

LncRNA-HEIH suppresses hepatocellular carcinoma cell growth and metastasis by up-regulating miR-199a-3p

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Abstract. – **OBJECTIVE:** The aim of this study was to explore the functional changes of long non-coding ribonucleic acid (lncRNA)-HEIH on hepatocellular carcinoma (HCC) Huh7 and Hep3B cells.

PATIENTS AND METHODS: The expression changes of HEIH in 18 pairs of HCC tissues and adjacent normal tissues were detected by quantitative real time-polymerase chain reaction (qRT-PCR). According to its expression changes in HCC cells silenced by short hairpin ribonucleic acid (shRNA) transfected *in vitro*, these cells were divided into sh-NC group and sh-HEIH group. The effects of highly expressed HEIH on the proliferation, migration and apoptosis of HCC cells were examined *in vitro* with flow cytometry. Western blotting was used to determine the expression changes of epithelial-mesenchymal transition (EMT) proteins, E-cadherin, matrix metalloproteinase (MMP)-2 and MMP-3. In addition, the role of miR-199a-3p as a downstream effector of HEIH in HCC was explored.

RESULTS: Compared with adjacent normal tissues, HEIH was highly expressed in HCC tissues ($p < 0.01$). HEIH silencing significantly inhibited the proliferation and migration, but induced apoptosis of Huh7 cells ($p < 0.05$). The expression of E-cadherin and MMP-2 in sh-HEIH group were markedly lower than those in sh-NC group ($p < 0.05$). Furthermore, miR-199a-3p was identified as a downstream effector of HEIH. The expression of miR-199a-3p increased markedly in Huh7 and Hep3B cells with silenced HEIH ($p < 0.01$). Moreover, when miR-199a-3p expression was inhibited, the effects of HEIH on Huh7 and Hep3B cells were weakened, manifested as notably enhanced cell proliferation and migration capabilities ($p < 0.05$).

CONCLUSION: LncRNA-HEIH suppresses HCC cell growth and metastasis by up-regulating miR-199a-3p. Our findings suggest that HEIH may be a promising target for HCC treatment.

Key Words:

lncRNA-HEIH, Hepatocellular carcinoma (HCC), miR-199a-3p.

Introduction

Primary liver cancer is the second leading cause of cancer-related death in the world¹. In recent years, the incidence rate of liver cancer increases gradually by 4% in men and 3% in women². Hepatocellular carcinoma (HCC) is the main type of primary liver cancer. In the latest decades, the therapeutic methods (surgical resection, liver transplantation, etc.) for HCC has dramatically improved the quality of life of patients. However, the overall 5-year survival rate is still far from satisfactory³. Therefore, improving the 5-year survival rate of HCC is still the direction of our efforts. The development of HCC is a complex and multi-step process implicating diverse molecules and signal transduction pathways⁴. A better knowledge of the incidence and development of HCC will be conducive to the further improvement of its treatment methods. In addition, pinpointing the molecular occurrence mechanism of HCC and discovering effective therapeutic targets can provide new approaches for HCC treatment.

Long non-coding ribonucleic acids (lncRNAs) are a kind of RNAs unable to encode proteins. lncRNAs exert crucial effects in regulating cell cycle, growth and death^{5,6}. Therefore, they are considered as vital factors that can influence the development of various diseases, including malignant tumors⁷. Multiple lncRNAs tend to be disordered in HCC, such as CCHE1⁸, GIHCG⁹, ZNFX1-AS1¹⁰, LINC00052¹¹ and HOTAIR¹². A fraction of lncRNAs is identified as tumor suppressor genes or oncogenes. Consistently, lncRNA HANR facilitates the occurrence and chemoresistance of HCC¹³, while lncRNA uc.134 inhibits the progression of HCC¹⁴. Besides, lncRNAs are considered as promising therapeutic targets for HCC treatment due to their involvement in the occurrence and development of HCC¹⁵. Yang et al¹⁶ reported for the first time that HEIH is a lncRNA highly expressed in HCC tissues. HEIH becomes well-known for its high expression in HBV-related HCC. Zhang et al¹⁷ have revealed that HEIH is conducive to the invasion of HCCMHCC97L and HepG2 cells. They have also proved that HEIH exerts a tumor promoting effect on HCC development. Moreover, HEIH exhibits a high expression in various types of cancers, including colorectal cancer, melanoma, and lung cancer¹⁸⁻²⁰. Besides, the tumor-promoting effect of HEIH in these cancers has been confirmed. Although HEIH is proved to be associated with the pathogenesis of human cancers, its function in HCC has not been fully elaborated. Therefore, the aim of this study was to investigate the effect of HEIH on the growth and metastasis of human HCC tissues and to reveal the potential occurrence mechanism of HCC.

Patients and Methods

Experimental Materials

The investigation was approved by the Ethics Committee of Jiangyin Hospital Affiliated to Nanjing University. Signed written informed consents were obtained from all participants. The study selection of patients was based on the guideline proposed by the Union for International Cancer Control (UICC). A total of 12 patients aged 45-55 years old admitted to our hospital from April 2016 to January 2017 were enrolled in this study. Paired tumor samples were obtained from each patient. U6/Neo plasmid (GenePharma, Shanghai,

China), miRNeasy Mini Kit (Qiagen, Shenzhen, China), TB Green Premix Ex Taq II (Takara, Dalian, China), fluorescence-activated cell sorting (FACS) kit (Beckman, California, CA, USA), bicinchoninic acid (BCA) (Beijing Puli, Beijing, China) and 0.22 µm filter membrane (Millipore, Billerica, MA, USA).

Research Objects

The obtained 18 HCC tissue samples were assigned into HCC group and para-cancer group. HCC Huh7 and Hep3B cells were transfected with short hairpin (sh) HEIH plasmid (sh-HEIH group) and empty plasmid (sh-NC group). Thereafter, HCC Huh7 cells with silenced HEIH expression were selected as research objects and transfected with inhibitor reagent (sh-HEIH+inhibitor group) and negative control (Negative control (sh-HEIH+NC group)).

Cell Culture and Transfection

Huh7 and Hep3B cells were cultured in complete high-glucose Dulbecco Modified Eagle's Medium (DMEM). The sh structure of sh-HEIH was applied in U6/Neo plasmid, and empty U6/Neo plasmid sh-NC was regarded as a negative control. The sh-HEIH and sh-NC were designed and synthesized by GenePharma (Shanghai, China). Cell transfection was performed in 6-well plates according to the instructions of Lipofectamine 3000 (Invitrogen, Carlsbad, CA, USA). After 6 h, the transfection reagent was discarded, and the cells were collected for subsequent assays.

Western Blotting

Transfected cells were first digested with trypsin. Cell precipitates were collected and added with cell lysate, followed by ultrasonic disruption. The supernatant was then collected, and the concentration of extracted protein was determined in accordance with the BCA kit. Next, protein samples were separated *via* sodium dodecyl sulphate-polyacrylamide gel electrophoresis (SDS-PAGE) and transferred onto polyvinylidene difluoride (PVDF) membranes (Roche, Basel, Switzerland). After blocking with 5% skimmed milk powder for 10 min, the membranes were incubated with primary antibodies overnight at 4°C. On the next day, the membranes were incubated with corresponding secondary antibody at room temperature for 2 h. Immuno-reactive bands were finally exposed by enhanced electrochemiluminescence (ECL) method.

Determination of Cell Apoptosis

Transfected cells were collected and precipitated in Eppendorf (EP; Hamburg, Germany) tubes following transient trypsinization. After washing with cold phosphate-buffered saline (PBS), the cells were incubated in cell dye solution [5 µL of Annexin V-FITC (fluorescein isothiocyanate) and 5 µL of Propidium Iodide (PI)] and 95 µL of dye solution binding buffer for 10 min in a dark place. Finally, cell apoptosis was detected using a flow cytometer.

Detection of Cell Proliferation Via MTS

After digestion, transfected cells were uniformly inoculated into 96-well plates at a density of 5,000 cells per well, with a total volume of 200 µL. After incubation at 37°C for 48 h, 10 µL of Cell Counting Kit-8 (CCK-8) was added (Dojindo Molecular Technologies, Kumamoto, Japan) to each well, followed by incubation at 37°C for another 4 h in the dark. Absorbance at 490 nm in each cell was determined by a micro-plate reader.

Quantitative Real Time-Polymerase Chain Reaction (qRT-PCR)

Total RNAs in cells and tissues were extracted using TRIzol reagent (Invitrogen, Carlsbad, CA, USA). To analyze the expression of HEIH, PrimeScript RT Master Mix and Green Pre-mix Ex Taq II were used for complementary DNA (cDNA) synthesis and qPCR, respectively. In addition, messenger RNAs (mRNAs) in Huh7 cells were isolated using RNeasy Lysis Kit to examine the expression of miR-199a-3p. Mir-X miRNA one-strand synthesis kit and Mir-X miRNA qPCR SYBR kit were utilized for reverse transcription and qPCR. Glyceraldehyde 3-phosphate dehydrogenase (GAPDH) and U6 were used as internal controls for mRNA and miRNA, respectively. Expression level was calculated using the $2^{-\Delta\Delta Ct}$ method. The primer sequences are shown below: HEIH: Forward: 5'-GCGAGGCGCCATGAGACC-30; Reverse: 5'-AACCCTTGTGTGACCGA-3'; miR-199a-3p: Forward: 5'-ACACTCCAGCTGCAGTAGGACACAT-3'; Reverse: 5'-CAACTGGGTGTCGTGGAGTCGGCAATTCCTTAA CCAAT-3'; GAPDH: Forward: 5'-CTGGCTCCTCCTGTTCGAC-3'; Reverse: 5'-GACTCCGACCTTCACCTTCC-3'; U6: Forward: 5'-CTCGCTTCGGCAGCACACA-3'; Reverse: 5'-AACGCTTCACGAATTTGCGT-3'.

Wound Healing Assay

To determine the changes in relative migration, the healing changes after cell scratching were measured to reflect the changes of cell migration. In short, the cells were evenly inoculated into 6-well plates. When cell confluence reached 100%, the effect of cell proliferation was eliminated by replacing the original medium with serum-free medium. Three straight lines in each well were drawn using a 10 µL syringe head. The beginning of drawing was recorded as 0h, and images were continuously captured after incubation for 24 h.

Statistical Analysis

Statistical Product and Service Solutions (SPSS) 19.0 (SPSS Corp., Armonk, NY, USA) was used for all statistical analysis. The *t* test was used to compare the difference between two groups. $p < 0.05$ was considered statistically significant.

Results

HEIH Was Highly Expressed In HCC Tissues

The expression of HEIH in 18 pairs of HCC tissues and adjacent normal tissues was quantitatively analyzed by qRT-PCR. The results showed that HEIH expression in HCC tissues was markedly higher than that in para-cancerous tissues ($p < 0.01$) (Figure 1). This indicated that HEIH might be associated with the occurrence and development of HCC.

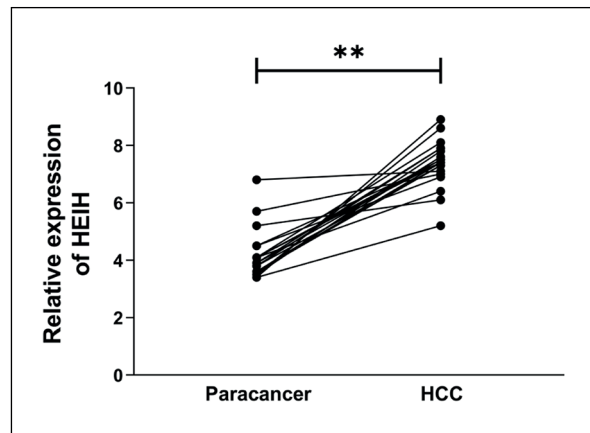


Figure 1. HEIH expression in HCC tissues and para-cancer tissues. HEIH is highly expressed in HCC tissues compared with para-cancer tissues ($p < 0.01$) (** $p < 0.01$).

HEIH Silencing Inhibited the Proliferation and Migration of Huh7 Cells

To testify the authenticity of the above hypothesis, HEIH was silenced in Huh7 cells by shRNA transfection. QRT-PCR data revealed that sh-HEIH transfection significantly reduced HEIH expression in HCC cells in comparison with NC transfection ($p < 0.01$). Next, the effects of HEIH silencing on the growth and metastasis of Huh7 cells were assessed by functional assays. According to MTS results, cell proliferation in sh-HEIH group was significantly inhibited ($p < 0.05$). In addition, in contrast to sh-NC group, sh-HEIH group exhibited remarkably shortened cell migration distance and lower wound healing rate ($p < 0.01$) (Figure 2).

Silencing of HEIH Suppressed Invasion but Triggered Apoptosis of Huh7 Cells

Western blotting manifested that the expressions of matrix metalloproteinase (MMP)-2, MMP-

3 and vimentin were overtly lower in sh-HEIH group than those in sh-NC group ($p < 0.05$), which indicated that suppressing HEIH expression promoted the invasion of HCC cells. Flow cytometry was then adopted to detect the percentage of apoptotic Huh7 cells with silenced HEIH expression. It was found that the percentage of apoptotic cells in sh-HEIH group was remarkably higher than that in sh-NC group ($p < 0.01$) (Figure 3). These findings suggested that the inhibition on HEIH expression induced the apoptosis of HCC cells.

HEIH Silencing Increases miR-199a-3p Expression

Cancer-associated miR-199a-3p has been widely recognized as a tumor suppressor miRNA in HCC^{21,22}. It shows strong ability to inhibit multiple biological processes, such as growth, migration, invasion and angiogenesis²³. In this study, the correlation between HEIH and miR-199a-3p was

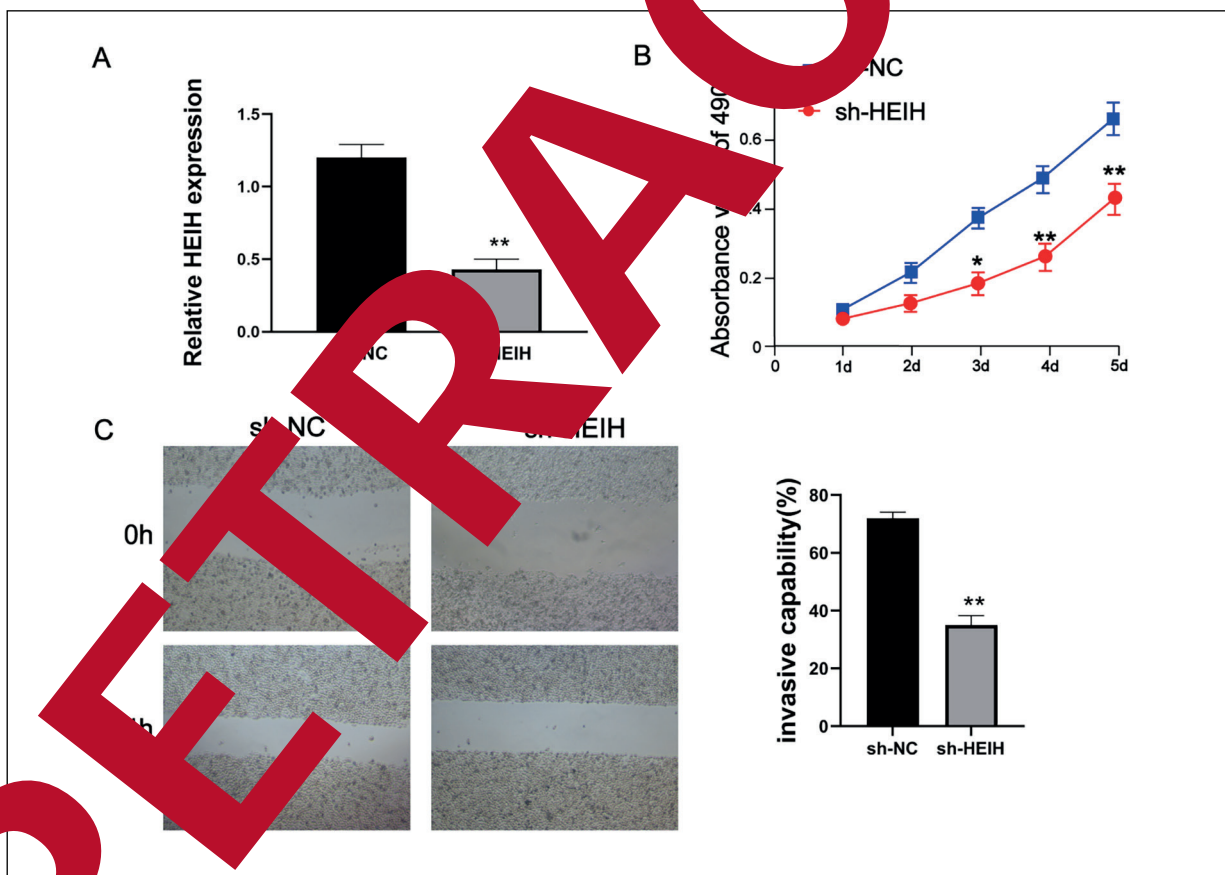


Figure 2. Effects of HEIH expression on HCC cell proliferation and migration. **A**, HEIH expression in Huh7 cells transfected with shRNA. HEIH is lowly expressed in sh-HEIH group compared with that in sh-NC group ($p < 0.01$). **B**, Effects of silenced HEIH expression on Huh7 cell proliferation. Absorbance at 490 nm of cells in sh-HEIH group is notably lower than that in sh-NC group ($p < 0.05$). **C**, Effects of silenced HEIH expression on the migration of Huh7 cells. Sh-HEIH group exhibits a prominently lower wound healing rate than sh-NC group (magnification: 40 \times) ($p < 0.01$) (* $p < 0.05$, ** $p < 0.01$).

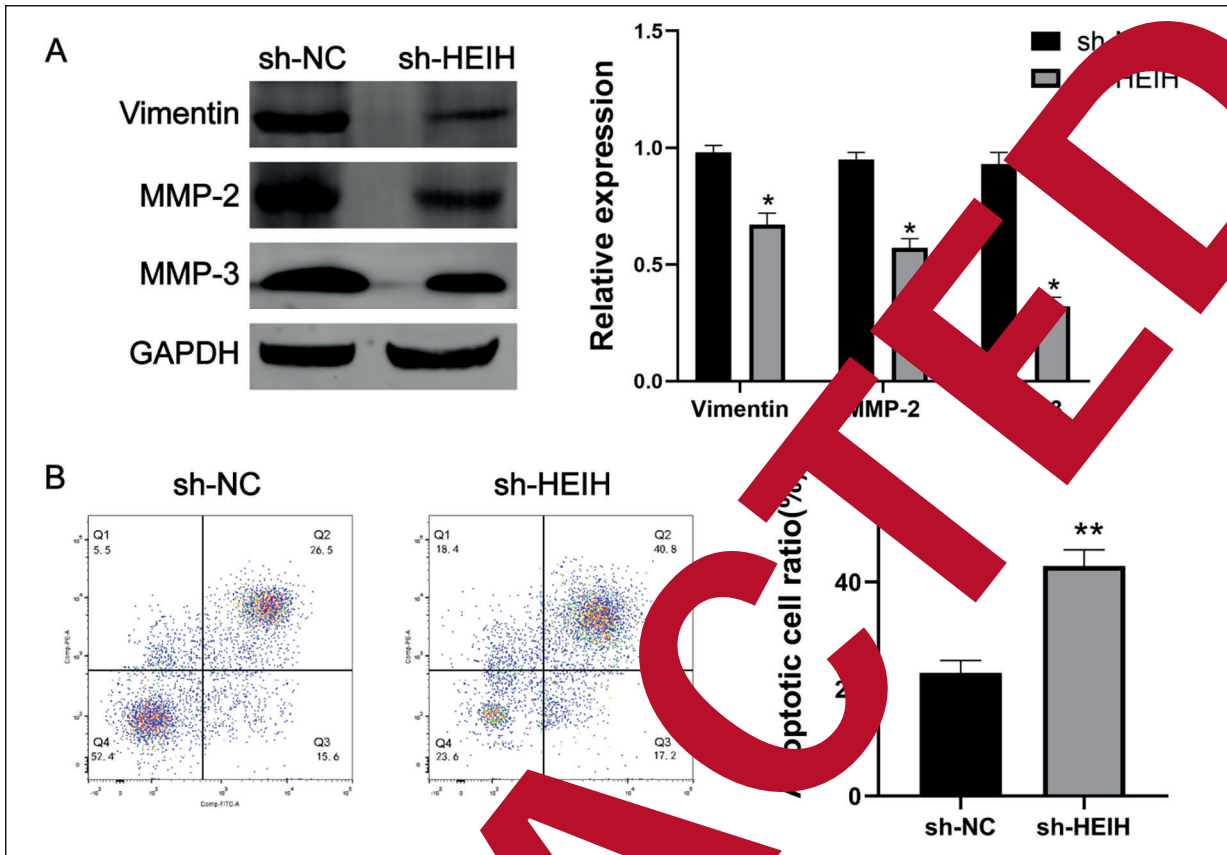


Figure 3. Impacts of suppressing HEIH expression on HCC invasion and apoptosis. **A**, Effect of HEIH on HCC cell invasion. In comparison with sh-NC group, sh-HEIH significantly reduced expressions of epithelial-mesenchymal transition (EMT) proteins ($p < 0.05$). **B**, Effect of HEIH expression on HCC cell apoptosis. The percentage of apoptotic cells in sh-HEIH group is markedly higher than sh-NC group ($p < 0.01$) (* $p < 0.05$, ** $p < 0.01$).

explored to explain the inhibitory effect of HEIH silencing on tumors. QRT-PCR analysis indicated that sh-HEIH significantly up-regulated the ex-

pression of miR-199a-3p through silencing HEIH expression in both Huh7 and Hep3B cells compared with sh-NC ($p < 0.01$) (Figure 4).

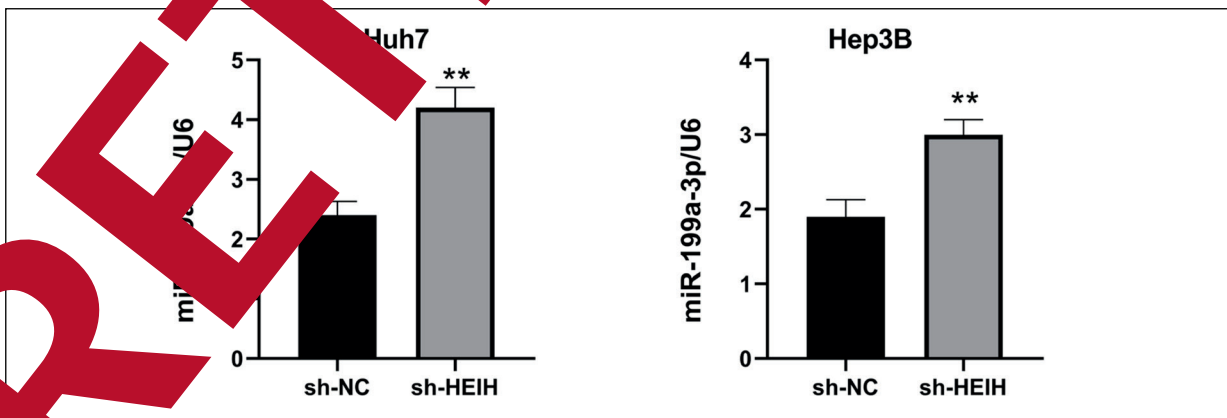


Figure 4. Effect of HEIH silencing on miR-199a-3p expression. In contrast to cells in sh-NC group, those in sh-HEIH group have a markedly up-regulated expression of miR-199a-3p ($p < 0.01$) (** $p < 0.01$).

HEIH Inhibited the Growth and Metastasis of Huh7 and Hep3B Cells by Modulating miR-199a-3p

HCC Huh7 cells were selected and transfected with miR-199a-3p inhibitor to inhibit miR-199a-3p expression, so as to further determine whether HEIH silencing up-regulated the expression of miR-199a-3p and was related to the function of HEIH. QRT-PCR results demonstrated that the expression of miR-199a-3p in Huh7 cells was indeed significantly inhibited after transfection of miR-199a-3p inhibitor reagent ($p < 0.01$). Subsequently, sh-NEIN and miR-199a-3p inhibitor were co-transfected into Huh7 cells, and the changes in cell growth and metastasis were reevaluated. Experimental results showed that co-transfection of sh-HEIH and miR-199a-3p inhibitor weakened the inhibition on cell proliferation by sh-HEIH, manifested as evidently higher absorbance at 450 nm in sh-HEIH+inhibitor group than sh-HEIH+NC group ($p < 0.05$). In addition, co-transfection with sh-HEIH and miR-199a-3p inhibitors weakened

the inhibitory effect of sh-HEIH on cell migration as well. Wound healing rate in sh-HEIH+inhibitor group was overtly higher than that in sh-HEIH+NC group ($p < 0.05$). To summarize, the growth and metastasis of Huh7 cells were suppressed by HEIH silencing, and this effect was mediated by regulating the expression of miR-199a-3p after silencing HEIH expression. For another HCC cell line Hep3B, a similar trend was observed. HEIH silencing prominently suppressed the proliferation and migration of Hep3B cells ($p < 0.05$). When the expression of miR-199a-3p was inhibited, the anti-growth and anti-metastasis effects of HEIH silencing on Hep3B cells were weakened. All our findings demonstrated that HEIH silencing inhibited the growth and metastasis of Hep3B cells by regulating miR-199a-3p (Figure 5).

Discussion

Currently, the overall survival rate of HCC patients is relatively low. Meanwhile, high metastasis

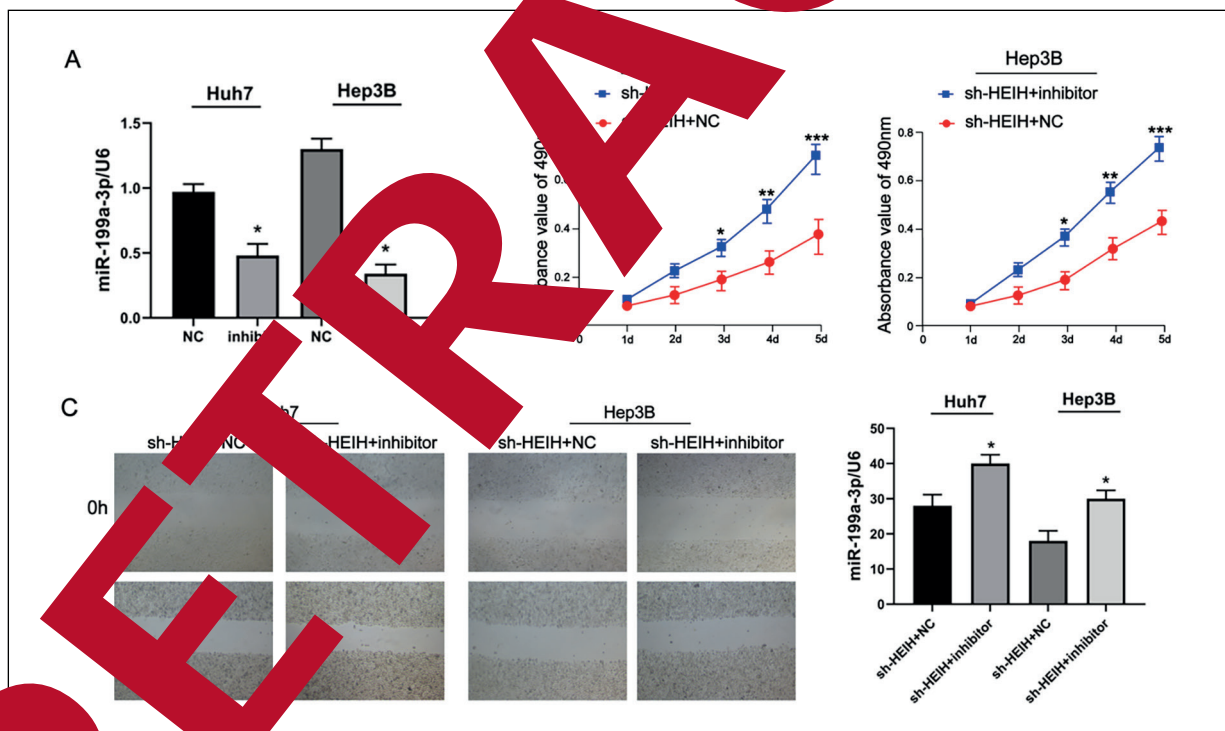


Figure 5. Impacts of HEIH silencing and miR-199a-3p inhibition on the growth and metastasis of Huh7 cells. **A**, Expression of miR-199a-3p in Huh7 and Hep3B cells transfected with miR-199a-3p inhibitor or NC. It is discovered that compared with sh-HEIH+NC group, the expression of miR-199a-3p in sh-HEIH+inhibitor group evidently declines ($p < 0.05$). **B**, Changes in the proliferation of the two cell lines co-transfected with shRNA and miR-199a-3p inhibitor. In contrast to cells in sh-HEIH+NC group, those in sh-HEIH+inhibitor group display a significantly increased absorbance at 450 nm ($p < 0.01$). **C**, Changes in cell migration after co-transfection with shRNA and miR-199a-3p inhibitor. Wound healing rate of cells in sh-HEIH+inhibitor group is significantly higher than that in sh-HEIH+NC group (magnification: 40×) ($p < 0.05$) (* $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$).

and recurrence rates become huge obstacles to the treatment of HCC. Specific lncRNAs exert regulatory roles in the growth, apoptosis, migration and invasion of HCC cells¹⁵. In this study, we mainly aimed to explore the effect of HEIH on HCC. The difference in HEIH expression in 18 pairs of HCC tissues and para-cancer tissues was detected by qRT-PCR. The results showed that HEIH was highly expressed in HCC tissues compared with para-cancer tissues, which is consistent with the findings of previous studies^{17,19}. It is suggested that HEIH level increases in tumor microenvironment. To further determine whether elevated HEIH participates in regulating the growth and metastasis of HCC cells, human HCC cell lines (Huh7 and Hep3B) were transfected with shRNA to silence HEIH expression. Huh7 cell line with silenced HEIH expression was selected for subsequent functional assays. According to the results, HEIH silencing notably reduced the proliferation and migration of Huh7 cells but induced their apoptosis. Of note, miR-199a-3p was highly expressed in HCC cells with silenced HEIH expression. Therefore, it was regarded as a downstream target molecule of HEIH in HCC. In the case of increased miR-199a-3p expression, the functions of Huh7 and Hep3B cells were weakened.

Up to date, there are few studies on the tumor-promoting effect of HEIH on human cancer. Overexpression of HEIH has been confirmed to promote the migration and invasion of tumor cells, which may also increase the mortality of non-small cell lung cancer A549 and Hep-1 cells. Similar results have been observed in rectal cancer and melanoma cells^{18,19}. However, there are no reports exist in the function of HEIH in HCC. Yang et al¹⁶ have initially proved that HEIH is able to modulate the cell cycle and proliferation of HCC cells, indicating the involvement of HEIH in regulating the proliferation of HCC cells. Highly expressed HEIH facilitates the invasion of HCC MHC cells but does not influence its proliferation ability¹⁷. In addition, epithelial-mesenchymal transition (EMT) is an important process in the progression and metastasis of cancer. In this study, the results of western blot showed that the expressions of matrix metalloproteinase (MMP-2, MMP-3 and vimentin) were significantly lower in sh-HEIH group than those in sh-NC group. All these results suggested that after silencing the expression of HEIH in Huh7 and Hep3B cells, cell proliferation and migration were significantly inhibited. However, cell apoptosis was induced. The findings of this study are partially consistent with those in previous researches,

illustrating that HEIH is conducive to the proliferation and metastasis of HCC cells. All the above data imply that HEIH silencing may be a potential therapeutic target for HCC treatment.

LncRNAs have a very complex functional mechanism. Physically, lncRNAs can combine with DNAs, proteins, mRNAs and miRNAs to modulate the expression, localization and stability of binding partners²¹. Among these different mechanisms of action, there are increasingly more interactions between lncRNAs and miRNAs. Meanwhile, lncRNAs can knock down miRNAs and mRNAs like molecular sponges. In the present study, the co-linking between HEIH and miR-199a-3p was proved. MiR-199a-3p ranks third among miRNAs with the highest expression in the normal liver, which is remarkably down-regulated in HCC. Besides, miR-199a-3p exerts a crucial effect in driving the occurrence and development of HCC. In recent years, it becomes one of the most extensively researched miRNAs in HCC²³. In this study, the expression changes of miR-199a-3p in HCC cells with lowly expressed HEIH were detected. qRT-PCR results demonstrated that miR-199a-3p was highly expressed in HCC cells with low expression of HEIH, suggesting that miR-199a-3p expression is negatively regulated by HEIH. Subsequent functional analysis manifested that when miR-199a-3p expression was inhibited, the tumor-promoting effect of HEIH silencing on Huh7 and Hep3B cells was weakened. The above results prove that HEIH functions in the growth and metastasis of HCC cells, which may be partially realized by negative regulation on miR-199a-3p.

Conclusions

The novelty of this study was that HEIH silencing may be a promising target for inhibiting HCC cell growth and metastasis. In addition, HEIH silencing exerts its anti-tumor impact mainly by up-regulating miR-199a-3p. However, the results of this study still need to be verified *in vivo*.

Conflict of Interest

The Authors declare that they have no conflict of interests.

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