

# Long non-coding RNA LINP1 induces tumorigenesis of Wilms' tumor by affecting Wnt/ $\beta$ -catenin signaling pathway

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**Abstract.** – **OBJECTIVE:** Recent studies have discovered that long non-coding RNAs (lncRNAs) play an important role in the development of malignant tumors. The aim of this work was to investigate the exact role of lncRNA LINP1 in the development of Wilms' tumor and to explore the possible underlying mechanism.

**PATIENTS AND METHODS:** The expression of lncRNA in non-homologous end joining pathway 1 (LINP1) in tissue samples of Wilms' tumor was detected by Real Time-quantitative Polymerase Chain Reaction (RT-qPCR). The relationship between the expression of lung cancer associated transcript 1 (LUCAT1) and patients' overall survival time was analyzed. Subsequent functional experiments were conducted to identify the changes in biological behavior of Wilms' tumor cells after the gain or loss of LINP1. Moreover, the underlying mechanism of LINP1 function was explored.

**RESULTS:** QRT-PCR results showed that LINP1 expression level in Wilms' tumor tissues was significantly higher than that in adjacent tissues. LINP1 expression was negatively associated with the overall survival time of patients with Wilms' tumor. Cell growth ability was markedly inhibited and promoted after down-regulation and overexpression of LINP1 *in vitro*, respectively. Moreover, after the loss and gain of LINP1 *in vitro*, cell migration and invasion abilities were markedly suppressed and promoted, respectively. Furthermore, the loss of LINP1 *in vitro* could significantly decrease the expression of target proteins in the Wnt/ $\beta$ -catenin signaling pathway. However, the expressions of target proteins in the Wnt/ $\beta$ -catenin signaling pathway were remarkably up-regulated after over-expression of LINP1.

**CONCLUSIONS:** LINP1 could enhance cell proliferation and proliferation via inducing the Wnt/ $\beta$ -catenin signaling pathway. Our findings might provide a new prospect for the diagnosis and therapy of Wilms' tumor.

**Key Words:**

Long non-coding RNA, LINP1, Wilms' tumor, Wnt/ $\beta$ -catenin signaling pathway.

## Introduction

Wilms' tumor is the most frequent pediatric renal cancer, which affects one in 10,000 children annually. The overall survival rate of Wilms' tumor is more than 90%<sup>1</sup>. When embryonic nephroblasts fail to undergo terminal differentiation, Wilms' tumor may happen. Great advances have been achieved in combination therapy to improve the prognosis of most patients. However, almost 10% of patients with Wilms' tumor eventually develop metastasis and recurrence, contributing to poor prognosis<sup>2,3</sup>. Thus, it is crucial to uncover the molecular mechanism of the progression of Wilms' tumors and to identify potential targets to improve the prognosis of this pediatric disease.

As a subtype of non-coding RNA (ncRNA), long noncoding RNAs (lncRNAs) regulate a variety of cellular processes and pathways in cancer development. For instance, the expression level of lncRNA-CCHE1 is positively correlated with the malignancy of colorectal carcinoma, which regulates the ERK/COX-2 pathway<sup>4</sup>. Through regulation of OIP5 expression, lncRNA OIP5-AS1 promotes the proliferation and inhibits the apoptosis of bladder cancer cells<sup>5</sup>. Activated by ZEB1, lncRNA HCCL5 accelerates cell viability, cell migration, epithelial-mesenchymal transition (EMT), as well as the malignancy of hepatocellular carcinoma<sup>6</sup>. Moreover, the over-expression of lincRNA-p21 represses the proliferation of gastric cancer cells, whereas increases cell sensitivity of radiotherapy by regulating the beta-catenin

signaling pathway<sup>7</sup>. However, the specific role of lncRNA LINP1 in the progression of Wilms' tumor and the possible underlying molecular mechanism have not been fully elucidated yet.

In this work, LINP1 was highly expressed in Wilms' tumor tissues when compared with adjacent normal tissues. Moreover, LINP1 significantly promoted the proliferation and metastasis of Wilms' tumor *in vitro*. Previous studies have showed that the Wnt/ $\beta$ -catenin signaling pathway is a fundamental pathway in tumor development. Our findings also demonstrated that LINP1 participated in the tumorigenesis of Wilms' tumor by regulating the Wnt/ $\beta$ -catenin signaling pathway.

## Patients and Methods

### Clinical Samples

Tumor samples and adjacent tissues ( $\geq 5$  cm away from the edge of tumor) were collected from Wilms' tumor patients ( $n = 52$ ) who underwent surgery at Jining No. 1 People's Hospital from 2010 to 2012. Written informed consent was obtained from each patient before the operation. All fresh tissues were preserved at  $-80^{\circ}\text{C}$  for subsequent use. This study was approved by the Human Research Ethics Committee of the Jining No. 1 People's Hospital.

### Cell Culture

Cells were first collected and cultured from fresh Wilms' tumor tissues. All cells were cultured in Dulbecco's Modified Eagle's Medium (DMEM, Gibco, Grand Island, NY, USA) containing 10% fetal bovine serum (FBS, Gibco, Grand Island, NY, USA), and maintained in an incubator with 5%  $\text{CO}_2$  at  $37^{\circ}\text{C}$ .

### Cell Transfection

After synthesis, short hairpin RNA (shRNA) targeting LINP1 (sh-LINP1), lentivirus targeting LINP1 (LINP1) or scrambled oligonucleotides (NC) were packaged into pLVH1/GFP+Puro vector (GenePharm, Shanghai, China). Then, Wilms' tumor cells were transfected with sh-LINP1, LINP1 lentivirus (LINP1) and NC according to the instructions. Subsequently, GFP-positive cells were selected for the following experiments.

### RNA Extraction and Real Time Quantitative Polymerase Chain Reaction (RT-qPCR)

Total RNA in tissues and cells was extracted in strict accordance with the TRIzol reagent (Invit-

rogen, Carlsbad, CA, USA). After that, extracted total RNA was reverse-transcribed to complementary deoxyribose nucleic acids (cDNAs) through the Reverse Transcription Kit (TaKaRa, Dalian, China). Primers used for Real Time-quantitative Polymerase Chain Reaction (RT-qPCR) were as follows: LINP1, forward: 5'-AGCCGGTCCAGTACAC-3' and reverse: 5'-GGAAAGCA-3'; GAPDH, forward: 5'-CAAAATCAGAT-3' and reverse: 5'-TGATGG-CATGGACTGTC-3'. Thermal cycle was as follows: 95 $^{\circ}\text{C}$  for 30 sec, 95 $^{\circ}\text{C}$  for 5 sec, 60 $^{\circ}\text{C}$  for 35 sec at 60 cycles.

### Western Blot Analysis

Cells were first washed with pre-cooled Phosphate Buffered Saline (PBS, Gibco, Grand Island, NY, USA) and lysed with cell lysis solution (RIPA; Beyotime, Shanghai, China). The concentration of extracted protein was detected by the bicinchoninic acid (BCA) method (Thermo Fisher Scientific, Waltham, MA, USA). After separation, proteins were transferred onto polyvinylidene difluoride (PVDF) membranes (Millipore, Billerica, MA, USA). Then, the membranes were blocked with Tris-Buffered Saline and Tween (TBST; Sigma-Aldrich, St. Louis, MO, USA; 25 mM Tris, 140 mM NaCl, and 0.1% Tween 20, pH 7.5) containing 5% skimmed milk for 2 h. Subsequently, the membranes were incubated with primary antibodies of target proteins including Wnt3a,  $\beta$ -catenin, C-myc and Survivin (Abcam, Cambridge, MA, USA) in the Wnt/ $\beta$ -catenin signaling pathway and GAPDH (Abcam, Cambridge, MA, USA) at  $4^{\circ}\text{C}$  overnight. After washing with TBST three times (10 min for each), the membranes were incubated with the corresponding secondary antibody at room temperature for 1 h. Finally, immunoreactive bands were analyzed by Image J software (NIH, Bethesda, MD, USA).

### MTT (3-(4,5-Dimethylthiazol-2-yl)-2,5-Diphenyl Tetrazolium Bromide) Assay

A total of  $2 \times 10^3$  transfected cells were first seeded into 96-well plates. Following manufacturer's protocol, cell proliferation was assessed by Cell Proliferation Reagent Kit I (MTT; Roche, Basel, Switzerland) every 24 h. Absorbance at 490 nm was assessed using an enzyme-linked immunosorbent assay (ELISA) reader system (Multiskan Ascent, LabSystems, Helsinki, Finland).

### Colony Formation Assay

Transfected cells were first seeded into 6-well plates, followed by culture for 2 weeks. Formed colonies were fixed with methanol for 30 min and stained with 0.5% crystal violet for 5 min. Colonies containing more than 50 cells were counted, and the mean number of formed colonies was calculated. The analysis was conducted with Image-Pro Plus 6.0 (Media Cybernetics, Silver Springs, MD, USA).

### Wound Healing Assay

Cells were first seeded into 6-well plates and incubated overnight. After being scratched with a pipette tip, the cells were cultured in serum-free DMEM. At 48 h, the relative distance was observed under a light microscope (Olympus, Tokyo, Japan). Each assay was independently repeated in triplicate.

### Transwell Assay

A total of  $1 \times 10^5$  cells in serum-free DMEM were seeded into the upper chamber (Corning Inc., Lowell, MA, USA) of 24-well plates pre-coated with Matrigel Matrix dilution (BD Biosciences, Franklin Lakes, NJ, USA). Meanwhile, the lower chamber was added with DMEM and FBS. 48 h later, after being wiped by a cotton swab, the top surface of chambers was immersed with 4% paraformaldehyde for 10 min and stained with 0.5% crystal violet for 30 min. The cells were randomly for each sample, and the number of invading cells was counted under a Leica DM14000B microscope (Leica Microsystems, Wetzlar, Germany).

### Statistical Analysis

GraphPad Prism 5.0 (La Jolla, CA, USA) was adopted for all statistical analysis. Data were expressed as mean  $\pm$  SD (Standard Deviation). Student's *t*-test and Kaplan-Meier method were utilized when appropriate. *p*-value  $< 0.05$  was considered statistically significant.

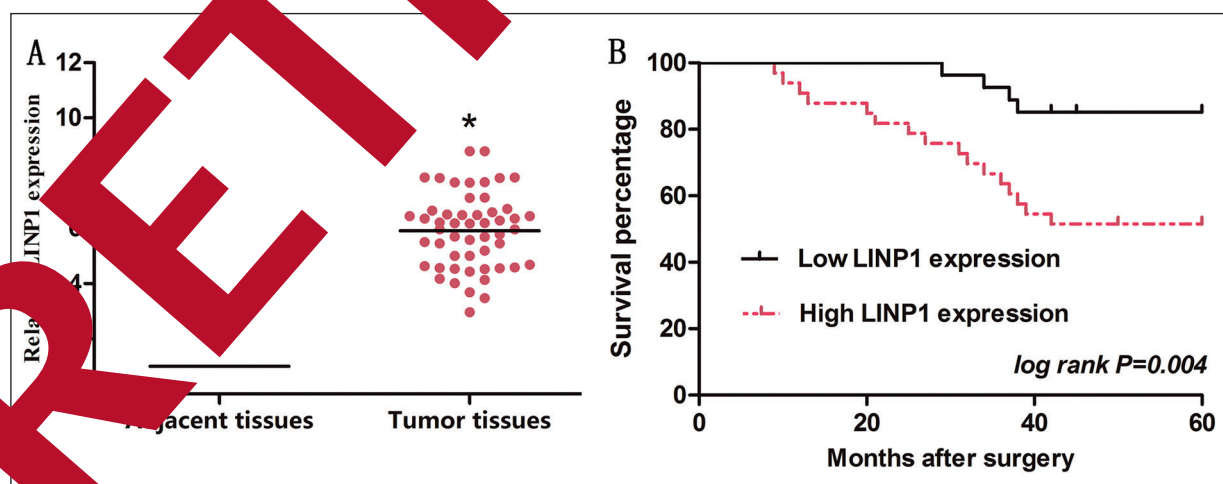
## Results

### LINP1 Expression in Wilms' Tumor Tissues

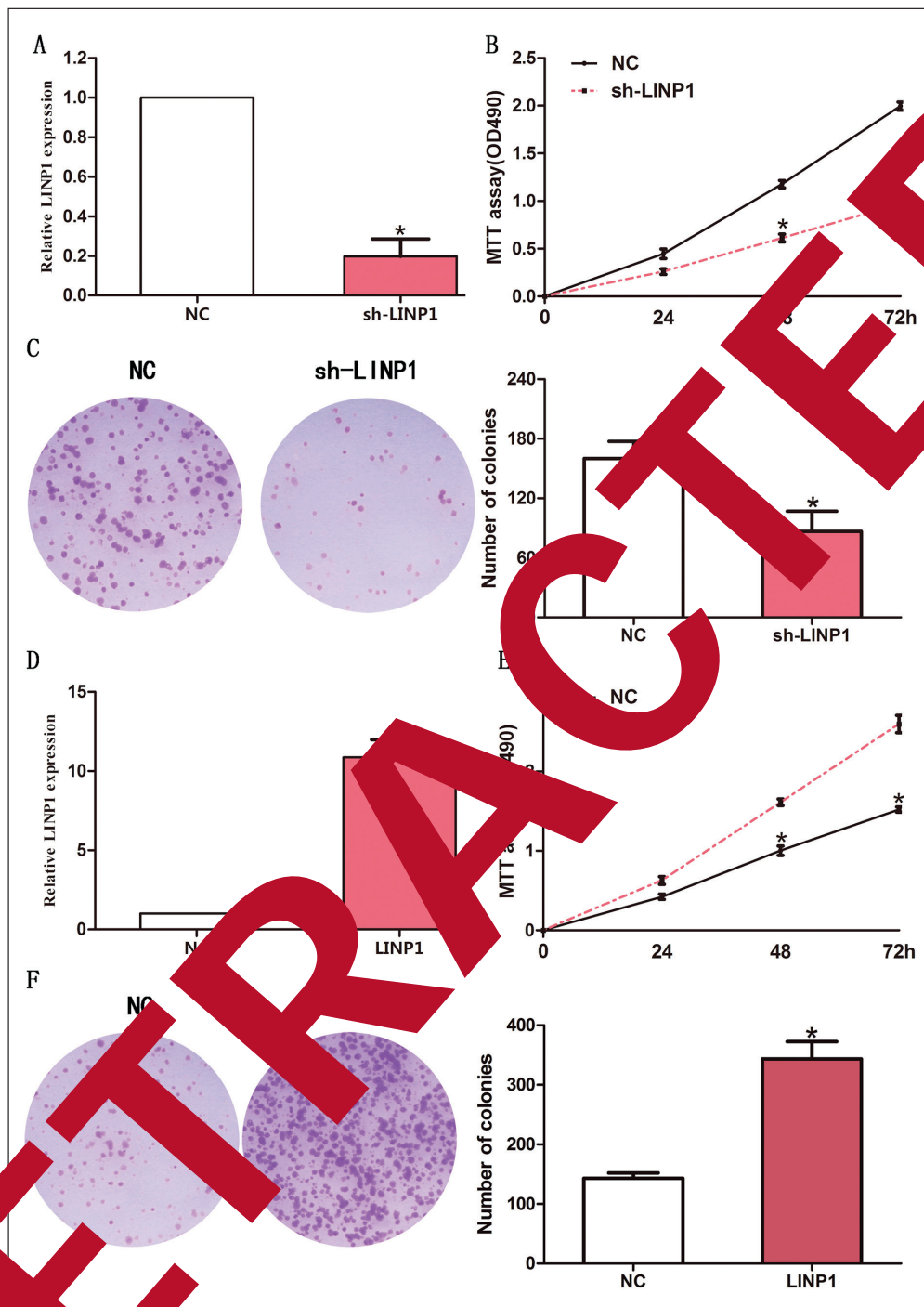
First, RT-qPCR was conducted to detect LINP1 expression in 52 patients' tumor tissues. The results showed that LINP1 was markedly up-regulated in Wilms' tumor tissue samples (Figure 1A). According to the mean expression of LINP1, 52 patients were divided into two groups, including high LINP1 expression group and low LINP1 expression group. The Kaplan-Meier analysis showed that the overall survival of patients in high LINP1 expression group was significantly worse when compared with the low LINP1 expression group (Figure 1B).

### LINP1 Overexpression and Regulation of LINP1 Inhibited Proliferation of Wilms' Tumor Cells

Collected and digested cells from Wilms' tumor tissues were used for transfection of LINP1 shRNA or scrambled oligonucleotides (NC). RT-qPCR was then utilized to detect LINP1 expression (Figure 2A). The results of the MTT assay revealed that the growth ability of Wilms' tumor cells was remarkably repressed after



**Figure 1.** Expression level of LINP1 was significantly increased in Wilms' tumor patients. **A**, LINP1 expression was markedly increased in Wilms' tumor tissues when compared with adjacent tissues. **B**, The expression level of LINP1 was negatively associated with patients' overall survival time. \**p*<0.05.



**Figure 2.** LIN1 promoted Wilms' tumor cell proliferation. **A**, LIN1 expression in Wilms' tumor cells transfected with LIN1 shRNA (sh-LIN1) or scrambled oligonucleotides (NC) was detected by RT-qPCR. GAPDH was used as an internal control. **B**, MTT assay revealed that the growth ability of Wilms' tumor cells was remarkably repressed in the sh-LIN1 group when compared with the NC group. **C**, Colony formation assay revealed that the number of Wilms' tumor cell colonies was remarkably reduced in the sh-LIN1 group when compared with the NC group (Magnification  $\times 40$ ). **D**, LIN1 expression in Wilms' tumor cells transfected with LIN1 lentivirus (LIN1) or scrambled oligonucleotides (NC) was detected by RT-qPCR. GAPDH was used as an internal control. **E**, MTT assay revealed that the growth ability of Wilms' tumor cells was remarkably enhanced in the LIN1 lentivirus (LIN1) group when compared with the NC group. **F**, Colony formation assay also revealed that the number of Wilms' tumor cell colonies was remarkably increased in the LIN1 lentivirus (LIN1) group when compared with the NC group (magnification  $\times 40$ ). The results represented the average of three independent experiments (mean  $\pm$  standard error of the mean). \* $p < 0.05$ , compared with control cells.



down-regulation LINP1 (Figure 2B). Colony formation assay also revealed that the number of formed colonies was markedly reduced due to loss of LINP1 *in vitro* (Figure 2C). To further identify the function of LINP1 in Wilms' tumor, collected and digested cells from Wilms' tumor tissues were used for transfection of LINP1 lentivirus (LINP1) or scrambled oligonucleotides (NC). Similarly, RT-qPCR was utilized to detect LINP1 expression (Figure 2D). The results of the MTT assay revealed that the growth ability of Wilms' tumor cells was significantly promoted after over-expression of LINP1 (Figure 2E). Furthermore, colony formation assay demonstrated that the number of formed colonies was remarkably increased after over-expression of LINP1 in Wilms' tumor cells (Figure 2F).

#### **Downregulation of LINP1 Inhibited Migration and Invasion of Wilms' Tumor Cells**

Wound healing assay revealed that the relative migrating distance of Wilms' tumor cells was markedly repressed after the loss of LINP1 (Figure 3A). Subsequent transwell assay also indicated that the number of invaded cells was remarkably reduced after the loss of LINP1 in Wilms' tumor cells (Figure 3B). Similarly, after over-expression of LINP1 *in vitro*, the relative migrating distance of Wilms' tumor cells was significantly promoted (Figure 3C). In addition, transwell assay illustrated that the number of invaded cells was remarkably increased after over-expression of LINP1 in Wilms' tumor cells (Figure 3D).

#### **Interaction Between Wnt/ $\beta$ -catenin Signaling Pathway and LINP1 in Wilms' Tumor**

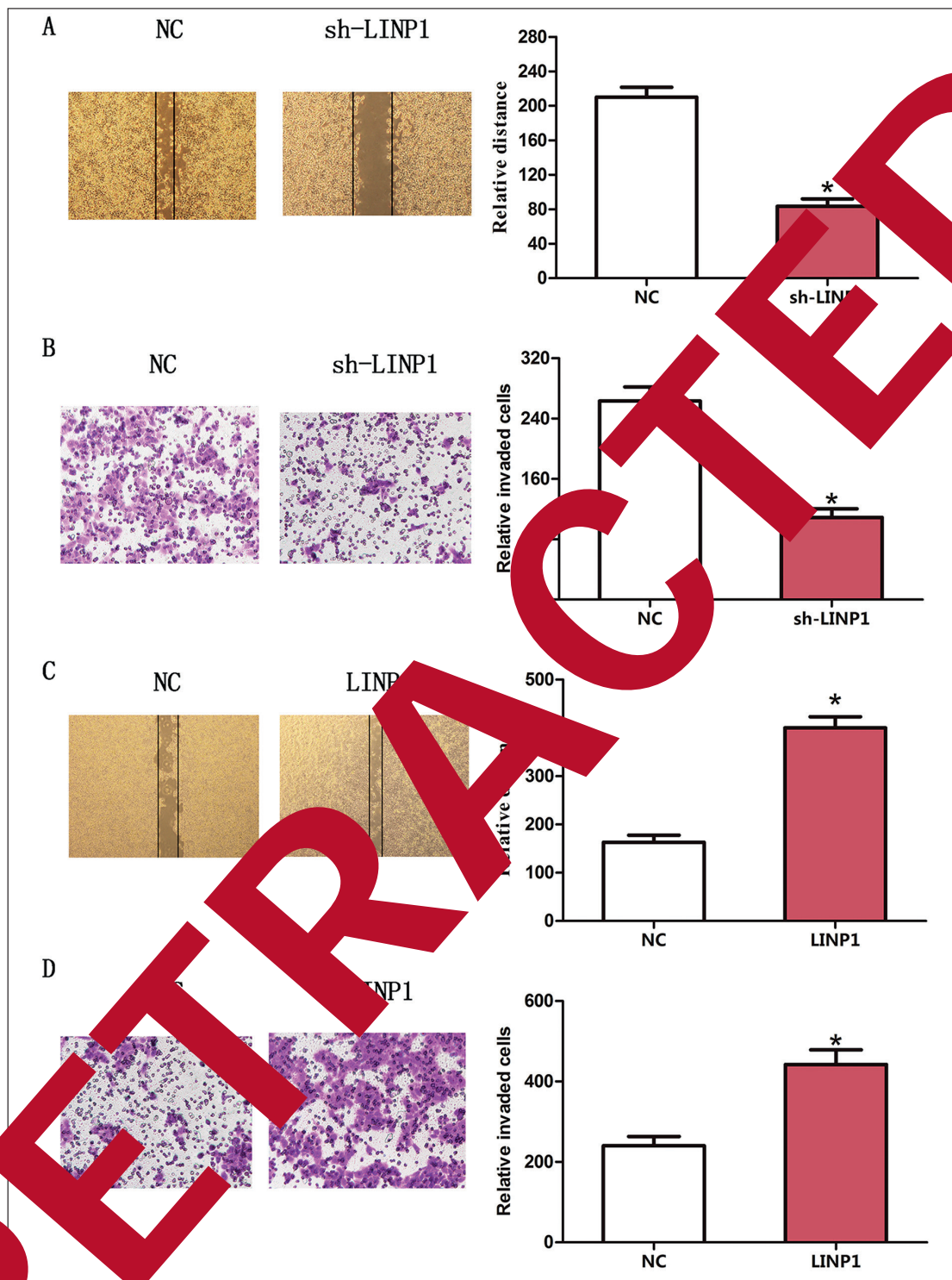
To explore the underlying mechanism of LINP1 function in Wilms' tumor, Western blot assays were conducted to detect the expressions of target proteins in the Wnt/ $\beta$ -catenin signaling pathway, including Wnt3a,  $\beta$ -catenin, C-myc and Survivin. The results showed that the protein expressions of Wnt3a,  $\beta$ -catenin, C-myc and Survivin were significantly down-regulated after the loss of LINP1 (Figure 4A). However, the expressions of Wnt3a,  $\beta$ -catenin, C-myc and Survivin were markedly up-regulated after over-expression of LINP1 (Figure 4B). These results suggested that LINP1 participated in the regulation of the Wnt/ $\beta$ -catenin signaling pathway, further promoting Wilms' tumor development and progression.

## **Discussion**

Numerous studies have proved that ncRNAs participate in a variety of important biological processes, including tumor growth. Previous evidence has revealed that several ncRNAs play an important role in the development of Wilms' tumor. For instance, the over-expression of miR-21 and low-expression of PTEN inhibit the proliferation and invasion of Wilms' tumor cells by targeting FRS2, miR-195 depresses the proliferation and migration of Wilms' tumor. This may provide an innovative target for the diagnosis and therapy of Wilms' tumor. Over-expressing tumor suppressor miR-195, LINP1 functions as an oncogene in Wilms' tumor by regulating IKK $\alpha$ .

Non-homologous end joining (NHEJ) is one of the major mechanisms for repairing damaged DNA in cancer cells. Previous studies have demonstrated that lncRNA in non-homologous end joining pathway 1 (LINP1) promotes NHEJ-mediated DNA repair in multiple malignancies. For example, LINP1 regulates NHEJ signal pathway and promotes DNA damage repair in cervical cancer cells, further decreasing radiation sensitivity<sup>11,12</sup>. LINP1 functions as an oncogene in breast cancer, which facilitates the progression and chemoresistance<sup>13</sup>. The over-expression of LINP1 enhances the malignant progression of prostate cancer by negatively modulating p53<sup>14</sup>. However, Zhang et al<sup>15</sup> have shown that LINP1 serves as a tumor suppressor in lung cancer by inhibiting EMT. Therefore, we explored the role of LINP1 in Wilms' tumor. The results showed that LINP1 was significantly up-regulated in Wilms' tumor tissues. Meanwhile, LINP1 expression was associated with the prognosis of patients. Besides, the proliferation and metastasis of Wilms' tumor cells were markedly inhibited after the loss of LINP1. However, the proliferation and metastasis of Wilms' tumor cells were remarkably promoted after over-expression of LINP1. The above results indicated that LINP1 promoted tumorigenesis of Wilms' tumor and might act as an oncogene.

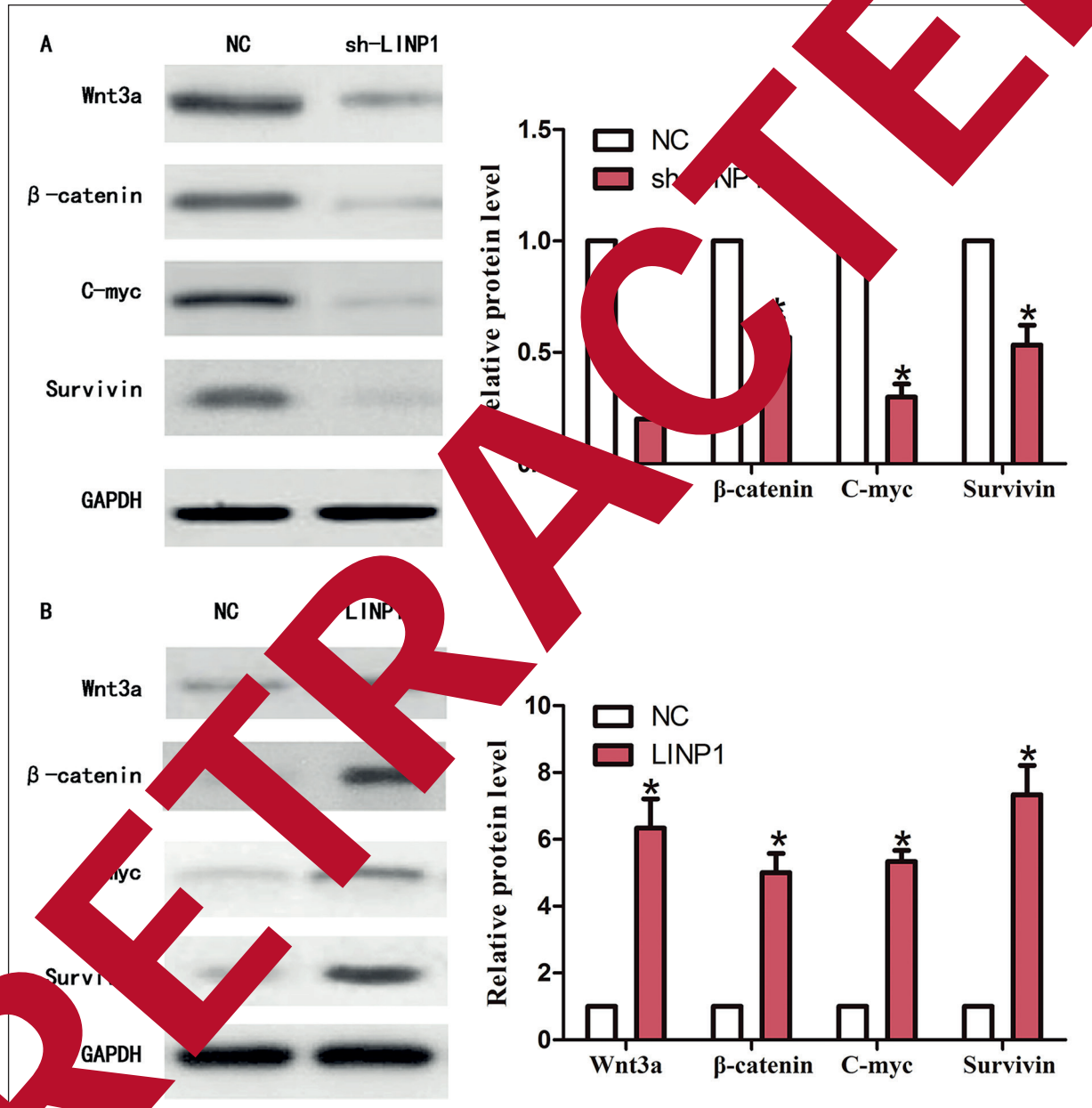
Wnt proteins mediate diverse processes during embryogenesis by modulating stem cell division and migration. Wnt3a,  $\beta$ -catenin, C-myc and Survivin are target proteins in the Wnt/ $\beta$ -catenin signaling pathway. Li et al<sup>16</sup> have suggested that aberrant activation of the Wnt/ $\beta$ -catenin signaling pathway plays an important role in regulat-



**Fig. 4.** LINP1 promoted Wilms' tumor cell migration and invasion. **A**, The relative migrating distance of Wilms' tumor cells was significantly decreased in the sh-LINP1 group when compared with the NC group (Magnification  $\times 10$ ). **B**, The transwell assay showed that the number of invaded Wilms' tumor cells was markedly decreased in the sh-LINP1 group when compared with the NC group (Magnification  $\times 40$ ). **C**, The relative migrating distance of Wilms' tumor cells was remarkably increased in the LINP1 lentivirus (LINP1) group when compared with the NC group (Magnification  $\times 10$ ). **D**, The transwell assay showed that the number of invaded Wilms' tumor cells was significantly increased in the LINP1 lentivirus (LINP1) group when compared with the NC group (magnification  $\times 40$ ). The results represented the average of three independent experiments (mean  $\pm$  standard error of the mean). \* $p < 0.05$ , compared with control cells.

ing the development of several human cancers. For instance, by modulating the Wnt/ $\beta$ -Catenin/Axin2 signaling, c-Myb facilitates the invasion and migration of breast cancer cells. Through the activation of the Wnt/ $\beta$ -catenin signaling inhibitors (DKK1 and SFRP2), TET1 serves as a tumor suppressor in ovarian cancer *via* inhibiting EMT<sup>17</sup>. LncRNA CRNDE enhances the prolifer-

ation and chemoresistance of colorectal cancer *via* modulating the expression of miR-181a-5p, which directly mediates the regulation of the Wnt/ $\beta$ -catenin pathway<sup>18</sup>. Our work shows that the expressions of target proteins in the Wnt/ $\beta$ -catenin signaling pathway were significantly down-regulated after the knockdown of LINP1. However, the protein expression level of target



**Figure 4.** Interaction between LINP1 and Wnt/ $\beta$ -catenin signaling pathway. **A**, Western blot assay revealed that the expressions of target proteins in the Wnt/ $\beta$ -catenin signaling pathway were significantly down-regulated in the sh-LINP1 group when compared with the NC group. **B**, Western blot assay indicated that the expressions of target proteins in the Wnt/ $\beta$ -catenin signaling pathway were markedly up-regulated in the LINP1 lentivirus (LINP1) group when compared with the NC group. The results represented the average of three independent experiments. \* $p < 0.05$ .

proteins in the Wnt/ $\beta$ -catenin signaling pathway were remarkably up-regulated after over-expression of LINP1. All the results above suggested that LINP1 might promote tumorigenesis of Wilms' tumor *via* regulating the Wnt/ $\beta$ -catenin signaling pathway.

## Conclusions

LINP1 enhanced Wilms' tumor cell proliferation and metastasis by regulating the Wnt/ $\beta$ -catenin signaling pathway. Our findings indicated that LINP1 might contribute to therapy for Wilms' tumor as a candidate target.

## Conflict of Interest

The Authors declare that they have no conflict of interests.

## References

- CHARLTON J, PAVASOVIC V, PRITCHARD-JONES K. Biomarkers to detect Wilms tumors in pediatric patients: where are we now? *Future Oncol* 2015; 11: 21-2234.
- CONE EB, DALTON SS, VAN NOORD M, TRACY ET, HE, ROUTH JC. Biomarkers for Wilms tumor: a systematic review. *J Urol* 2016; 196: 1535.
- LIU G, ZHANG Y, FU K, HUANG Z, JIA W, LIU G. Meta-analysis of the effect of preoperative chemotherapy on Wilms' tumor. *J BUON* 2018; 23: 211-217.
- GABALLAH HH, GABALLAH RA, ELHAYAT MA, EL SHAHAT DA, HABLUS M, ELBEID AM. Expression of long non-coding RNA H19 in colorectal carcinoma: correlation with clinicopathological features and ERK/COX-2 pathway. *Cell Biol Rep* 2018; 10.1007/s11038-18-4521-0.
- WANG H, SHI F, XIA Y, ZHAO Y. lncRNA OIP5-AS1 predicts poor prognosis and regulates cell proliferation and apoptosis in bladder cancer. *J Cell Biochem* 2018; 10.1002/jcb.28024.
- PENG Y, CHEN B, YU X, QIU Y, PENG J, HUANG Y, ZHANG Y, LI Z, LI J, YAO W, DENG W, ZHANG Y, MENG M, LI C, YIN D, BI X, LI G, LIN DC. ZEB1 up-regulates and activates long noncoding RNA LINC00473, which is activated by ZEB1 and promotes the proliferation of hepatocellular carcinoma. *Cancer Res* 2017; 79: 572-584.
- CHEN L, YUAN D, YANG Y, REN M. LincRNA-p21 enhances the sensitivity of radiotherapy for gastric cancer by targeting the beta-catenin signaling pathway. *J Cell Biochem* 2018; 10.1002/jcb.27905.
- CUI M, LIU W, ZHANG L, GUO F, LIU Y, CHEN MA R, WU R. Over-expression of miR-195 up-regulates PTEN levels in Wilms' tumor with aggressive behavior. *Tohoku J Exp Med* 2017; 2017: 43-52.
- WANG HF, ZHANG YY, ZHUANG H, LIU JM. MicroRNA-613 attenuates the proliferation, migration and invasion of Wilms' tumor *via* targeting PTPN22. *Eur Rev Med Pharmacol Sci* 2017; 41: 3369.
- ZHU S, FU W, ZHANG Y, FU K, LIU J, JIA W, LIU G. LINC00473 antagonizes the tumor suppressor miR-195 to promote the proliferation of Wilms tumor *via* IKK $\alpha$ . *Cell Prolif* 2018; 1: e12416.
- ZHANG Y, HUANG Z, HU Z, FENG Y, LIU J, YUAN J, SHAN W, GUO R, TANYI JL, FAN Y, GUANG Q, MONTONE T, DANG C, ZHANG L. Long noncoding RNA LINP1 regulates the repair of DNA double-strand breaks in triple-negative breast cancer. *Nat Struct Mol Biol* 2016; 23: 522-529.
- WANG X, LIU H, SHI L, YU X, GU Y, SUN X. LINP1 facilitates DNA damage repair through non-homologous end joining (NHEJ) pathway and subsequently decreases the sensitivity of cervical cancer cells to ionizing radiation. *Cell Cycle* 2018; 17: 407-416.
- LIANG Y, LI Y, SONG X, ZHANG N, SANG Y, ZHANG H, LIU Y, CHEN B, ZHAO W, WANG L, GUO R, YU Z, YANG Q. Long noncoding RNA LINP1 acts as an oncogene and promotes chemoresistance in breast cancer. *Cancer Biol Ther* 2018; 19: 120-131.
- WU HF, REN LG, XIAO JQ, ZHANG Y, MAO XW, ZHOU LF. Long non-coding RNA LINP1 promotes the malignant progression of prostate cancer by regulating p53. *Eur Rev Med Pharmacol Sci* 2018; 22: 4467-4476.
- ZHANG C, HAO Y, WANG Y, XU J, TENG Y, YANG X. TGF-beta/SMAD4-regulated lncRNA-LINP1 inhibits epithelial-mesenchymal transition in lung cancer. *Int J Biol Sci* 2018; 14: 1715-1723.
- LI Y, JIN K, VAN PELT GW, VAN DAM H, YU X, MESKER WE, TEN DUKE P, ZHOU F, ZHANG L. c-Myb enhances breast cancer invasion and metastasis through the Wnt/beta-catenin/Axin2 pathway. *Cancer Res* 2016; 76: 3364-3375.
- DUAN H, YAN Z, CHEN W, WU Y, HAN J, GUO H, QIAO J. TET1 inhibits EMT of ovarian cancer cells through activating Wnt/beta-catenin signaling inhibitors DKK1 and SFRP2. *Gynecol Oncol* 2017; 147: 408-417.
- HAN P, LI JW, ZHANG BM, LV JC, LI YM, GU XY, YU ZW, JIA YH, BAI XF, LI L, LIU YL, CUI BB. The lncRNA CRNDE promotes colorectal cancer cell proliferation and chemoresistance *via* miR-181a-5p-mediated regulation of Wnt/beta-catenin signaling. *Mol Cancer* 2017; 16: 9.