

MiR-155 regulates the proliferation and apoptosis of pancreatic cancer cells through targeting SOCS3

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Abstract. – **OBJECTIVE:** Reduced expression of suppressors of cytokine signaling 3 (SOCS3) is associated with a variety of tumors. The elevated miR-155 expression is associated with the onset of pancreatic cancer. Bioinformatics analysis revealed a targeted relation between miR-155 and the 3'-UTR of SOCS3. This study investigated whether miR-155 regulates SOCS3 expression and affects the biological effects of pancreatic cancer cells.

PATIENTS AND METHODS: QRT-PCR was used to detect the expression of miR-155 and SOCS3 mRNA in tumor tissues and pancreatic tissues of patients with pancreatic cancer. The dual luciferase reporter gene assay validated the target interaction between miR-155 and SOCS3. Pancreatic cancer cell line SW1990 cells were divided into miR-NC group and miR-155 inhibitor group followed by detection of the expressions of SOCS3, p-JAK2 and p-STAT3, cell apoptosis by flow cytometry, and cell proliferation by EdU staining.

RESULTS: Compared with normal tissues, miR-155 expression was increased in tumor tissues of patients with pancreatic cancer, and SOCS3 expression was decreased. miR-155 was a targeted regulatory relationship between miR-155 and SOCS3 mRNA. Compared with HP-DE6-C7 cells, miR-155 expression in pancreatic cancer SW1990 and Capan-1 cells was increased and SOCS3 expression was decreased. Transfection of miR-155 inhibitor significantly increased SOCS3 expression in pancreatic cancer cells, decreased the expression of p-JAK2 and p-STAT3, increased cell apoptosis, and decreased cell proliferation.

CONCLUSION: Increased miR-155 expression and decreased SOCS3 expression are related to the pathogenesis of pancreatic cancer. miR-155 inhibits the proliferation and apoptosis of pancreatic cancer cells by inhibition of SOCS3 expression.

Keywords:

miR-155, SOCS3, JAK-STAT, Pancreatic cancer.

Introduction

Pancreatic carcinoma (PC) is a common malignant tumor of the digestive system. It has a very high degree of malignancy, so the treatment effect and prognosis are extremely poor with a high mortality rate.

SOCS3 is a member of the cytokine signal transduction inhibitory protein (SOCS) family, which blocks the activation and transmission of the JAK-STAT signaling pathway by inhibiting the activity of JAK kinase². As an important tumor suppressor, the expression or decreased function of SOCS3 plays an important role in the occurrence, progression, and metastasis of various tumors³⁻⁵. There is research evidence that abnormal expression or dysfunction of SOCS3 is associated with the occurrence, progression, and metastasis of pancreatic cancer⁶⁻⁸.

MicroRNA (microRNA) is an endogenous non-coding small RNA molecule in eukaryotes that bind to the 3'-untranslated region (3'-UTR) of the target gene mRNA through complementary pairing, leading to degradation of mRNA or inhibition of mRNA translation, thus regulating the expression of target genes and participating in the regulation of biological processes such as cell survival, proliferation, apoptosis, migration. MicroRNA expression and dysfunction have been shown to be associated with non-small cell lung cancer⁹, gastric cancer¹⁰, prostate cancer¹¹, bladder cancer¹², and other tumors. A number of studies¹³⁻¹⁶ have shown that the abnormal expression and function of miR-155 are related to the occurrence, progression, metastasis, and prognosis of various tumors such as breast cancer, gastric cancer, lung cancer and intestinal cancer. Several researches¹⁷⁻¹⁹ have shown that abnormal changes in the expression of miR-155 are associated with the

development, progression, and prognosis of pancreatic cancer. Bioinformatics analysis revealed a targeted binding site between miR-155 and the 3'-UTR of SOCS3 mRNA. This work investigated whether miR-155 plays a role in regulating SOCS3 expression, affecting JAK-STAT pathway activity, pancreatic cancer cell proliferation, and apoptosis.

Patients and Methods

Patients

30 patients with pancreatic cancer who were treated in our hospital from June 2018 to November 2018 were enrolled. The tumor tissues of pancreatic cancer confirmed by pathological examination were collected and the paracancerous tissues located at least 2 cm away from the tumor tissue were collected as control. This research was approved by the Ethic Committee of our hospital and informed consent was obtained from the patients.

Main Reagents and Materials

Human normal pancreatic epithelial cells HPDE6-C7 were purchased from Hunan Zhonghui organism; pancreatic cancer cells SW1990 and Capan-1 were purchased from Shanghai Siyuan organism; Dulbecco's Modified Medium (DMEM) was purchased from Gibco (Grand Island, NY, USA); Fetal bovine serum (FBS) was purchased from Shanghai Sai Biotechnology Company; Lip2000 was purchased from Invitrogen (Carlsbad, CA, USA); PrimeScript™ RT reagent Kit, SYBR Green dye purchased from TaKaRa (Dalian, China); miR-155 inhibitor, miR-NC, miR-155 mimic were designed and synthesized by Ruibo Bio (Guangzhou, China); rabbit anti-human STAT3, p-JAK2 antibody purchased in the CST; rabbit anti-human antibody was purchased from Santa Cruz Biotechnology (Santa Cruz, CA, USA); rabbit anti-human SOCS3, β -actin antibody, HRP-conjugated secondary antibody were purchased from Abcam (Cambridge, MA, USA); pGL3 plasmid was purchased from Chemicon (La Jolla, CA, USA); luciferase detection kit was purchased from Promega (Madison, WI, USA); the EdU-Alexa488 cell proliferation assay kit was purchased from Thermo Fisher Scientific (Waltham, MA, USA); the Annexin V-FITC/PI apoptosis assay reagent was purchased from Jiangsu Biyuntian Science and Technology.

Cell Culture

HPDE6-C7, SW1990, and Capan-1 were inoculated in DMEM medium containing 10% FBS and 1% streptomycin, and cultured in a cell culture incubator at 37°C with 5% CO₂ until the cells reached a confluence of 90%. After digestion with 0.25% trypsin, the cells were collected by centrifugation, sub-cultured at a ratio of 1:1:5, and cells in the logarithmic growth phase were selected for experiments.

Dual Luciferase Reporter Gene Test

Using the HPK293T genome as a template, the full-length 3'-UTR fragment of the SOCS3 gene and the fragment containing the mutant was amplified and cloned into the pMIR vector, transformed into DH5 α competent cells, and the correct plasmids were sequenced and named as pMIR-SOCS3-WT and pMIR-SOCS3-MUT. pMIR-SOCS3-WT (or pMIR-SOCS3-MUT) and miR-155 mimic (or miR-NC) were co-transfected into HEK293T cells with Lip2000. After 48 h of culture, the luciferase activity kit was used to detect the relative luciferase activity.

Cell Transfection

SW1990 cells were cultured *in vitro* and divided into two transfection groups: miR-NC transfection group and miR-155 inhibitors transfection group. The general procedure for transfection was as follows: 10 μ L of Lip2000, 50 nmoL miR-NC, and 50 nmoL miR-155 inhibitor were diluted with 100 μ L of serum-free Opti-MEM, and incubated for 5 min at room temperature, respectively, and Lip2000 and miR-NC, miR-155 inhibitor, respectively. They were gently mixed, incubated for 20 min at room temperature, and the transfectants were added to the cell culture medium, mixed gently and continued to culture for 72 hours followed by a collection of the cells to measure the relevant parameters.

RT-PCR Detection of Gene Expression

RNA was extracted using TRIzol method and RNA was reversely transcribed into cDNA using PrimeScript™ RT reagent Kit. The resulting cDNA was stored in a refrigerator at -20°C. PCR amplification was carried out under the action of Taq DNA polymerase using cDNA as a template in a total of 10 μ L PCR reaction system including 2 \times SYBR Green Mixture 5.0 μ L, 2.5 μ m/L forward primer 0.5 μ L, 2.5 μ m/L reverse primer 0.5 μ L, cDNA 1 μ L, and ddH₂O. PCR conditions were designed as follows: 95°C for 5 min, 95°C for 15 s, 60°C for 1 min, fluorescence amplification data were stored on a Bio-Rad (Hercules,

CA, USA) CFX96 Real-time PCR instrument for 40 cycles. Primer sequences were: SOCS3-F: 5'-CCTGCGCCTCAAGACCTTC-3', SOCS3-R: 5'-GTCAGTGCCTCCAGTAGAA-3'; β -actin-F: 5'-CATGTACGTTGCTATCCAGGC-3', β -actin-R: 5'-CTCCTTAATGTCACGCACGAT-3'.

Western Blot

The cells were collected, and the protein was extracted from the RIPA lysate. The concentration was determined by bicinchoninic acid (BCA) assay and 40 μ g was separated in sodium dodecyl sulphate-polyacrylamide gel electrophoresis (SDS-PAGE) gel for 3 h, transferred to a polyvinylidene difluoride (PVDF) membrane by electroporation for 90 min, blocked with 5% skim milk powder, and incubated with the primary antibody at 4°C overnight (the dilution ratios of SOCS3, p-JAK2, p-STAT3, and β -actin were 1:2000, 1:800, 1:800, 1:8000, respectively), and subsequent washing three times with Phosphate-Buffered Saline and Tween (PBST). Next, the membrane was incubated with horseradish peroxidase (HRP)-labeled secondary antibody (1:8000 dilution) for 60 min at room temperature followed by washing 3 times with PBST. Enhanced chemiluminescence (ECL) working solution was added to blotting membrane for 1 min at room temperature, and subsequent exposure and development.

Flow Cytometry Detection of Cell Apoptosis

The above-mentioned miR-NC and miR-155 inhibitor transfection groups SW1990 cells were collected by trypsinization. After washing with PBS, 300 μ L of Binding Buffer was added to the cell pellet followed by adding 5 μ L of Annexin V-FITC for 15 min incubation in the dark. After that, 5 μ L of 1% sodium iodide (containing

was added, and 200 μ L of Binding Buffer was added followed by an analysis of cell apoptosis by flow cytometry.

Edu Staining for Cell Proliferation

The above two miR-NC and miR-155 inhibitor transfection group SW1990 cells were collected by trypsinization. SW1990 cells were reseeded in DMEM complete medium containing 10% FBS, incubated with 10 μ M EdU for 120 min, and cultured for 48 h. After enzymatic digestion and collection of cells, cell proliferation was detected by Beckman Coulter FC 500 flow cytometry according to the kit instructions. The proliferation of cells was reflected by EdU positive rate.

Statistical Analysis

Statistical analysis was performed using SPSS 17.0 software (SPSS Inc., Chicago, IL, USA). The measurement data were expressed as mean \pm standard deviation ($\bar{x} \pm s$). The comparison between the measurement data of the groups was analyzed by Student's *t*-test. $P < 0.05$ was considered statistically significant.

Results

Abnormal Expression of MiR-155 and SOCS3 in Pancreatic Cancer

The results of qRT-PCR showed that the expression of SOCS3 mRNA in tumor tissues of pancreatic cancer patients was significantly lower than that in adjacent tissues (Figure 1A). The results of qRT-PCR showed that the expression of miR-155 was significantly increased in tumor tissues of patients with pancreatic cancer compared with adjacent tissues (Figure 1B).

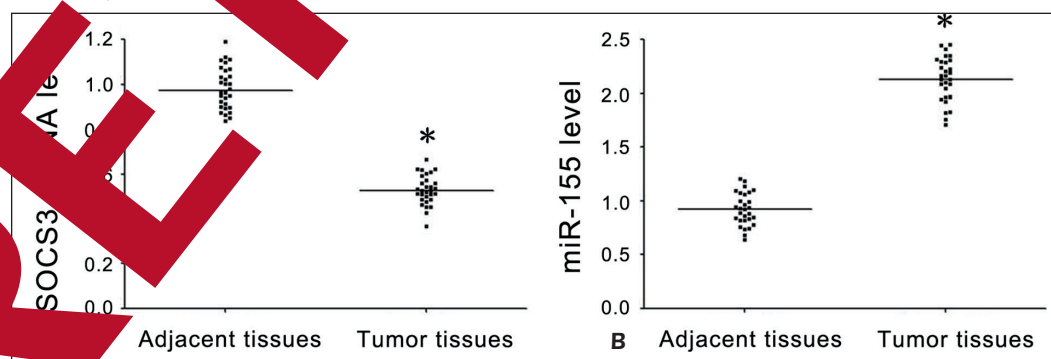


Figure 1. Abnormal expression of miR-155 and SOCS3 in pancreatic cancer. **A**, QRT-PCR detection of SOCS3 mRNA expression in pancreatic cancer tissues; **B**, QRT-PCR detection of miR-155 expression in pancreatic cancer tissues. *represents $p < 0.05$ compared with adjacent tissues.

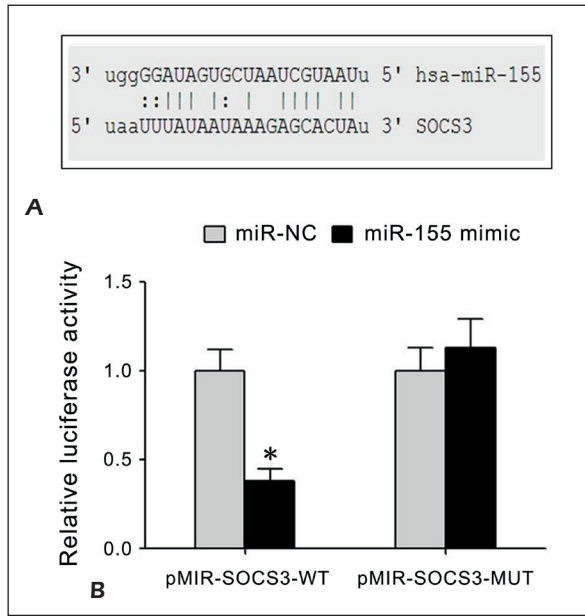


Figure 2. Targeted regulatory relationship between miR-155 and SOCS3 mRNA. A, Schematic diagram of the interaction site between miR-155 and the 3'-UTR of SOCS3 mRNA; B, Dual luciferase gene reporter assay. *represents $p < 0.05$ compared to miR-NC.

Targeted Relationship Between miR-155 and SOCS3 mRNA

Bioinformatics analysis revealed a complementary binding site between miR-155 and the 3'-UTR of SOCS3 mRNA (Figure 2A). Dual luciferase gene reporter assays showed that transfection of miR-155 mimic significantly reduced relative luciferase activity in pMIR-SOCS3-WT transfected HEK293T cells, but not in pMIR-SOCS3-MUT

mimic did not have significant effect on relative luciferase activity in HEK293T cells transfected with pMIR-SOCS3-MUT (Figure 2B), indicating that miR-155 has a targeted regulatory relationship with the 3'-UTR region of SOCS3 mRNA.

Increased MiR-155 and Decreased SOCS3 Expression in Pancreatic Cancer Cells

The qRT-PCR analysis showed that compared with normal pancreatic epithelial HPDE6-C7 cells, the expression of miR-155 in pancreatic cancer SW1990 and Capan-1 cells was significantly increased, while the expression of SOCS3 mRNA was significantly decreased (Figure 3A). Western blot analysis showed that the expression of SOCS3 protein in pancreatic cancer SW1990 and Capan-1 cells was significantly lower than that in HPDE6-C7 cells (Figure 3B).

Inhibition of miR-155 Expression Significantly Inhibits Pancreatic Cancer Cell Proliferation and Promotes Apoptosis

The qRT-PCR analysis showed that the expression of miR-155 in SW1990 cells was significantly increased in miR-155 inhibitor transfection group compared with miR-NC transfection group (Figure 4A), while the expression of SOCS3 mRNA was significantly increased (Figure 4B). Western blot analysis showed that compared with miR-NC transfection group, the expression of SOCS3 protein in SW1990 cells was significantly increased in miR-155 inhibitor transfection group, while the expression of p-JAK2 and p-STAT3 protein was

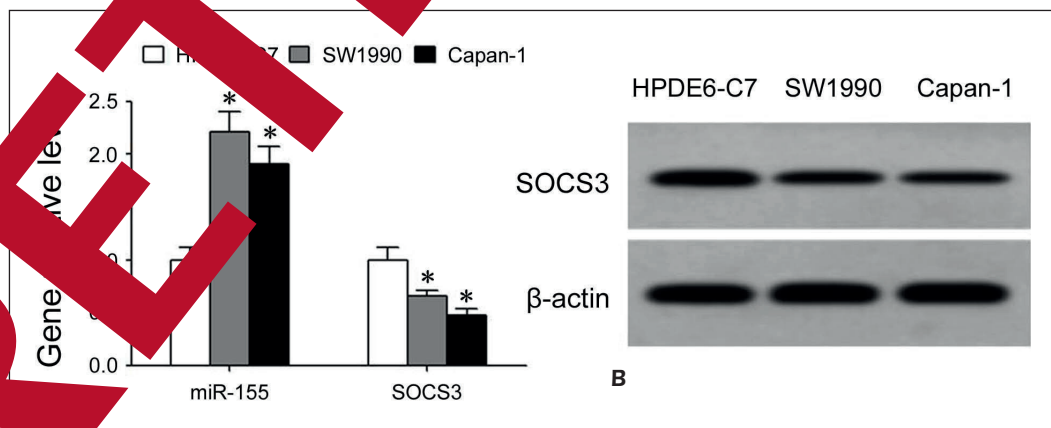
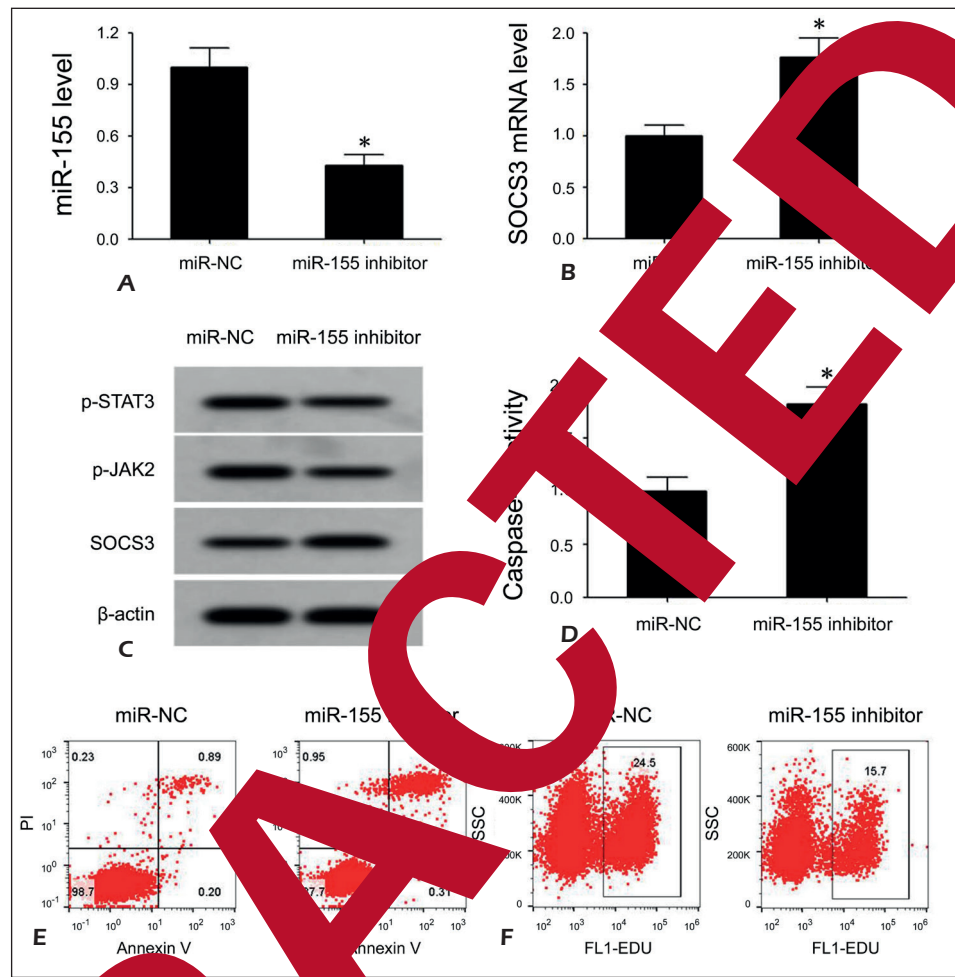


Figure 3. Increased expression of miR-155 in pancreatic cancer cells and decreased expression of SOCS3. A, QRT-PCR was used to detect the expression of miR-155 and SOCS3 mRNA in cells; B, Western blot was used to detect the expression of SOCS3 protein in cells. *represents $p < 0.05$ compared to HPDE6-C7 cells.

Figure 4. Inhibition of miR-155 expression can significantly inhibit pancreatic cancer cell proliferation and promote apoptosis. **A**, QRT-PCR detection of intracellular miR-155 expression; **B**, QRT-PCR detection of intracellular SOCS3 mRNA expression; **C**, Western blot detection of intracellular protein expression; **D**, Spectrophotometric detection of intracellular caspase-3 enzyme activity; **E**, Flow detection of apoptosis; **F**, EdU staining for cell proliferation. *represents $p < 0.05$ compared to miR-NC.



significantly decreased (Figure 4C). Spectrophotometry analysis showed that the caspase-3 activity was significantly increased in SW1990 cells of miR-155 inhibitor transfection group compared with miR-NC group (Figure 4D). Flow cytometry analysis showed that transfection of miR-155 inhibitor significantly increased apoptosis (Figure 4E) and inhibited proliferation of SW1990 cells (Figure 4F).

Discussion

The tyrosine kinase (JAK)-signal transducer and activator of transcription (STAT) signaling pathway is a signal-regulating pathway widely found in various tissues and cells of mammals. The JAK-STAT pathway is involved in a variety of biological processes that regulate cell survival, proliferation, migration, and invasion^{20,21}. SOCS is a negative feedback regulator in the JAK-STAT

signaling pathway and plays an important role in maintaining homeostasis in the cell. SOCS structurally includes an N-terminal kinase inhibitory region (KIR), a central SH2 domain, and a C-terminal SOCS-box structure. As one of the most widely expressed and most functional members of the cytokine signaling inhibitory (SOCS) family, SOCS3 directly inhibits JAK kinase activity and STAT phosphorylation and inhibits JAK-STAT signaling transduction². Researches¹⁷⁻¹⁹ have shown that abnormal changes in the expression of miR-155 are associated with the development, progression, and prognosis of pancreatic cancer. We investigated whether miR-155 plays a role in regulating SOCS3 expression, affecting JAK-STAT pathway activity, and pancreatic cancer cell proliferation and apoptosis.

The results of qRT-PCR showed that compared with the adjacent tissues, the expression of miR-155 was significantly increased in the tumor tissues of patients with pancreatic cancer, while

the expression of SOCS3 mRNA was significantly decreased, suggesting that increased miR-155 expression may play a role in reducing the expression of SOCS3 and promoting the pathogenesis of pancreatic cancer. In addition, the results of comparative studies showed that compared with normal pancreas HPDE6-C7 cells, the expression of miR-155 in pancreatic cancer SW1990 and Capan-1 cells was significantly increased, while the expression of SOCS3 was significantly decreased. The dual luciferase gene reporter assay showed that transfection of miR-155 mimic significantly reduced relative luciferase activity in pMIR-SOCS3-WT transfected HEK293T cells, but miR-155 mimic did not affect the relative luciferase activity in HEK293T cells transfected with pMIR-SOCS3-MUT, confirming the targeted regulation relationship between miR-155 and SOCS3 mRNA. In the study of the relationship between miR-155 and pancreatic cancer, Ali et al²² showed that the expression of miR-155 in pancreatic tumor tissues of pancreatic cancer model mice was significantly increased. LaConti et al²³ observed that the expression of miR-155 was significantly elevated in tumor tissues of patients with pancreatic cancer compared with normal pancreatic tissue. Liu et al²⁴ detected that the expression of miR-155 was significantly increased in tumor tissues of patients with pancreatic cancer compared with chronic pancreatitis and normal pancreatic tissue. Liu et al¹⁹ revealed that the expression of miR-155 was significantly increased in pancreatic cancer compared with adjacent tissues, and the expression of target gene SOCS3 was significantly decreased, and miR-155 promotes the pathogenesis of pancreatic cancer by inhibiting BLH1. Papanicolaou et al²⁵ pointed out that the expression of miR-155 was abnormally increased in pancreatic cancer tissues compared with the control, and the increase in miR-155 expression was associated with tumor stage and prognosis. Qu et al²⁶ found that the expression of miR-155 in peripheral blood of patients with pancreatic cancer was abnormally elevated. Seux et al²⁷ indicated that the expression of the tumor suppressor gene p53INP1 was associated with abnormally elevated expression of miR-155 in pancreatic cancer tissues, and miR-155 plays a role in tumor promotion by inhibiting the expression of p53INP1. In this study, the expression of miR-155 was significantly increased in pancreatic cancer tumor tissues and pancreatic cancer cell lines, and elevated miR-155 was a

cancer-promoting factor in the pathogenesis of pancreatic cancer, which was consistent with the results of LaConti et al²³, Liu et al²⁴ and Seux et al²⁷.

To further investigate the role and mechanism of miR-155 in the regulation of the biological effects of pancreatic cancer cells, this study transfected miR-155 inhibitor in pancreatic cancer cells and observed changes in the biological effects of pancreatic cancer cells. The results showed that transfection of miR-155 inhibitor significantly down-regulated the expression of miR-155 in pancreatic cancer cells and significantly increased the expression of SOCS3, while the expression of p-JAK2 and p-STAT3 was significantly decreased, thereby reducing cell proliferation and increasing cell apoptosis. In the study of the relationship between miR-155 and the biological effects of pancreatic cancer cells, Ali et al²² found that the proliferation of pancreatic cancer Rlnk-cells can significantly inhibit cell proliferation and weaken the ability of cell cloning. Lin et al²⁸ observed that the expression of miR-155 is related to pancreatic cancer, and emodin can inhibit the angiogenesis by up-regulating the expression of miR-155, thereby exerting an anti-tumor effect on pancreatic cancer. Wang et al¹⁸ revealed that under oxidative stress, K-Ras gene activation can significantly up-regulate the expression of the oncogene miR-155, and miR-155 can inhibit the expression of Foxo3a and reduce the level of SOD and catalase, and the proliferative capacity of pancreatic cancer cells was significantly enhanced. This study combines the targeted regulatory relationship between miR-155 and SOCS3, revealing that miR-155 plays a role in regulating SOCS3 expression, affecting JAK-STAT pathway activity, and pancreatic cancer cell proliferation and apoptosis. However, whether the role of miR-155 targeting SOCS3 in regulating the biological effects of pancreatic cancer cells *in vivo* is unclear, and requires further investigation using animal models in the future.

Conclusions

We showed that the increase of miR-155 expression and the decrease of SOCS3 expression are related to the pathogenesis of pancreatic cancer. miR-155 can inhibit the proliferation and apoptosis of pancreatic cancer cells by targeting the inhibition of SOCS3 expression.

Conflict of Interests

The authors declare no conflict of interest

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