. The primary antibodies were TGF- $\beta 1$, p-Smad2, Smad3, N-cad, Vimentin, MMP-9 and β -actin, and secondary antibodies were anti-mouse IgG and anti-rabbit, all purchased from Cell Signaling Technology (CST, Danvers, MA, USA).

Statistical Analysis

After the data was checked, the database was recorded using Statistical Product and Service Solutions (SPSS) 22.0 (IBM, Armonk, NY, USA). Measurement data were expressed as mean \pm standard deviation ($\bar{x} \pm s$). The continuous variables were analyzed by *t*-test, while the categorical ones were analyzed using χ^2 -test or Fisher's exact probability method. Kaplan-Meier method was applied to evaluate the survival time of patients, and the difference between each curve was compared by Log-rank test. $p < 0.05$ was considered to be statistically significant.

Results

BX357664 was Lowly Expressed in NSCLC Tissues and Cell Lines

RT-qPCR analysis revealed that the mRNA level of BX357664 was markedly decreased in tumor tissues compared with the adjacent normal tissues. The difference was statistically significant (Figure 1A, 1B). At the same time, BX357664 in

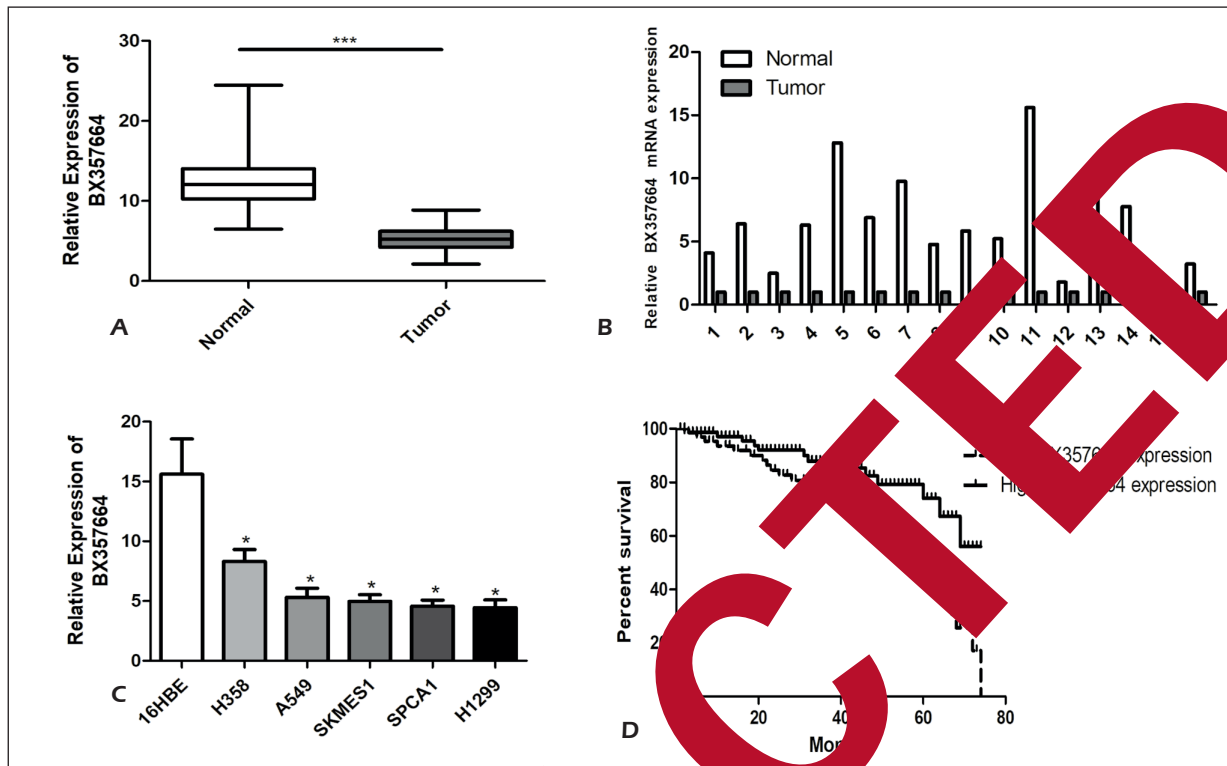


Figure 1. A-B, The expression of BX357664 in 82 NSCLC tissue was significantly decreased by RT-qPCR. C, Expression levels of BX357664 in 5 NSCLC cell lines (H358, H1299, A549) and normal lung cell line (16HBE) by RT-qPCR. D, Kaplan-Meier survival curves of patients with NSCLC stratified on BX357664 expression. Patients in the low expression group had a significantly more unfavorable prognosis than that in high expression group. A representative data set is displayed as mean \pm SD values. * $p < 0.05$, ** $p < 0.01$.

NSCLC cells was also found significantly lower than that in normal lung tissue (Figure 1C), especially in SPCA1 and H1299 cell lines, which were hence chosen for subsequent experiments.

BX357664 Inhibits Cell Proliferation, Correlates with Clinical Stage, Lymph Node or Distant Metastasis, and Overall Survival of NSCLC Patients

Based on the RT-qPCR results, tissues were divided into high-expressed BX357664 group and lowly-expressed group, with the number of each group being counted. Chi-square test was used to analyze the relationship between the lowly-expressed BX357664 and some individual information of patients including age, sex, tumor location, lymph node metastasis and distant metastasis. As shown in Table I, the low level of BX357664 was positively correlated with clinical stage, lymph node metastasis, and distant metastasis of NSCLC, but not with age, gender, and tumor location. In addition, we collected relevant follow-up data in order to figure out the relationship

between the level of BX357664 and the prognosis of patients with NSCLC. Kaplan-Meier survival curves indicated that low expression of BX357664 was conspicuously associated with poor prognosis of NSCLC. The lower the level of BX357664, the worse the prognosis ($p < 0.05$; Figure 1D). The above results indicated that BX357664 might be a new biological indicator for predicting the prognosis of NSCLC.

Over-Expression of BX357664 Inhibited Cell Proliferation

In order to investigate the effect of BX357664 on the proliferation of NSCLC cells, we first successfully constructed BX357664 overexpression model (Figure 2A, 2B), and tested cell proliferation of the control group and BX357664 overexpression group using CCK8 assay. As shown in Figures 2C and 2D, cells in BX357664 overexpression group presented a significant decrease in proliferative ability compared with the NC group. And cell clone formation experiments showed the same trend (Figure 2E).

Table I. Association of BX357664 expression with clinicopathological characteristics of NSCLC.

Parameters features	No. of cases	BX357664 expression		p-value
		High (%)	Low (%)	
Age (years)				0.034
< 60	35	21	14	
≥ 60	47	25	22	
Gender				0.030
Male	40	25	15	
Female	42	22	20	
T stage				0.045
T1-T2	46	31	15	
T3-T4	36	16	20	
Lymph node metastasis				0.001
No	48	33	15	
Yes	34	15	19	
Distance metastasis				0.034
No	63	38	11	
Yes	19	8	11	

Over-Expression of BX357664 Inhibited Cell Migration and Invasion

Transwell assay was performed to explore the effects of BX357664 on the migration and invasion of NSCLC cells. The results indicated that compared with the NC group, the number of migrated NSCLC cells in BX357664 over-expression group was remarkably reduced, suggesting that their migratory and invasive ability was strikingly enhanced (Figure 3A), which was consistent with the results of scratch test (Figure 3C, 3D).

Over-Expression of BX357664 Suppressed the Activation of TGF-β1/Smad Pathway

To figure out how BX357664 affect cell proliferation, invasive and migratory capacity, we examined the levels of key proteins including TGF-β1, p-Smad2, p-Smad3, N-cad, Vimentin and MMP-9 in the TGF-β1/Smad pathway after knockdown of BX357664. And the results indicated that the above proteins were strikingly suppressed (Figure 4).

Discussion

Lung cancer remains the leading cause of cancer-related deaths worldwide, with approximately 80% of cases being non-small cell lung cancer (NSCLC)¹⁻³. Carcinoma of lung is generally diagnosed in advanced stages, which predicts poor

prognosis and limitations in treatment options^{3,4}. Lung cancer progression involves the interaction of genetic, epigenetic and environmental factors that can cause the dysregulation of key genes and tumor suppressor genes, ultimately leading to the activation of cancer-related signaling pathways³⁻⁵. Over the past decade, the discovery of multiple molecular aberrations inducing lung cancer has driven the rapid development of lung cancer research⁵. There are still about half of NSCLC cases lacking targetable mutation sites, and there exist different degrees of drug resistance in targeted therapy⁴⁻⁷. Therefore, finding alternative treatment strategies for lung carcinoma is of great significance. Nowadays the pharmacological regulation of the epigenome has been used to treat lung cancer, and molecular biology has found that targeted therapy can prevent tumor growth to avoid its heterogeneity^{3,4}. In this study we discuss the current application of molecular targeted therapy in the treatment and discovery of diagnostic markers of lung cancer^{8,9}. Researches in early diagnosis, metastasis, recurrence and adjuvant therapy after advanced NSCLC have become the focus of current research⁹. Recent studies^{19,20} have discovered that BX357664 plays a vital role in a variety of diseases, including tumors, but it is unclear whether BX357664 is indispensable in the diagnosis, treatment, and prognosis of NSCLC. Therefore, investigating BX357664 level in NSCLC and analyzing its correlation with clinical prognosis will be beneficial to improve the diagnosis and therapy method of NSCLC as well as improve prognosis of patients.

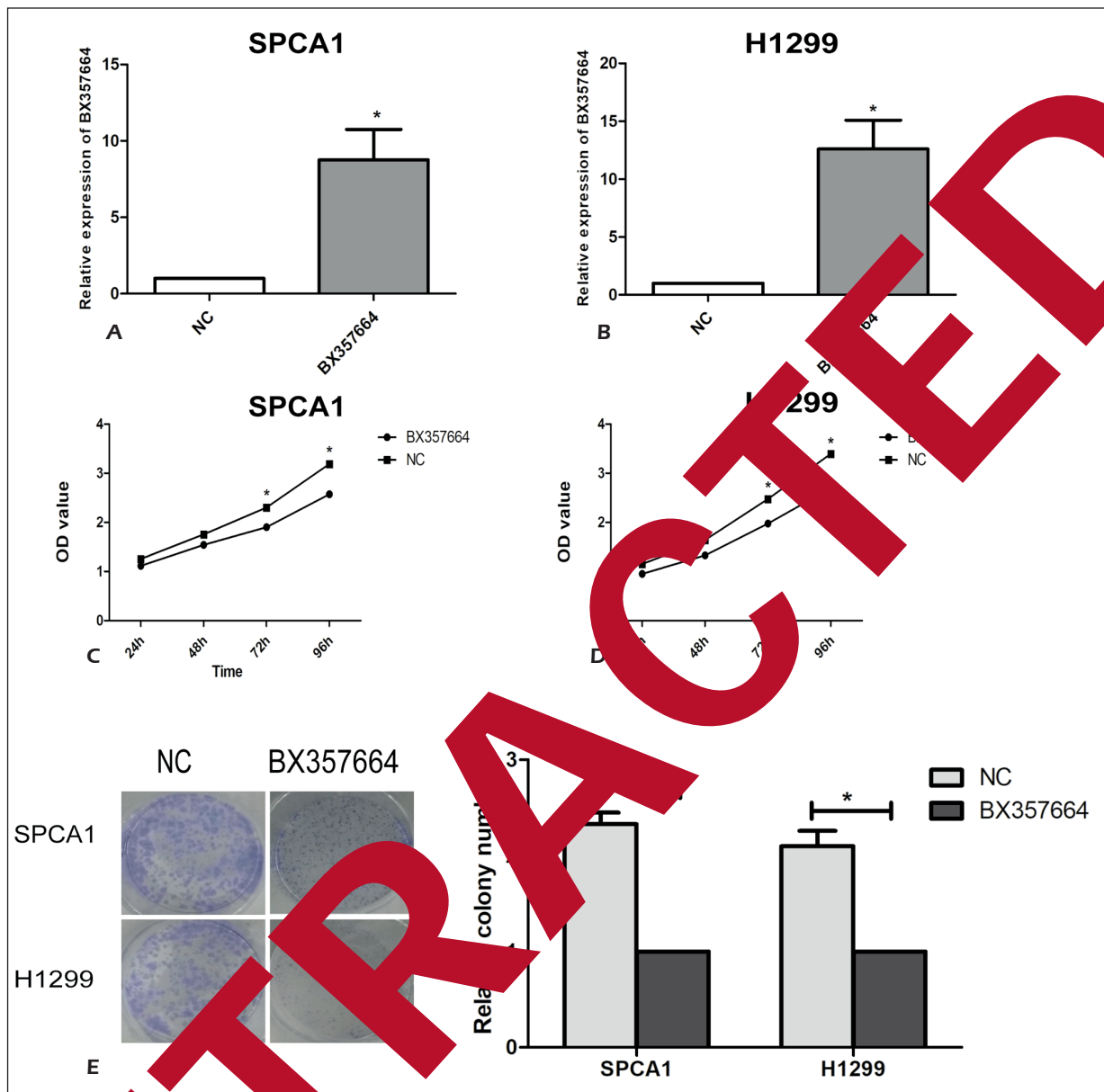


Figure 2 A, B, Western Blot and qPCR were used to verify the efficiency of BX357664 over-expression in SPCA1 and H1299 cell lines. C-D, Growth curve analysis showing the cell growth of SPCA1 and H1299 cells with BX357664 over-expression. E, The efficiencies of cell colony formation in SPCA1 and H1299 cells with BX357664 over-expression. A representative data is displayed as mean \pm SD values. * p <0.05, ** p <0.01.

ing non-coding RNA (LncRNA) is a non-coding RNA of greater than 200 nucleotides in length^{12,13}. It participates in a complex and diverse gene expression regulatory network, which can subtly regulate gene expression¹³. Previous researches have demonstrated that lncRNAs are essential in normal tissue development and regulation of cell pluripotency and differentiation. In addition, lncRNAs are involved in the control of multiple molecular pathways, causing chang-

es in gene expression, and ultimately regulating cell proliferation, apoptosis and migration^{14,15}. Therefore, the expression disorder of lncRNAs is closely related to many diseases, such as tumor formation¹⁵. In addition to being widely expressed in normal tissues, the protein product expressed by BX357664 gene has high levels of expression in various tumor cell tissues: kidney cancer, colorectal cancer, etc.^{19,20}. Studies have observed that BX357664 exerts great influence on migratory and

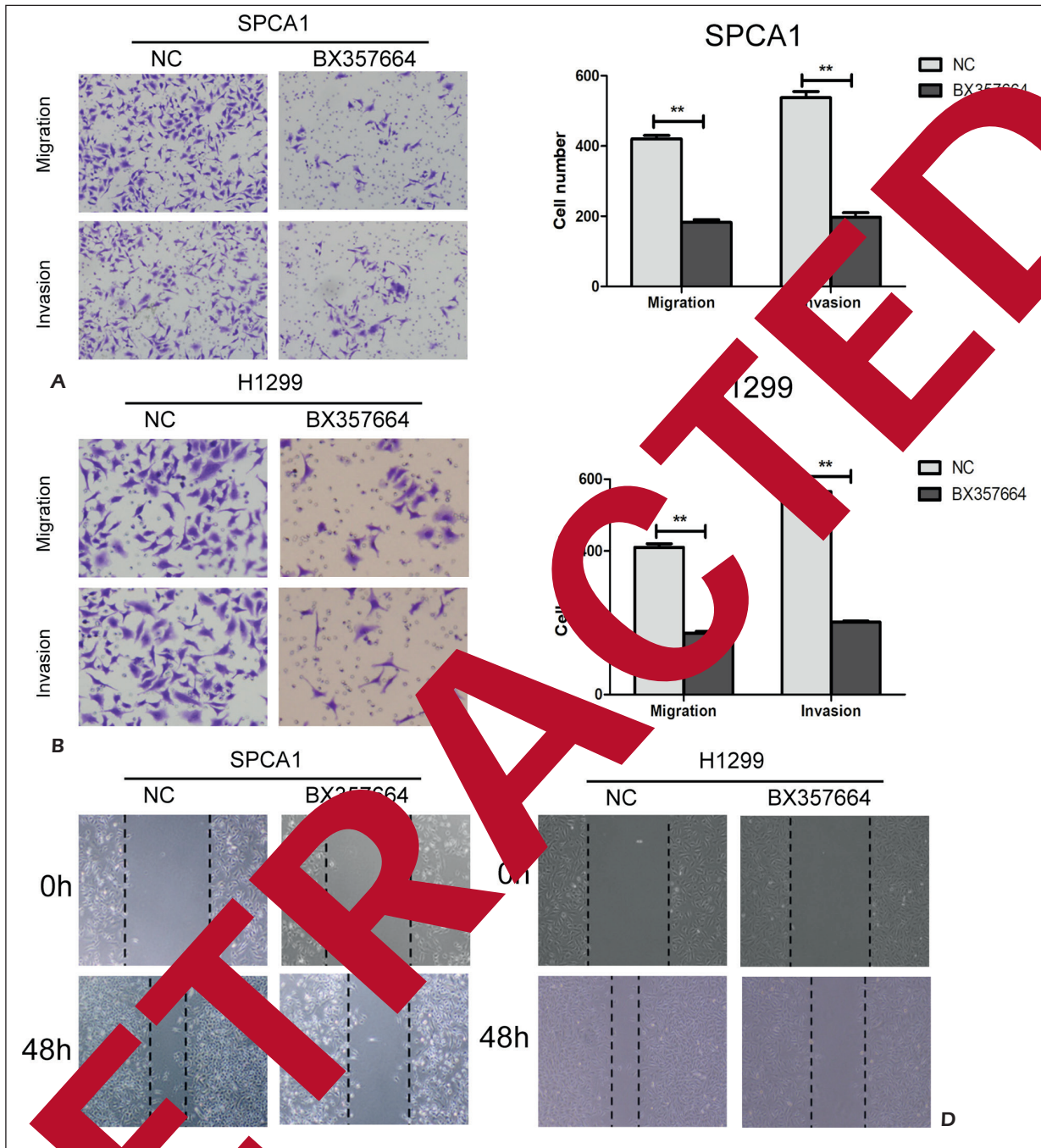


Figure 3. SPCA1 cells transfected with BX357664 displayed significantly lower migration and invasion capacity. **C-D**, H1299 cells transfected with BX357664 displayed significantly lower migration and invasion capacity. **E**, The efficiencies of cell scratch in SPCA1 and H1299 cells with BX357664 over-expression. A representative data set is displayed as mean \pm SD value. * $p < 0.05$, ** $p < 0.01$.

invasive capacity of tumor cells. The differential expression of BX357664 in tumor tissues is closely related to tumor metastasis and prognosis^{19,20}. In our work, we first verified that BX357664 level was dramatically down-regulated in NSCLC tissues compared with adjacent normal tissues

and positively related to NSCLC staging, lymph node or distant metastasis and poor prognosis. Therefore, we believe that BX357664 may play a role of suppressing cancer. To further understand the impact of BX357664 on the biological function of NSCLC cells, we constructed a

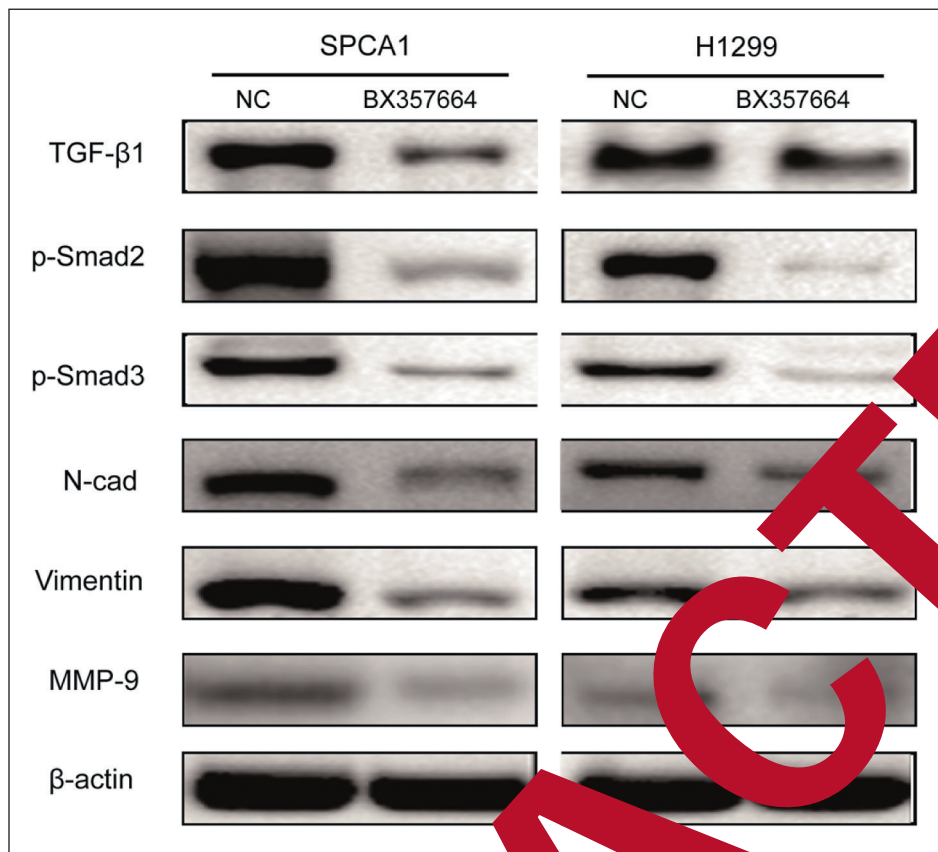


Figure 4. Over-expression of BX357664 expression significantly decreased the expression of TGF- β 1/Smad signal pathway proteins including TGF- β 1, p-Smad2, p-Smad3, N-cad, Vimentin and MMP-9.

BX357664 overexpression modulating lentivirus. And the results of CCK8 and cell assay demonstrated that BX357664 could inhibit the progression of NSCLC; however, the specific molecular mechanism remains unclear. The TGF- β /Smad signaling pathway can be found in various biological tissues in human. Its main functions include transduction of complex receptor signals on the cell surface through autocrine and paracrine pathways, regulation of cell growth, differentiation, apoptosis, and other functions²¹⁻²³. It has been found that TGF- β stimulates epithelial cells to first form a ligand receptor complex with TGF- β type I receptor on the cell membrane, and then phosphorylates TGF- β type I receptor by TGF- β type II receptor (T β RI) to transmit intracellular signal transduction substances. Smad2/3 is phosphorylated and then binds to Smad4 in the cytoplasm to form a heterotrimer or tetramer and undergo nuclear translocation, which enters the nucleus and interacts with nuclear transcription factors to promote the expression of the EMT-related markers so as to finally activate EMT. Most of these transcription factors are transcriptional repressors of E-cadherin, which induce the trans-

formation of epithelial phenotype by inhibiting the expression of E-cadherin²⁴⁻²⁶. It was observed that in keratinized epithelial cells, TGF- β -activated Smad2/3 controls the transdifferentiation of keratinocytes and the expression of the oncogene antagonist MAD1²⁵. Mutations in the Smad2 and Smad4 genes were found in some breast cancer patients²⁷. Similarly, Smad3 expression is associated with bladder cancer and colorectal cancer^{28,29}. In the Smad-dependent pathway, Smad7 and Smad6 prevent the development of EMT as an EMT negative factor²¹⁻²³. Smad6 competitively binds to Smad1 to form an inactive homologous polymer, which negatively regulates the TGF- β /Smad pathway²⁴⁻²⁶. When the ligand activates the TGF- β type I receptor, Smad7 binds to the membrane receptor, which dephosphorylates Smad2/3 and inhibits the TGF- β transduction pathway^{24,26}. To explore whether BX357664 regulates the development of NSCLC through TGF- β 1/Smad pathway, we examined the key proteins including TGF- β 1, p-Smad2, p-Smad3 in the TGF- β 1/Smad pathway after overexpression of BX357664 by Western Blot. The results revealed that the expression levels of the above proteins were signifi-

cantly decreased, suggesting a negative regulation of BX357664 on the TGF- β 1/Smad pathway.

Conclusions

BX357664 was found to have an extremely low expression in NSCLC tissues or cells, which could affect the tumor staging, lymph node or distant metastasis, and poor prognosis. In addition, BX357664 may inhibit cell proliferation or invasive and migratory capacity of NSCLC by regulating TGF- β 1/Smad pathway.

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Conflict of Interests

The authors declared no conflict of interest.

References

- 1) HO JC, LEUNG CC. Management of co-existing tuberculosis and lung cancer. *Cancer* 2012; 122: 83-87.
- 2) WANG S, HU Y. α 7 nicotinic acetylcholine receptors in lung cancer. *Oncol Rep* 2018; 35: 1375-1382.
- 3) NANAVATY P, ALVAREZ JC, ALLEN M. Early cancer screening: advantages, controversies, and applications. *Cancer Control* 2014; 23: 1-14.
- 4) ZHANG Y, YANG S, CHENG S. MicroRNAs: a new key in lung cancer. *Chin J Chemother Pharmacol* 2014; 75: 1105-1111.
- 5) HENNING T, CHAWLA A, BARNETT S, SALGIA R. A personalized treatment for lung cancer: molecular pathways, targeted therapies, and genomic characterization. *Adv Exp Med Biol* 2014; 799: 85-117.
- 6) BRYAN J, CALEIRON M, ROUSSEAU-BUSSAC G, CHOUAID C. New EGFR inhibitors for non-small cell lung cancer. *Oncol Rep* 2018; 35: 4051-4064.
- 7) LIAN Y, LI H. MiRNAs as biomarkers and for the detection of non-small cell lung cancer. *Chin J Thorac Dis* 2018; 10: 3119-3131.
- 8) TIEFENBACHER A, PIRKER R. Systemic treatment of advanced non-small cell lung cancer: controversies and perspectives. *Memo* 2018; 11: 112-115.
- 9) PARUMS DV. Current status of targeted therapy in non-small cell lung cancer. *Drugs Today (Barc)* 2014; 50: 503-525.

- 10) CHEN J, WANG R, ZHANG K, CHEN LB. Long non-coding RNAs in non-small cell lung cancer as biomarkers and therapeutic targets. *J Cell Mol Med* 2014; 18: 2425-2436.
- 11) RECK M, HEIGENER DF, MOK T, SORIA JC. Management of non-small-cell lung cancer: recent developments. *Lancet* 2013; 382: 709-719.
- 12) ANASTASIADOU E, FAGGIONI A, TRIVETTINI C, SLACK FJ. The nefarious nexus of noncoding RNAs in cancer. *Int J Mol Sci* 2018; 19: 2072.
- 13) LIAO Y, CHENG S, XIANG J, LIU C. lncRNA H19 increased proliferation, metastasis and invasion of non-small lung cancer cells and predicted poor survival in non-small lung cancer patients. *Eur Rev Med Pharmacol* 2018; 122: 1686-1692.
- 14) ADAMS BD, PATTERSON C, WANG X, ZHANG C, SLACK FJ. Targeting noncoding RNAs in disease. *J Clin Invest* 2017; 127: 761-771.
- 15) DANG W, WEI X, LIU W, WEN F, GU J, ZHENG H. Long non-coding RNA HOTAIR: target miRNA and signaling pathways. *Cell Mol Lab* 2018; 64: 887-894.
- 16) LIU Y, ZHU QN, DENG JL, LI ZX, WANG G, ZHU YS. Emerging role of long non-coding RNAs in cisplatin resistance. *Onco Targets Ther* 2018; 11: 3185-3194.
- 17) TAKAKA M, WANG DZ. NONCODING RNAs IN CARDIOVASCULAR DISEASE. In: NAKANISHI T, MARKWALD RR, BALDWIN HS, WANG DZ, SRIVASTAVA D, YAMAGISHI H, EDITORS. source: etiology and morphogenesis of congenital heart disease: from gene function and cellular interaction to morphology [internet]. Tokyo: Springer; 2016. Chapter 44.
- 18) DENG J, YANG M, JIANG R, AN N, WANG X, LIU B. Long non-coding RNA HOTAIR regulates the proliferation, self-renewal capacity, tumor formation and migration of the Cancer Stem-Like Cell (CSC) subpopulation enriched from breast cancer cells. *PLoS One* 2017; 12: e170860.
- 19) LIU F, WANG X, LIU H, WANG Y, LIU X, HAO X, LI H. LncRNA BX357664 inhibits cell proliferation and invasion and promotes cell apoptosis in human colorectal cancer cells. *Oncol Lett* 2018; 15: 8237-8244.
- 20) LIU Y, QIAN J, LI X, CHEN W, XU A, ZHAO K, HUA Y, HUANG Z, ZHANG J, LIANG C, SU S, LI P, SHAO P, LI J, QIN C, WANG Z. Long noncoding RNA BX357664 regulates cell proliferation and epithelial-to-mesenchymal transition via inhibition of TGF- β 1/p38/HSP27 signaling in renal cell carcinoma. *Oncotarget* 2016; 7: 81410-81422.
- 21) GAO J, ZHU Y, NILSSON M, SUNDFELDT K. TGF- β 1 isoforms induce EMT independent migration of ovarian cancer cells. *Cancer Cell Int* 2014; 14: 72.
- 22) CHANG H, KIM N, PARK JH, NAM RH, CHOI YJ, PARK SM, CHOI YJ, YOON H, SHIN CM, LEE DH. Helicobacter pylori might induce TGF- β 1-mediated EMT by means of cagE. *Helicobacter* 2015; 20: 438-448.
- 23) KO H, JEON H, LEE D, CHOI HK, KANG KS, CHOI KC. Sanguin H6 suppresses TGF- β 1 induction of the epithelial-mesenchymal transition and inhibits migration and invasion in A549 lung cancer. *Bioorg Med Chem Lett* 2015; 25: 5508-5513.

- 24) PANG L, WEI C, DUAN J, ZOU H, CAO W, QI Y, JIA W, HU J, ZHAO W, JIANG J, LIANG W, LI F. TGF-beta1/Smad signaling, MMP-14, and MSC markers in arterial injury: discovery of the molecular basis of restenosis. *Int J Clin Exp Pathol* 2014; 7: 2915-2924.
- 25) ZEGLINSKI MR, HNATOWICH M, JASSAL DS, DIXON IM. SnoN as a novel negative regulator of TGF-beta/Smad signaling: a target for tailoring organ fibrosis. *Am J Physiol Heart Circ Physiol* 2015; 308: H75-H82.
- 26) HU HH, CHEN DO, WANG YN, FENG YL, CAO G, VAZIRI ND, ZHAO YY. New insights into TGF-beta/Smad signaling in tissue fibrosis. *Chem Biol Interact* 2018; 292: 76-83.
- 27) CASCIONE M, DE MATTEIS V, TOMA CC, LEPORATTI S. Morphomechanical alterations induced by transforming growth factor-beta1 in epithelial breast cancer cells. *Cancers (Basel)* 2018; 10: 234.
- 28) LI J, XU X, MENG S, LIANG Z, WANG X, XU M, LI S, ZHU Y, XIE B, LIN Y, ZHENG X, LIU F, LI L. MiR-323a-3p is involved in regulating epithelial-mesenchymal transition progression in bladder cancer. *Cell Death Dis* 2017; 8: e3010.
- 29) YANG L, LIU Z, TAN J, DONG Y, ZHANG X. Microstructural imaging reveals hyperactive TGF-beta signaling in colorectal cancer. *Cancer Biol Ther* 2016; 17: 105-112.

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