

Regulation by Pink1 on the mitochondrial dysfunction in endothelial cells post the hypoxia mimetic agent CoCl₂ treatment

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Abstract. – OBJECTIVE: To explore the role of miR-451a in the migration and invasion of non-small cell lung cancer (NSCLC) cells.

MATERIALS AND METHODS: Quantitative Real time-polymerase chain reaction (qRT-PCR) and Western blot were performed to detect the levels of miR-451a and activating transcription factor 2 (ATF2) in NSCLC. Transwell assay was employed to analyze the migratory and invasive abilities in NSCLC cells. Dual-luciferase reporter assay was applied to confirm the binding condition of miR-451a and its target gene in NSCLC cells.

RESULTS: MiR-451a was downregulated in NSCLC tissues and lung cancer cell lines A549 and NCI-H460, while ATF2 was upregulated. The mRNA level of miR-451a was negatively correlated with ATF2. Additionally, miR-451a regulated cell migration and invasion through targeting ATF2. Furthermore, ATF2 could reverse the inhibitory effect of miR-451a on migration and invasion of A549 cells induced by miR-451a.

CONCLUSIONS: MiR-451a inhibited the migratory and invasive abilities of NSCLC cells through ATF2 regulation. The miR-451a/ATF2 axis provides a new insight into the pathogenesis of NSCLC.

Key Words: miR-451a, Migration, Invasion, ATF2, Non-small cell lung cancer

Introduction

Lung cancer is the most common malignant tumor with the highest mortality rate^{1,2}. Among all the diagnostic cases of the lung cancer, approximately 85% were non-small cell lung cancer (NSCLC), including adenocarcinoma and squamous cell carcinoma^{3,4}. Most NSCLC patients were diagnosed at advanced stage and had

a poor prognosis. Therefore, it is necessary to find biological markers for early diagnosis and survival prediction of NSCLC.

MicroRNAs (miRNAs), with a length of 22-28 nucleotides, could induce mRNA degradation or inhibit gene expression through binding to 3' UTR of target mRNA^{5,6}. Evidence has revealed that miRNAs participate in almost 60% of all human genes at post-transcriptional level^{7,8}.

MiR-451a plays a crucial role in the occurrence and development in various tumors, including pancreatic ductal adenocarcinoma, renal cell carcinoma, gastric cancer and dilated cardiomyopathy⁹⁻¹².

MiR-451a could abrogate the treatment resistance in FLT3-ITD-positive acute myeloid leukemia¹³.

Moreover, miR-451a is acted as a noninvasive biomarker for early prediction of recurrence and prognosis of pancreatic ductal adenocarcinoma¹⁴.

Therefore, we strongly believed that miR-451a may play great roles in NSCLC. Activating transcription factor 2 (ATF2) belongs to the cAMP response element binding family, which is activated by c-Jun N-terminal kinases phosphorylation and responses to regulatory biological progresses^{15,16}.

ATF2 activates the transcription of genes by dimer formation with other members of the activator protein superfamily and stimulates cytokine productions in response to DNA damage and cell death^{17,18}. ATF2, a paradigm of the multifaceted regulation of transcription factors, is involved in glucose metabolism and tumorigenesis^{19,20}.

In addition, ATF2 is associated with the malignant phenotypes and poor prognosis of ovarian adenocarcinoma patients and renal cell carcinoma^{21,22}. Considering these functions, we proposed that miR-451a could regulate cell migration and invasion in NSCLC via regulating ATF2 expression.

Materials and Methods

Tissue Specimens

In accordance to WHO classification, 55 paired lung carcinoma and paracancerous tissues were collected from the Affiliated Yantai Yuhuangding Hospital of Qingdao University, from 2015 to 2017. All the specimens were surgically resected and immediately stored at -80°C before RNA extraction. Patients did not receive preoperative therapies, including radiotherapy and chemotherapy. This study was approved by the Ethical Committee of Affiliated Yantai Yuhuangding Hospital of Qingdao University. All patients signed informed consent.

Cell Culture

We obtained two human lung cancer cell lines A549 and NCI-H460 and normal lung cells MRC-5 from American Type Culture Collection (ATCC, Rockville, MD, USA). All cells were maintained in Roswell Park Memorial Institute-1640 (RPMI-1640) medium (Gibco, Carlsbad, CA, USA) with 10% fetal bovine serum (FBS, Sigma-Aldrich, St. Louis, MO, USA) cultured at 37°C with 5% CO₂.

Transfection

MiR-451a mimic, miR-451a inhibitor, pCDNA3.1-ATF2 and control vector were constructed. A549 cells were seeded into 6-well plates and cultivated overnight before transfection. The plasmid vectors were transfected into A549 cells using Lipofectamine 2000 Reagent (Invitrogen, Carlsbad, CA, USA).

RNA Isolation and Quantification by Real-Time-Polymerase Chain Reaction (RT-PCR)

TRIzol Reagent (Invitrogen, Carlsbad, CA, USA) or MIRcute Extraction and Separation of miRNAs Kit (Tiangen, Beijing, China) were utilized to isolate and extract total RNAs and miRNAs, respectively. To detect the expression of miRNA or mRNA, PrimeScript II 1st Strand complementary Deoxyribose Nucleic Acid (cDNA) Synthesis Kit (TaKaRa, Otsu, Shiga, Japan) was conducted to reversely transcribe total RNA into complementary Deoxyribose Nucleic Acid (cDNA) chips. Then, SYBR Premix kit (TaKaRa, Otsu, Shiga, Japan) was applied to perform the RT-qPCR. Glyceraldehyde 3-phosphate dehydrogenase (GAPDH) and U6 were used as the loading controls. The relative expression of miRNA and mRNA were calculated through the $2^{-\Delta\Delta Ct}$ method. The primer sequences were as the following: MiR-451a: F: 5'-ACACTCCAGCTGG-

GAAACCGTTACCATTACT-3', R: 5'-CTGGT-GTCGTGGAGTCGGCAA-3'; ATF2: F: 5'-GATCCATGAAATTCAAGTTACAT-3', R: 5'-GGCTCGAGTCAACTTCCTGAGGCTGTG-3'; U6: F: 5'-GCTTCGGCAGCAATACTAAA-AT-3', R: 5'-CGCTTCAGAATTTCATCAT-3'; GAPDH: F: 5'-CGCTCTCTCTCCCTTCTTTC-3', R: 5'-ATCCGTTGACTCCACCTTCAC-

Protein Extraction and Western Blotting

Radioimmunoprecipitation assay (RIPA) Lysis Buffer supplemented with phenylmethanesulfonyl fluoride (PMSF) (both from Beyotime, Beijing, China) were employed to extract the total proteins from lung cancer cells. After centrifugation, protein concentration was measured by bicinchoninic acid (BCA) reagent kit (Sangon, Beijing, China). After electrophoresis, the separated proteins were transferred to the polyvinylidene difluoride (PVDF) membrane (Millipore, Billerica, MA, USA). The membrane was incubated with rabbit anti-ATF2 monoclonal antibody (1:1000; Abcam, Cambridge, MA, USA) at 4°C overnight. A rabbit secondary antibody (1:1000; ZSGB-BIO, Beijing, China) containing horseradish peroxidase (HRP)-conjugated was used to incubate the membrane for 1 h at room temperature. Anti-β-actin mouse antibody (1:3000; ZSGB-BIO, Beijing, China) was used as internal control. The interest proteins were performed by electrochemiluminescence (ECL) Western Blotting Detection System (BestBio, Beijing, China).

Transwell Assay

Transwell chamber (8 μm in pore size, Costar, Beijing, China) was pre-coated with Matrigel (Clontech, Mountain View, CA, USA). A549 cells were prepared for cell suspension. 200 μL of cell suspension in serum-free medium were added in the upper chamber. Meanwhile, 500 μL of normal medium containing 15% FBS were added in the lower chamber. 24 h later, non-adherent cells were removed using cotton swab cautiously. Cells were fixed with methanol and stained with crystal violet. Cell counting was then carried out through the microscope.

Plasmid Construction and Luciferase Reporter Assay

TargetScan software (www.targetscan.org) was used to predict target genes of miR-451a; moreover, ATF2 was screened out. Plasmids with wild-type and mutant-type 3'UTR oligonucleotides fragment inserting in pmirGlvectors of ATF2 were constructed (pmirGlo-ATF2-WT, pmirGlo-ATF2-MUT). Plasmid efficacy was verified

by sequencing. After co-transfection of miR-451a mimic or negative control (NC) and WT or MUT into A549 cells, the dual-luciferase activity was detected using Dual-Luciferase Reporter Assay System (Promega, Madison, WI, USA) with Renilla luciferase as normalization.

Statistical Analysis

Statistical analyses were performed using software Statistical Product and Service Solutions (SPSS) 19.0 (SPSS Inc., Chicago, IL, USA). Measurement data were compared by Student's *t*-test. The differences among groups were compared by one-way ANOVA, followed by LSD post-hoc test. $p < 0.05$ indicated that the results were statistically significant.

Results

Correlation of miR-451a and ATF2 in NSCLC Tissues

To explore the function of miR-451a in NSCLC, the expression level of miR-451a was measured in

55 pairs of NSCLC tissues using qRT-PCR. The data showed that miR-451a was lower in NSCLC tissues compared with corresponding adjacent normal tissues ($p < 0.001$, Figure 1A). In addition, we also determined the mRNA level of ATF2 and it was higher in NSCLC tissues than that in corresponding adjacent normal tissues ($p < 0.0001$, Figure 1B). Correlation analysis showed that miR-451a negatively correlated with ATF2 in NSCLC tissues ($p = 0.001$, $r = -0.4252$, Figure 1C). Besides, miR-451a was downregulated in NSCLC cells A549 ($p = 0.0005$) and H1299 ($p = 0.0004$) compared with that of normal lung epithelium cell HREC-5 (Figure 1D).

MiR-451a Inhibits Migration and Invasion of NSCLC Cells

To determine the function of miR-451a in NSCLC, the migratory and invasive abilities were evaluated. Transfection efficacies of miR-451a mimic and miR-451a inhibitor in A549 cells were verified by qRT-PCR. As expected, mRNA level of miR-451a was incre-

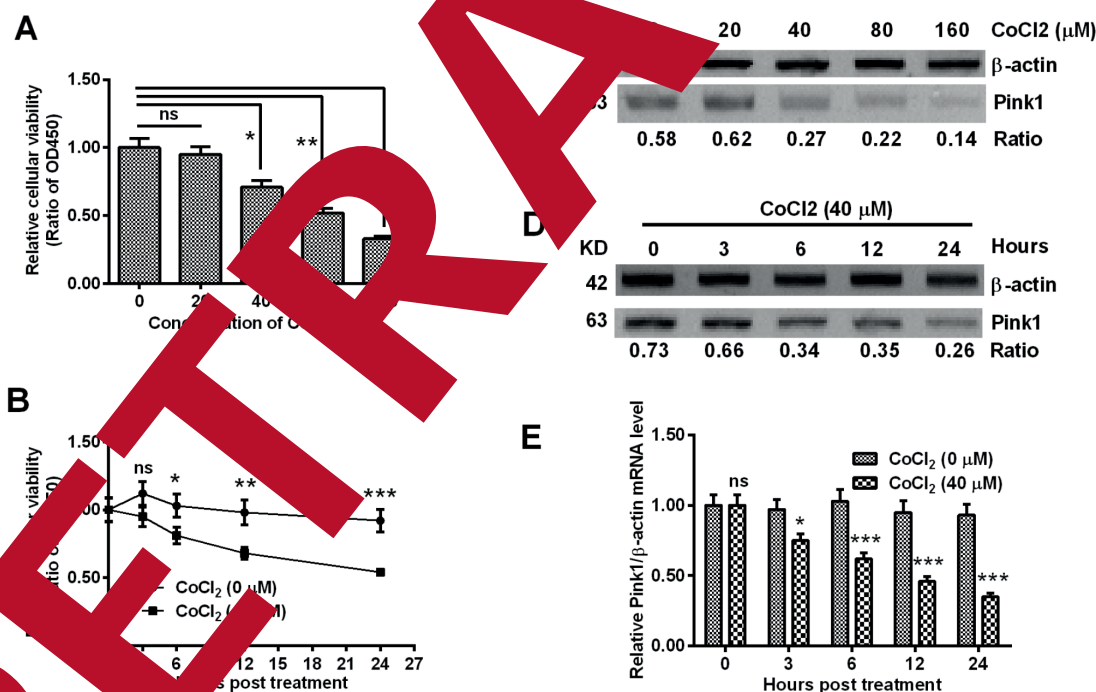


Figure 1. Regulation by cobalt chloride (CoCl_2) on cell viability and Pink1 expression in human endothelial EA.hy926 cells. Cells were treated with MTT assay for human endothelial EA.hy926 cells, which were treated with 20, 40, 80 or 160 μM CoCl_2 for 12 hours (A), with 40 μM CoCl_2 for 0, 3, 6, 12 or 24 hours (B). Western blotting assay was performed to quantified Pink1 expression in protein level in the EA.hy926 cells, which were treated with 20, 40, 80 or 160 μM CoCl_2 for 12 hours (C) and with 40 μM CoCl_2 for 0, 3, 6, 12 or 24 hours (D). Real-time quantitative PCR was performed to examine the relative mRNA level of Pink1 to β -actin in the EA.hy926 cells, which were treated with 40 μM CoCl_2 for 0, 3, 6, 12 or 24 hours (E). Experiments were performed independently for triplicate. * $p < 0.05$, ** $p < 0.01$ or *** $p < 0.001$, ns: no significance.

ased ($p=0.0013$) after transfection of miR-451a mimic, which was reduced ($p=0.0080$) after transfection of miR-451a inhibitor in A549 cells (Figure 2A). Subsequently, the migratory and invasive cell numbers were reduced ($p=0.0015$ and 0.0035) when A549 cells were transfected

miR-451a mimic. On the contrary, the migratory and invasive cell numbers were increased after transfection of miR-451a inhibitor ($p=0.00114$ and 0.0043 , Figure 2B). The above results indicated that miR-451a inhibits migration and invasion of NSCLC cells.

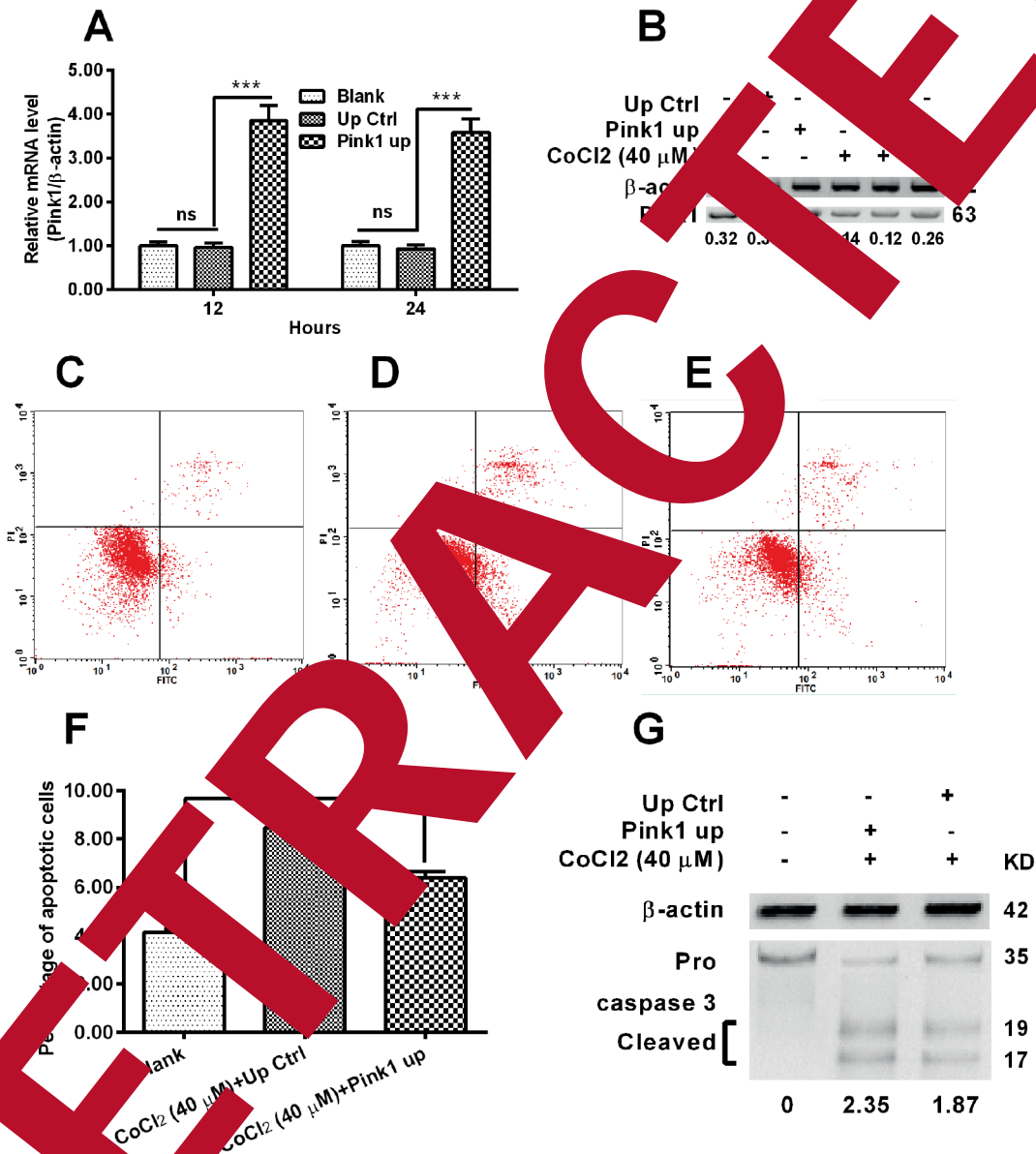


Figure 2. Apoptosis of EA.hy926 cells post the CoCl₂ treatment and the Pink1 overexpression. **A** and **B**, Relative Pink1 to β -actin mRNA level for or Western blotting assay of Pink1 (**B**) in the blank or the CoCl₂-treated (40 μ M) EA.hy926 cells, which were transfected with CAT-pcDNA3.1(+) (Up Ctrl), with Pink1-pcDNA3.1(+) (Pink1 up). **C-E**, Apoptosis (Flow cytometry analysis) of blank (**C**) or the CoCl₂-treated EA.hy926 cells, which were pre-transfected with CAT-pcDNA3.1(+) (**D**), with Pink1-pcDNA3.1(+) (**E**) for 12 hours. **F**, Apoptosis quantification of blank or the CoCl₂-treated EA.hy926 cells, which were pre-transfected with CAT-pcDNA3.1(+) (CoCl₂(40 μ M)+Up Ctrl), with Pink1-pcDNA3.1(+) (CoCl₂(40 μ M)+Pink1 up) for 12 hours. Early apoptotic cells were denoted in the upper right quadrant. **G**: Western blotting assay of cleaved caspase 3 in the three groups of EA.hy926 cells (blank, CoCl₂(40 μ M)+Up Ctrl and CoCl₂(40 μ M)+Pink1 up). Experiments were performed independently for triplicate. * $p < 0.05$ or ** $p < 0.01$.

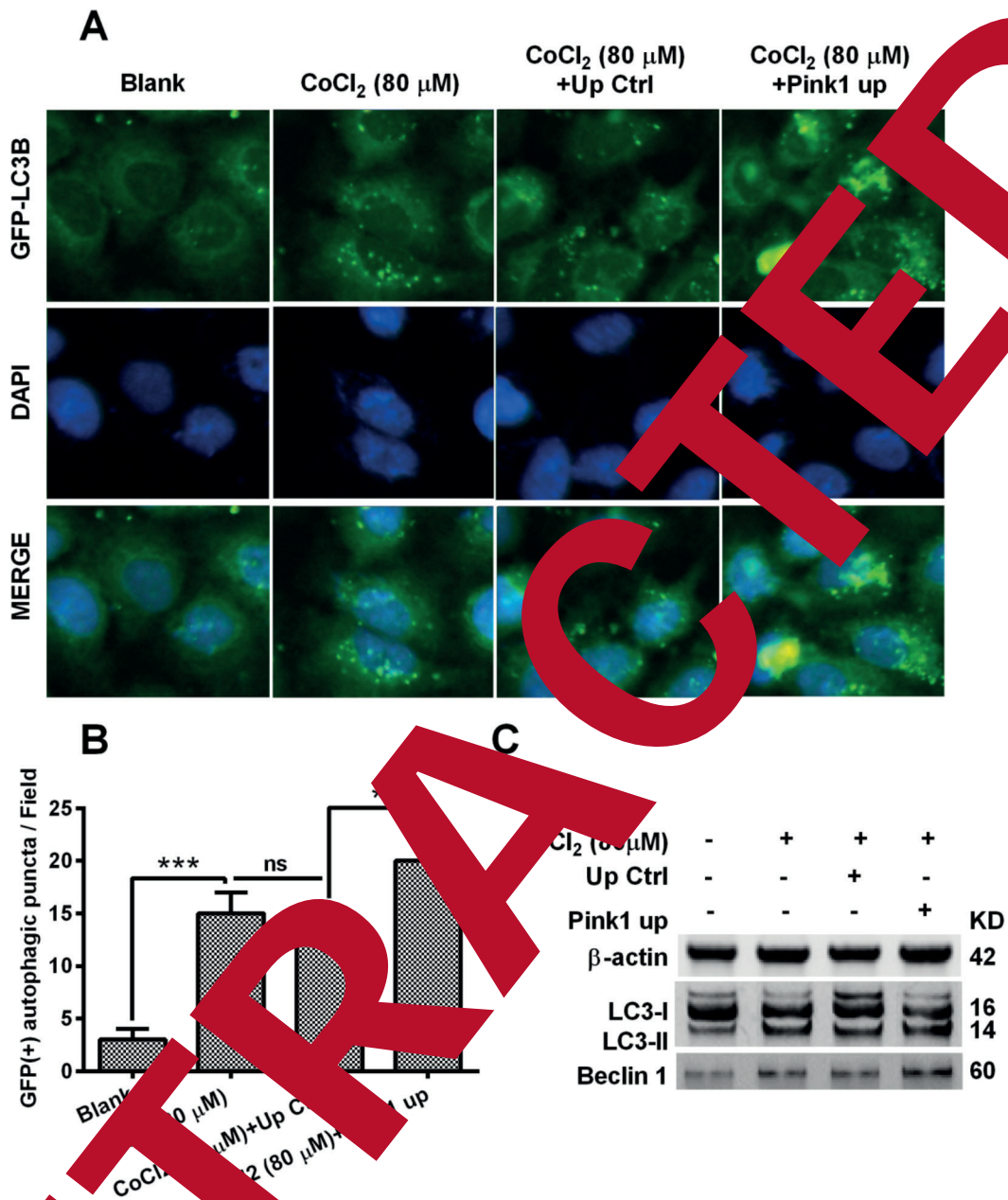


Figure 3 Autophagy in the EA.hy926 cells post the CoCl₂ treatment and the Pink1 overexpression. **A**, Autophagic vesicles were indicated as green fluorescence-positive puncta in the four groups of EA.hy926 cells (blank, CoCl₂ treatment with 80 μM for 6 hours), CoCl₂ treatment and Up Ctrl, CoCl₂ treatment and Pink1 up). **B**, Counting of autophagic vesicles in the four groups of EA.hy926 cells. **C**, Western blotting of autophagy-related proteins, LC3 and Beclin1 in the four groups of EA.hy926 cells; Experiments were performed independently for triplicate. **p* < 0.05, ****p* < 0.001, ns no significance.

miR-451a Targets ATF2 and Inhibits its

ATF2 was predicted to be a target gene of miR-451a by TargetScan and the binding site was located at 2960-2967 in the mRNA of ATF2 3'-UTR. To verify whether miR-451a could direct target to

ATF2, the binding pmirGlo-ATF2-WT (WT) and pmirGlo-ATF2-MUT (MUT) were constructed, respectively (Figure 3A). Subsequently, WT or MUT and miR-451a mimic were co-transfected into A549 cells and then luciferase abilities were detected. As expected, the luciferase activity was

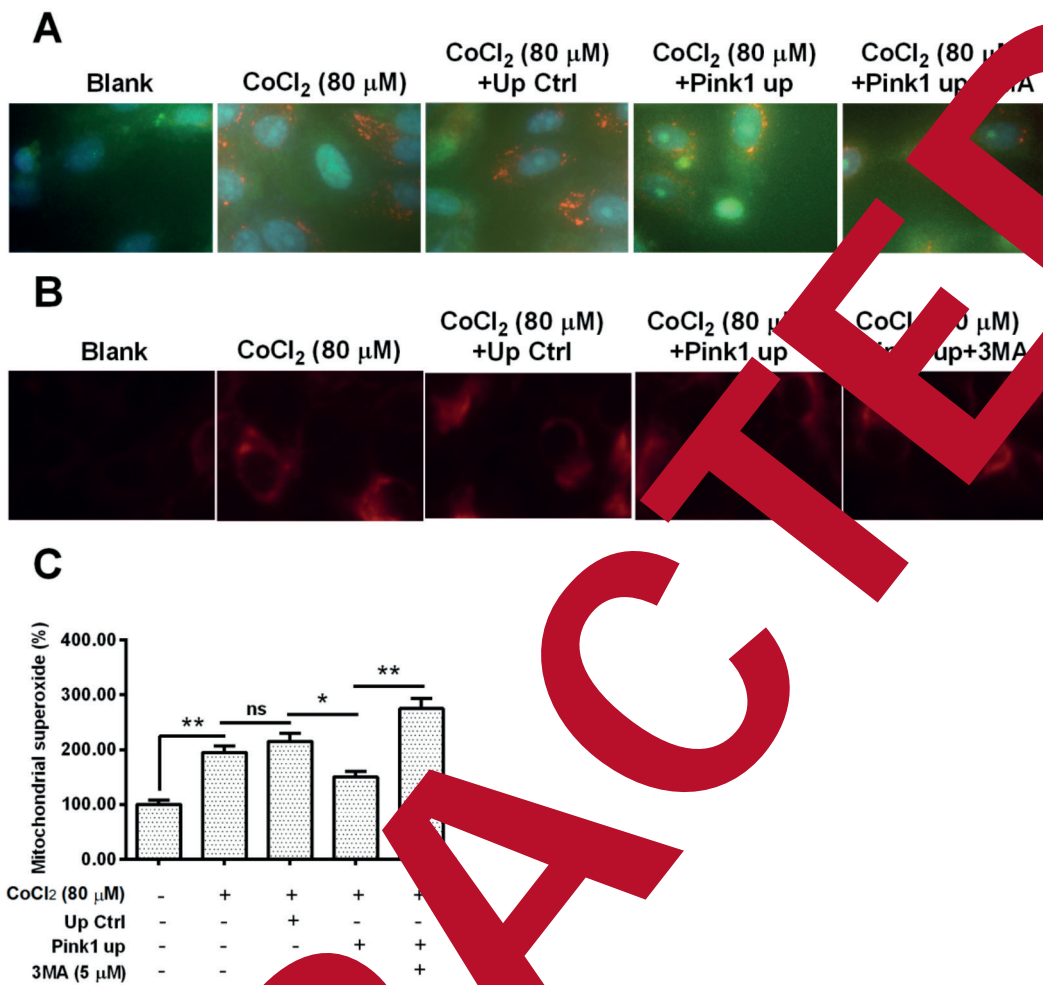


Figure 4. Regulation by Pink1 expression of the CoCl₂-induced mitochondrial dysfunction in EA.hy926 cells. EA.hy926 cells were transfected with pcDNA3.1(+)-Up Ctrl, with Pink1-pcDNA3.1(+) (Pink1 up) for 12 hours, were treated with CoCl₂ (80 μM) and 3MA (0 or 5 μM) and blank cells as blank control, then the JC-1 staining for mitochondrial membrane potential (MMP) (A), the fluorescence 5-(and-6)-chloromethyl-2,7-dichlorodi-hydrofluorescein diacetate (DCFH-DA) staining for mitochondrial reactive oxygen species (ROS) (B) and the MitoSOXTM Red staining for mitochondrial superoxide (C) was performed. All experiments were performed independently in triplicate. **p*< 0.05 or ***p*< 0.01, ns: no significance.

reduced (*p*=0.0011) in cells co-transfected with WT and miR-451a, while the kinase activity of MURK1 showed no alteration (*p*=0.7717) in A549 cells (Figure 3B). To further determine whether miR-451a mediated ATF2, ATF2 expression was evaluated after exogenous alteration of miR-451a. As expected, mRNA level of ATF2 was repressed (*p*=0.0030) when cells were transfected with miR-451a mimic, which was increased (*p*=0.0017) after transfection with miR-451a inhibitor (Figure 3C).

ATF2 could Reverse Partial Effects of miR-451a on Cell Migration and Invasion

To further explore the mechanism of miR-99a in regulating migration and invasion through ATF2,

miR-99a mimic and pcDNA3.1-ATF2 were co-transfected into A549 cells. The mRNA level of ATF2 was decreased (*p*=0.0003) when transfected with miR-451a mimic and it was reversed after transfection of pcDNA3.1-ATF2 (*p*=0.089, Figure 4A). The protein level of ATF2 showed the same results with that of mRNA level (Figure 4B). Transwell assay revealed that the migratory number was increased (*p*=0.0224) when co-transfection with miR-451a mimic and pcDNA3.1-ATF2, which was reversed by miR-451a mimic transfection (Figure 4C). Similarly with migration results, the invasive ability was also enhanced (*p*=0.0199) when co-transfection with miR-451a mimic and ATF2 was compared with those only transfected with miR-451a mimic (Figure 4D).

Discussion

Non-small cell lung cancer accounts for 85% of lung cancer, including adenocarcinoma and squamous cell carcinoma^{3,4}. Most NSCLC patients are diagnosed at advanced stage and had a poor prognosis. Thus, it is necessary to find biological markers for the early diagnosis and survival prediction of NSCLC.

MicroRNAs (miRNAs) could induce mRNA degradation or inhibit gene expression through binding to 3'-UTR of target mRNA. MicroRNAs participate in almost 60% of all human genes at post-transcriptional level^{5,6}. MiR-451a is served as a noninvasive biomarker and it could alleviate drug resistance in miscellaneous cancers^{13,4}. Liu et al^{23,24} elucidated that miR-451a inhibited cell proliferation and enhanced tamoxifen sensitive in breast cancer. More importantly, miR-451a acted as a tumor suppressor by retarding cell migration and invasion in melanoma²⁵. Our results were consistent with the previous findings that miR-451a was downregulated in NSCLC tissues and lung cancer cells. In lung cancer cells A549, miR-451a overexpression suppressed cell migration and invasion while knockdown of miR-451a obtained the opposite results. We first proposed that miR-451a regulated cell migration and invasion through targeting ATF2.

ATF2 is involved in multiple regulatory biological progresses, including transcription, DNA damage, metabolism and tumorigenesis^{18,19}. Increasing evidences have revealed that ATF2 was a target of several miRNAs, including miR-26b, miR-204, miR-622 and miR-451a²⁶⁻²⁹. For our study in our research, we found the same results that ATF2 was a target of miR-451a and was regulated by miR-451a in lung cancer cells A549. We first proposed the correlation between miR-451a and ATF2 in NSCLC. In non-small cell carcinoma, ATF2 knockdown promoted anticancer activity of sorafenib³⁰. Similar findings were discovered by Li et al³⁰, and they indicated that ATF2 knockdown suppress cell growth and enhances sensitivity to chemotherapy in pancreatic cancer. Consistent with the previous results, we first verified that miR-451a regulated migration and invasion of A549 cells through regulating ATF2 expression.

Conclusions

Our study showed that miR-451a was downregulated in non-small cell lung cancer tissues and lung

cancer cells A549 and NCI-H460, while ATF2 was upregulated. The mRNA level of ATF2 had negative correlation with ATF2. MiR-451a regulated the migration and invasion of lung cancer cells through targeting ATF2. In addition, ATF2 could reverse partial function of miR-451a on migration and invasion of A549 cells.

Conflict of Interest

The Authors declare that they have no conflict of interest.

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