

MiR-155 regulates lymphoma cell proliferation and apoptosis through targeting SOCS3/JAK-STAT3 signaling pathway

X.-D. LI, X.-M. LI, J.-W. GU, X.-C. SUN

Dental Department, Yan'an People's Hospital of Shaanxi Province, Yan'an, Shaanxi, China

Abstract. – **OBJECTIVE:** Janus kinase (JAK)-signal transducer and activator of transcription (STAT) signaling pathway participates in regulating cell proliferation, differentiation, and apoptosis, and related to lymphoma. Suppressors of cytokine signaling 3 (SOCS3) is a negative regulator of the JAK-STAT signaling pathway. SOCS3 reduction and miR-155 up-regulation are associated with lymphoma pathogenesis. Bioinformatics analysis showed the complementary binding site between miR-155 and SOCS3. This study aimed to investigate the role of miR-155 in regulating SOCS3/JAK-STAT signaling pathway and affecting diffuse large B cell lymphoma (DLBCL) cell proliferation and apoptosis.

PATIENTS AND METHODS: DLBCL tumor sample was collected from the patients in our hospital. Lymphatic tissue derived from reactive lymphoid hyperplasia patients were selected as control. MicroRNA-155 (MiR-155) and SOCS3 expressions were detected. Dual luciferase assay was used to verify the targeted relationship between miR-155 and SOCS3. OCI-LY10 cells were cultured in vitro and divided into four groups, including miR-NC, miR-155 mimic, pIRES2-Blank, pIRES2-SOCS3, and miR-155 mimic + pIRES2-SOCS3 group. SOCS3, p-JAK, p-JAK2, p-STAT3, and Survivin expressions were tested. Cell apoptosis and proliferation were detected by flow cytometry.

RESULTS: MiR-155 expression significantly increased while SOCS3 level declined in DLBCL tissue compared with control. MiR-155 targeted reduced SOCS3 expression. MiR-155 inhibitor and pIRES2-SOCS3 transfection markedly up-regulated SOCS3 expression, reduced p-JAK1, p-JAK2, p-STAT3, and Survivin levels, attenuated cell proliferation, and enhanced cell apoptosis in OCI-LY10 cells.

CONCLUSIONS: Down-regulation of miR-155 inhibits DLBCL cell proliferation and facilitated apoptosis through up-regulating SOCS3 expression to suppress JAK-STAT3 signaling pathway.

Key Words:

MiR-155, SOCS3, JAK-STAT3, Lymphoma, Proliferation, Apoptosis.

Lymphoma is a kind of malignant tumor that takes place in the lymphoid and hematological system, such as lymph nodes, spleen, thymus, and extralymphatic tissues and organs^{1,2}. Lymphoma is associated with immune cell canceration, thus belonging immune system malignant tumor. Lymphoma contains Hodgkin's lymphoma (HL) and non-Hodgkin's lymphoma (NHL). NHL is the major type accounting for more than 90%³. Although tumor development has been achieved by surgical resection, whole body standard chemotherapy, local radiotherapy, and biological immunotherapy, about 60% patients still obtain non-ideal curative effect^{4,5}.

Janus kinase (JAK)-signal transducer and activator of transcription (STAT) signaling pathway excessive activation is closely related to multiple cancers occurrence, development, invasion, and metastasis by promoting cell proliferation and antagonizing apoptosis⁶⁻⁹. Suppressors of cytokine signaling 3 (SOCS3) is one of the members of SOCS family that can block cytokines response to negatively regulate JAK-STAT signaling pathway. SOCS3 down-regulation is associated with tumor occurrence, metastasis, drug resistance, and poor prognosis¹⁰⁻¹³. SOCS3 expression significantly reduced in NHL tissue, indicating that SOCS3 plays a tumor suppressor gene role in NHL¹⁴. MiRNA is a type of endogenous single-stranded noncoding RNA at the length of 22-25 nt discovered from eukaryote. It plays a degrading or inhibiting role on mRNA by binding with the 3'-UTR. MiRNA that accounts for 1% gene amount regulates more than 30% of human gene expressions¹⁵. Numerous studies revealed that miRNA abnormal expression and function play a crucial role in multiple tumor pathogeneses, including lymphoma^{16,17}. MiR-155 was found to play

an oncogene role in lymphoma¹⁸ and related to lymphoma disease degree and poor prognosis¹⁹. Bioinformatics analysis showed the complementary binding site between miR-155 and SOCS3. This study aimed to investigate the role of miR-155 in regulating SOCS3/JAK-STAT signaling pathway and affecting lymphoma cell proliferation and apoptosis.

Patients and Methods

Main Reagents and Materials

Human diffuse large B cell lymphoma (DLBCL) OCI-LY10 cells were purchased from Shanghai Yubo biological technology co., Ltd. (Shanghai, China). RPMI-1640 medium, Opti-MEM, penicillin, and streptomycin were purchased from Gibco BRL. Co. Ltd. (Grand Island, NY, USA). FBS was got from Gemini Bio Products (Woodland, CA, USA). EasyPure RNA Kit and Real-time PCR reagent TransScript Green One-Step qRT-PCR SuperMix were obtained from TransGen Biotech (Beijing, China). Mouse anti-human Survivin, JAK1, p-JAK1, JAK2, and p-JAK2 antibodies were purchased from Abcam (Cambridge, MA, USA). Rabbit anti-human STAT3, phosphorylated STAT3 (p-STAT3), SOCS3, and β -actin antibodies were both from Cell Signaling Technology (Beverly, MA, USA). Goat anti-rabbit and anti-mouse secondary antibodies were got from Bio-Rad Laboratories (Hercules, CA, USA). Annexin-V/PI apoptosis detection kit was from Affinity BioReagent (Hangzhou, China). Cell proliferation detection kit Click-iT[®] DU Alexa Fluor 488 Flow cytometry Assay Kit was obtained from Molecular Probes (Eugene, OR, USA). Dual-Glo[®] Luciferase Assay System and pMIR luciferase reporter gene plasmids were provided by Promega (Madison, WI, USA). Lipofectamine[®] was provided from Invitrogen Life Technologies (Carlsbad, CA, USA).

Cell Transfection

A total of 20 DLBCL patients received treatment between May 2016 and Oct 2016 in Yan'an People's Hospital of Shaanxi Province and another 32 patients suffered from reactive lymphoid hyperplasia were enrolled. The lymphatic tissue and tumor tissue were collected. The study was approved by the Ethics Committee of Yan'an People's Hospital of Shaanxi Province. All the patients signed the informed consent.

Cell Culture

OCI-LY10 cells were cultured in RPMI-1640 medium containing 10% fetal bovine serum (FBS) and 1% penicillin-streptomycin. The cells were passaged at 1:4.

Dual-Luciferase Reporter Gene Assay

The PCR products containing the full length of SOCS3 gene 3'-UTR segment were cloned to pMIR. Next, it was transfected into DH28 competent cells and sequenced to select the plasmids with correct sequence. Then, pMIR-SOCS3 3'-UTR-wt (or pMIR-SOCS3 3'-UTR-mut) was co-transfected with HEK293 cells using Lipofectamine 2000 together with miR-155 mimic (or miR-NC, or miR-155 inhibitor). The luciferase activity was detected according to the Dual-Glo[®] Luciferase Assay method after cultured for 48 h.

SOCS3 Overexpression Plasmid Construction

The CDS region fragment of SOCS3 gene was amplified and connected to pIRES2 plasmid after digested by XbaI and BamH I. Then it was transformed into DH28 competent cells and screened by penicillin. Next, the plasmid was extracted and sequenced. It was named as pIRES2-SOCS3, and pIRES2-Blank was treated as control.

Cell Transfection and Grouping

OCI-LY10 cells were cultured *in vitro* and divided into five groups, including miR-NC, miR-155 inhibitor, pIRES2-Blank, pIRES2-SOCS3, and miR-155 + pIRES2-SOCS3 groups. Nucleotide fragments and Lipofectamine 2000 were added to Opti-MEM and incubated at room temperature for 30 min, respectively. Then they were added to the cells cultured in Opti-MEM. After 6 h incubation, the medium was changed back to RPMI 1640 medium containing 10% FBS and 1% penicillin-streptomycin. The cells were used for detection after 72 h.

qRT-PCR

Total RNA was extracted using EasyPure RNA Kit and adopted for PCR reaction by TransScript Green One-Step qRT-PCR SuperMix. The reaction system contained 1 μ g RNA template, 0.3 μ M primers, 10 μ l 2 \times TransStart Tip Green qPCR SuperMix, 0.4 μ l RT Enzyme Mix, 0.4 μ l Dye II, and ddH₂O. The reverse transcription condition was 37°C for 15 min and 98°C for 5 min. The PCR reaction was composed of 45°C reverse transcription for 5 min, 94°C pre-denaturation for 30 s, followed

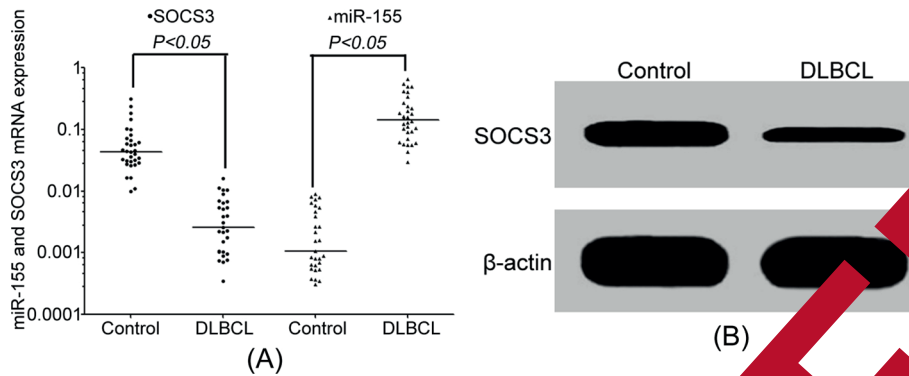


Figure 1. MiR-155 up-regulated, while SOCS3 reduced in DLBCL tissue. (A) qRT-PCR detection of miR-155 and SOCS3 mRNA expressions. (B) Western blot detection of SOCS3 protein expression.

by 40 cycles of 94°C for 5 s and 60°C for 30 s. Real-time PCR was performed on Bio-Rad CFX96/CFX connect to test the relative expression.

Western Blot

Total protein was extracted by RIPA from cells. A total of 40 μg protein was separated by

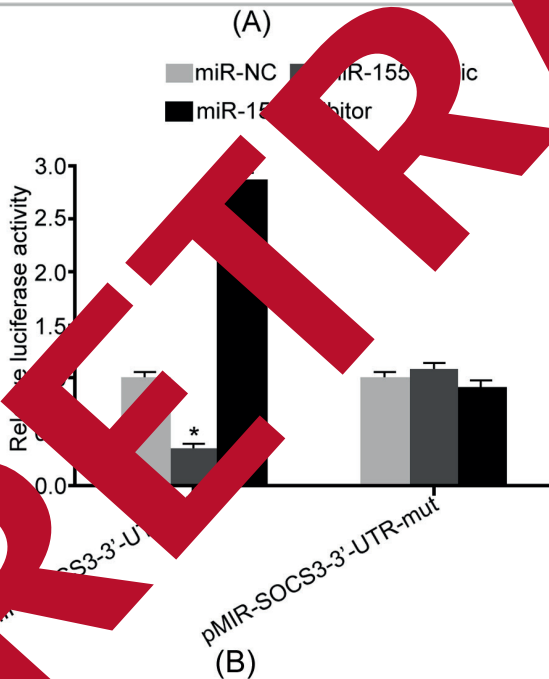
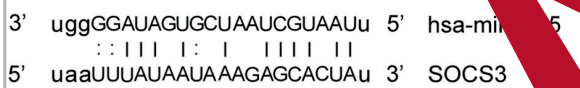


Figure 2. MiR-155 targeted regulated SOCS3 expression. (A) The binding site between miR-365 the 3'-UTR of DJ-1 mRNA. (B) Dual luciferase assay. *p < 0.05, compared with miR-NC.

8%-10% sodium lauryl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) for 3 h and transferred to membrane. Next, the membrane was blocked and incubated with primary antibody at 4°C overnight (SOCS3, JAK1, p-JAK1, JAK2, p-JAK2, STAT3, and STAT3, Survivin, and β-actin at 1:1000, 1:2000, 1:800, 1:2000, 1:800, 1:2000, and 1:10000, respectively). Then, the membrane was incubated with horseradish peroxidase (HRP) labeled secondary antibody (1:10000) for 60 min after washed by phosphate buffered saline-tween 20 (PBS-T) for three times. At last, the protein expression was detected by enhanced chemiluminescence (ECL).

Edu Staining

The cells were added to 10 μM EdU solution at 37°C for 120 min. After incubated for 48 h, the cells were digested by trypsin and collected. After washed with phosphate-buffered saline (PBS) containing 1% bovine serum albumin (BSA) and fixed in 100 μl Click-iT® fixative for 15 min, the cells were penetrated by 100 μl 1×Click-iT® saponin-based permeabilization and wash reagent at room temperature for 15 min. Next, the cells were incubated in 500 μl reaction liquid containing PBS, CuSO₄, Alexa Fluor 488, and Buffer additive at room temperature for 30 min. Then the cells were washed by 3 ml 1×Click-iT® saponin-based permeabilization and wash reagent for 1 time. At last, the cells were resuspended in 500 μl 1×Click-iT® saponin-based permeabilization and wash reagent and tested by Beckmann FC 500 MCL/MPL flow cytometry.

Flow Cytometry

The cells were resuspended in 500 μl binding buffer and incubated in 5 μl Annexin V-FITC and

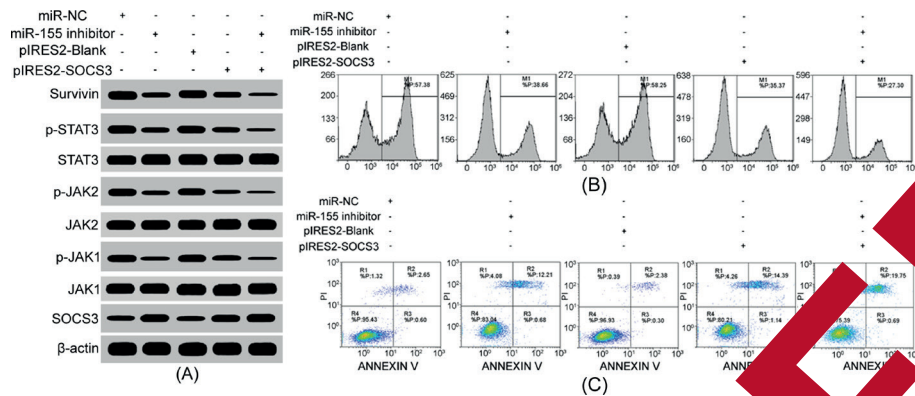


Figure 3. Down-regulation of miR-155 attenuated JAK-STAT3 signaling pathway activity, promoted cell apoptosis and inhibited proliferation. (A) Western Blot detection of protein expression. (B) Flow cytometry detection of cell proliferation. (C) Flow cytometry detection of cell apoptosis.

5 μl PI avoid of light for 15 min. Next, the cells were added with 5 μl PI and tested on Beckmann FC 500 MCL/MPL flow cytometry to evaluate cell apoptosis.

Statistical Analysis

All data analyses were performed by the SPSS 18.0 software (Chicago, IL, USA). The mean and standard deviation data were depicted as mean ± standard deviation and compared by *t*-test. *p*<0.05 was considered as statistical significance.

Down-regulation of miR-155 Attenuated JAK-STAT3 Signaling Pathway Activity, Promoted Cell Apoptosis and Inhibited Proliferation

miR-155 inhibitor and/or pIRES2-SOCS3 transfection significantly up-regulated SOCS3 expression, reduced p-JAK1, p-JAK2, p-STAT3, and Survivin levels (Figure 3A), attenuated cell proliferation (Figure 3B), and enhanced cell apoptosis (Figure 3C) in OCI-LY10 cells.

Results

MiR-155 Up-Regulated in DLBCL Tissue While SOCS3 Reduced in DLBCL Tissue

Quantitative RT-PCR (qRT-PCR) showed that SOCS3 mRNA significantly reduced while miR-155 elevated in DLBCL tissue from DLBCL patients compared with control (Figure 1A). Western blot revealed that SOCS3 protein markedly reduced in DLBCL tissue compared with control, indicating miR-155 up-regulation may play a role in down-regulating SOCS3 and promoting DLBCL pathogenesis (Figure 1B).

MiR-155 Mimics or Inhibitors Regulated SOCS3 Expression

miR-155 mimics or inhibitors transfection significantly declined or increased the relative luciferase activity of HEK293 cells, indicating the regulatory relationship between miR-155 and SOCS3 mRNA.

Discussion

JAK-STAT signaling pathway is first found in interferon study. It is discovered to be activated by multiple cytokines and growth factors, and mediates various cytokines signal transduction²⁰. JAK-STAT signaling pathway is mainly composed of tyrosine kinase associated receptor, STAT, and JAK⁶.

JAK-STAT signaling pathway activation is associated with tumor pathogenesis by up-regulating Bcl-2, Survivin, Cyclin D1, and c-Myc^{21,22}. As an oncogene transcription factor, STAT3 is an important member of STAT family that plays a critical role in a variety of cancer pathogenesis^{23,24}. JAK-STAT3 participates in multiple biological processes, such as cell proliferation, cycle, and apoptosis, thus, is associated with tumor occurrence, development, invasion, and metastasis, including lung cancer⁶, colorectal cancer⁹, prostate cancer⁷, and pancreatic cancer⁸. As the strongest member of SOCS family, SOCS3 plays a negative regulatory role in JAK-STAT3 signaling pathway by inhibiting JAK kinase binding with receptor to suppress JAK phosphorylation or

competitive binding with JAK to restrain STAT3 phosphorylation via N-terminal kinase inhibitory region (KIR)^{11,13}. SOCS3 is related to multiple cancers' occurrence, metastasis, drug resistance, and poor prognosis, such as breast cancer¹¹, prostate cancer¹³, lung cancer¹², and pancreatic cancer¹⁰. SOCS3 expression significantly reduced in NHL tissue, indicating that SOCS3 plays a tumor suppressor gene role in NHL¹⁴. MiR-155 was found to play an oncogene role in lymphoma¹⁸ and related to lymphoma disease degree and poor prognosis¹⁹. Bioinformatics analysis showed the complementary binding site between miR-155 and SOCS3. This research aimed to investigate the role of miR-155 in regulating SOCS3/JAK-STAT signaling pathway and affecting lymphoma cell proliferation and apoptosis.

Our investigation showed that miR-155 mimics or inhibitor transfection significantly declined or increased the relative luciferase activity of HEK293 cells, indicating the regulatory relationship between miR-155 and SOCS3 mRNA. SOCS3 significantly reduced, while miR-155 elevated in the tumor tissue from DLBCL patients compared with control, indicating miR-155 up-regulation may play a role in reducing SOCS3 and promoting DLBCL pathogenesis. Slezacek and Prochazka et al²⁵ reported that miR-155 expression in B-cell lymphoma abnormally up-regulated. Zhong et al²⁶ demonstrated that miR-155 increased in DLBCL compared with healthy lymphatic hyperplasia and was correlated with complete remission, overall response rate, and poor prognosis. Munch-Petersen et al²⁷ reported that miR-155 overexpressed limit in tumor tissue of DLBCL by nucleic acid hybridization in situ technique. Huang et al²⁸ revealed that miR-155 level markedly elevated in activated cell-like (ABC) DLBCL tissue compared with germinal center B-cell-like (GCL) DLBCL tissue, lower invasion and malignancy. Moreover, miR-155 expression was significantly higher in OCI-Ly3 and OCI-Ly10 compared with that in SU-DHL6 and SU-DHL16. Merkel et al¹⁸ exhibited that miR-155 increased in lymphoma tissue and played an oncogene role. In this study, miR-155 content was significantly higher in DLBCL tissues, suggesting that miR-155 is an oncogenic factor, which is similar to Zhong et al²⁶ and Munch-Petersen et al²⁷. Molavi et al¹⁴ showed that promoter methylation enhanced SOCS3, while SOCS3 mRNA expression reduced in B-cell lymphoma cell line Jeko, Mino, and Rec-1 compared with healthy bone marrow mononuclear cells, which is in accordance with

our results. Further detection demonstrated that miR-155 inhibitor and/or SOCS3 over-expression plasmid transfection markedly upregulated SOCS3 expression, reduced p-JAK1, p-STAT3, and Survivin levels, attenuated cell proliferation, and enhanced cell apoptosis in OCI-LY10 cells. Huang et al²⁸ found that miR-155 targeted inhibited PIK3R1 (p85) expression and enhanced PI3K-AKT signaling pathway, whereas down-regulation of miR-155 attenuated PI3K-AKT signaling pathway and restrained BCL cell line OCI-Ly3 and SU-DHL16 proliferation. Sandhu et al²⁹ revealed that miR-155 promoted OCI-Ly3 cell proliferation, enhanced colony formation, and inhibited apoptosis by targeting HDAC4 expression. Dagan et al¹⁹ discovered that miR-155 overexpression facilitated lymphoma cell migration and chemotactic ability by inhibiting HGF expression, which may promote lymphoma cell dissemination, progression, invasion, and pathogenesis. Merkel et al¹⁸ demonstrated that inhibition of miR-155 suppressed lymphoma cell proliferation *in vitro* and *in vivo*. In this work, miR-155 down-regulation markedly attenuated OCI-Ly3 malignancy. Molavi et al¹⁴ reported that SOCS3 over-expression apparently suppressed JAK-STAT3 phosphorylation, downregulated BCL2, BCL-XL, BCL1, BCL2, and BCL-XL expressions, and induced apoptosis in B-cell lymphoma cell line Mino and Rec-1. At present, the role of miR-155 in regulating SOCS3 expression and DLBCL cell proliferation and apoptosis has not been reported. This investigation showed that miR-155 plays a role in suppressing SOCS3 expression and promoting DLBCL pathogenesis. Down-regulation of miR-155 increased SOCS3 expression and blocked JAK-STAT3 signal transduction to inhibit lymphoma cell proliferation and accelerated cell apoptosis. This study only collected DLBCL tissue and investigated the impact of miR-155 on SOCS3-JAK-STAT3 signaling pathway and DLBCL cell proliferation and apoptosis, while whether miR-155 may regulate other types of B-cell lymphoma is still unclear.

Conclusions

MiR-155 over-expression plays a role in inhibiting SOCS3 and promoting DLBCL pathogenesis. Down-regulation of miR-155 inhibited DLBCL cell proliferation and facilitated apoptosis through up-regulating SOCS3 expression to suppress JAK-STAT3 signaling pathway.

Acknowledgments

This work was supported by the Yanan Department of Science and Technology Plan Item (2011ks-05).

Conflict of interest

The authors declare no conflicts of interest.

References

- CHENG H, TANG X, CHENG J, ZHANG B, ZHANG YL, WANG WO, TENG P. Pathologic character and diagnosis of female primary genital system diffuse large B cell lymphoma. *Eur Rev Med Pharmacol Sci* 2017; 21: 1471-1476.
- SMEDBY KE, HJALGRIM H. Epidemiology and etiology of mantle cell lymphoma and other non-Hodgkin lymphoma subtypes. *Semin Cancer Biol* 2011; 21: 293-298.
- SKRABEK P, TURNER D, SEFTEL M. Epidemiology of non-Hodgkin lymphoma. *Transfus Apher Sci* 2013; 49: 133-138.
- GHOSH I, BAKHSI S. Jaundice as a presenting manifestation of pediatric non-Hodgkin lymphoma: etiology, management, and outcome. *J Pediatr Hematol Oncol* 2010; 32: e131-135.
- XAVIER AC, ARMESON KE, HILL EG, COSTA LJ. The outcome of non-Hodgkin lymphoma among classical Hodgkin lymphoma survivors. *Cancer* 2013; 119: 3385-3392.
- LI HX, ZHAO W, SHI Y, LI YN, ZHANG LS, ZHANG M, WANG D. Retinoic acid amide inhibits the JAK/STAT3 pathway in lung cancer which leads to apoptosis. *Tumour Biol* 2015; 36: 8671-8678.
- LIU X, HE Z, LI CH, HUANG Q, QI DING C. Correlation analysis of JAK-STAT3 signaling pathway on prognosis of patients with prostate cancer. *Pathol Oncol Res* 2012; 18: 17-23.
- MACHA MA, SINGH S, GUPTA S, PANDYANONNUSAMY MP, BATRA SK, JAIN SK. Suggulsterone decreases proliferation and metastatic behavior of pancreatic cancer cells by modulating JAK/STAT and Src/FAK signaling. *Cancer Lett* 2013; 341: 166-177.
- METERY ML, HUNDGREEN A, KADLUBAR SA, BONDURANT WOLFF F. JAK/STAT/SOCS-signaling pathway in prostate and rectal cancer. *Mol Carcinog* 2013; 42: 155-166.
- HUANG Y, LI B, LIU W, WU J, JIANG W, CHEN C, YANG Z, ZENG Y, HU G, WANG X. Transcriptional repression of SOCS3 mediated by IL-6/STAT3 signaling via DNMT1 promotes pancreatic cancer cell proliferation and metastasis. *J Exp Clin Cancer Res* 2016; 35: 27.
- BARCLAY JL, ANDERSON ST, WATERS MJ, CURLEWIS JD. SOCS3 as a tumor suppressor in breast cancer cells, and its regulation by PRL. *Int J Cancer* 2009; 124: 1756-1766.
- LIN YC, LIN CK, TSAI YH, WENG HH, LI YC, YOU L, CHEN JK, JABLONS DM, YANG CT. Adenovirus-mediated SOCS3 gene transfer inhibits the growth and enhances the radiosensitivity of human non-small cell lung cancer cells. *Oncol Rep* 2010; 24: 1605-1612.
- PIERCONTI F, MARTINI M, PINTO F, CENCI T, CAPODIMONTE S, CALARCO A, BASSI PF, LAROCCA LM. Epigenetic silencing of SOCS3 identifies a subset of prostate cancer with an aggressive behavior. *Prostate* 2011; 71: 318-325.
- MOLAVI O, WANG P, ZAK Z, GELEBARTI A, LAI R. Gene methylation and silencing of SOCS3 in mantle cell lymphoma. *Br J Haematol* 2013; 161: 235-236.
- WANG K, XU Z, WANG H, XU T, ZHU M. Methylation and gene networks in human diffuse large B-cell lymphoma. *Oncol Rep* 2014; 31: 2215-2232.
- TAGAWA H. microRNA in mantle lymphoma. *Adv Exp Med Biol* 2015; 889: 1-12.
- HOAREAU M, MERKEL O, MORIGGL R, GRIEBEN F. MicroRNA-155-positive anaplastic large cell lymphoma. *Hum Biosci (Schol Ed)* 2015; 7: 217-225.
- LIU D, HAMACHER J, FICHTL R, GRABNER L, SCHIEBERER M, PRUTSCH N, BAERLEGGER G, SCHLEDERER M, KRENN PW, HARTMANN TN, SIMONITSCH-KLUPP I, PLASS C, STABER PB, MORIGGL R, TURNER SD, GREIL R, KENNER L. Oncogenic role of miR-155 in anaplastic large cell lymphoma lacking the t(2;5) translocation. *J Hematol Oncol* 2015; 8: 445-456.
- ALBERTINI M, MORTARINO M, AVALLONE G, GIOIA G, COMAZZI S, ROCCABIANCA P. The expression ratio of miR-17-5p and miR-155 correlates with grading in primary splenic lymphoma. *Vet Immunol Immunopathol* 2013; 155: 117-123.
- WU Z, HUANG W, CHEN B, BAI PD, WANG XG, XING JC. Up-regulation of miR-124 inhibits invasion and proliferation of prostate cancer cells through mediating JAK-STAT3 signaling pathway. *Eur Rev Med Pharmacol Sci* 2017; 21: 2338-2345.
- PENCIK J, PHAM HT, SCHMOELLERL J, JAVAHERI T, SCHLEDERER M, CULIG Z, MERKEL O, MORIGGL R, GREBIEN F, KENNER L. JAK-STAT signaling in cancer: From cytokines to non-coding genome. *Cytokine* 2016; 87: 26-36.
- KWON EM, HOLT SK, FU R, KOLB S, WILLIAMS G, STANFORD JL, OSTRANDER EA. Androgen metabolism and JAK/STAT pathway genes and prostate cancer risk. *Cancer Epidemiol* 2012; 36: 347-353.
- Ji K, ZHANG M, CHU Q, GAN Y, REN H, ZHANG L, WANG L, LI X, WANG W. The role of p-STAT3 as a prognostic and clinicopathological marker in colorectal cancer: a systematic review and meta-analysis. *PLoS One* 2016; 11: e0160125.
- BANERJEE K, RESAT H. Constitutive activation of STAT3 in breast cancer cells: A review. *Int J Cancer* 2016; 138: 2570-2578.
- SLEZAK-PROCHAZKA I, KLUIVER J, DE JONG D, SMIGIELSKA-CZEPIEL K, KORTMAN G, WINKLE M, RUTGERS B, KOERTS J, VISSER L, DIEPSTRA A, KROESEN BJ, VAN DEN BERG A. Inhibition of the miR-155 target NIAM phenocopies the growth promoting effect of miR-155 in B-cell lymphoma. *Oncotarget* 2016; 7: 2391-2400.
- ZHONG H, XU L, ZHONG JH, XIAO F, LIU Q, HUANG HH, CHEN FY. Clinical and prognostic significance

of miR-155 and miR-146a expression levels in formalin-fixed/paraffin-embedded tissue of patients with diffuse large B-cell lymphoma. *Exp Ther Med* 2012; 3: 763-770.

- 27) MUNCH-PETERSEN HD, RALFKIAER U, SJO LD, HOTHER C, ASMAR F, NIELSEN BS, BROWN P, RALFKIAER E, GRONBAEK K. Differential expression of miR-155 and miR-21 in tumor and stroma cells in diffuse large B-cell lymphoma. *Appl Immunohistochem Mol Morphol* 2015; 23: 188-195.
- 28) HUANG X, SHEN Y, LIU M, BI C, JIANG C, IOBAL J, MCKEITHAN TW, CHAN WC, DING SJ, FU K. Quantitative proteomics reveals that miR-155 regulates the PI3K-AKT pathway in diffuse large B-cell lymphoma. *Am J Pathol* 2012; 181: 26-33.
- 29) SANDHU SK, VOLINIA S, COSTINEAN S, GALASSO M, NEINAST R, SANTHANAM R, PARTHUN MR, PERROTTI D, MARCUCCI G, GARZON R, CROCE CM. miR-155 inhibits histone deacetylase 4 (HDAC4) and restores transcriptional activity of B-cell lymphoma 6 (BCL6) in the Emu-miR-155 transgenic mouse model. *Proc Natl Acad Sci USA* 2012; 109: 20051-20052.
- 30) DAGAN LN, JIANG X, BHATT S, SUBEDO V, WILKINSKY K, LOSSOS IS. miR-155 regulates HGAL expression and increases lymphoma cell motility. *Blood* 2012; 119: 513-520.

RETRACTED