Study on MKNK2 as a potential prognostic and immunological biomarker in pan-cancer

Y.-M. ZHANG¹, J.-K. FAN¹, X.-Y. WANG¹, J. LIU¹, T. LI¹, X.-S. WANG¹, X.-J. YANG^{1,2}

¹Department of Neurosurgery, Tianjin Medical University General Hospital, Tianjin, China ²Department of Neurosurgery, Beijing Tsinghua Changgung Hospital, Beijing, China

Abstract. – OBJECTIVE: This study aimed to investigate the expression levels of the MKNK2 gene in pan-cancer, its prognostic significance, and its relationship with the tumor immune microenvironment, as well as to assess its potential as an immunological and prognostic biomarker.

MATERIALS AND METHODS: The research utilized data from The Cancer Genome Atlas (TCGA), Genotype-Tissue Expression (GTEx), and Cancer Cell Line Encyclopedia (CCLE), including clinical and mutational information. Bioinformatic tools were employed to analyze the association of MKNK2 with carcinogenesis, including its links to prognosis, immune cell infiltration, tumor immune microenvironment, gene mutation, and the stemness of various tumor cells. A variety of statistical software and analytical tools were applied, including R software, SPSS 27.0, TIMER, CIBERSORT algorithm, and EPIC algorithm.

RESULTS: The study found that MKNK2 is abnormally expressed in pan-cancer and is associated with a poor prognosis. The levels of MKNK2 are highly related to immune cell infiltration and tumor stemness. Notably, in liver hepatocellular carcinoma, glioblastoma multiforme, low-grade gliomas, and acute myeloid leukemia, MKNK2 expression shows a strong correlation with clinical outcomes and immune infiltration. Furthermore, the expression of MKNK2 shows significant correlations with immune cell infiltration, immune checkpoints, tumor mutational burden (TMB), microsatellite instability (MSI), and stemness scores across various cancers.

CONCLUSIONS: The abnormal expression of MKNK2 is associated with tumor progression, immune checkpoint genes, immune cell infiltration, microsatellite instability (MSI), tumor mutational burden (TMB), and stemness in a variety of tumors, especially in glioblastoma multiforme low-grade gliomas (GBMLGG). Therefore, MKNK2 may serve as a potent prognostic physiological marker and provide new avenues for the development of tumor mechanisms and therapeutic strategies targeting MKNK2 to enhance the efficacy of immunotherapy.

Key Words:

MKNK2, Pan-cancer, Cancer Genome Atlas, Immune analysis, Tumor microenvironment.

Introduction

Cancer continues to be a significant challenge to global health, with a persistent rise in both its incidence and mortality rates. The World Cancer Report by the World Health Organization (WHO) for the year 2020 underscores the severity of this issue, highlighting that cancer accounts for one in every six deaths annually, with an ever-increasing burden on society and healthcare systems worldwide¹. The urgency of improving cancer detection and therapeutic strategies is evident, given the high mortality rate associated with the disease.

Despite advances in clinical treatments, the overall prognosis for many cancer patients remains suboptimal due to the development of drug resistance, debilitating side effects, and other treatment-related complications. The quest for novel therapeutic targets and more sensitive physiological biomarkers is, therefore, more critical than ever to enhance early diagnosis and treatment efficacy².

MKNK2 encodes a protein that is part of the calcium/calmodulin-dependent protein kinases (CAMK) family of Ser/Thr protein kinases, which are members of the broader protein kinase superfamily³. Encoded by the MKNK2 gene, this protein kinase is a crucial downstream effector in the MAPK signaling cascade, which is instrumental in various cellular functions, including the promotion of oncogenic transformation, mRNA translation, and cell proliferation upon stimulation by mitogen-activated protein kinases $(MAPK)^4$. The downstream signaling proteins, MAPK-interacting kinases (MKNK1/2), have been observed to modulate the synthesis of both non-cancerous and cancerous cell proteins, indicating their potential role in cancer development⁵. Emerging evidence has demonstrated elevated *MKNK2* expression in a variety of cancers, particularly in drug-resistant ovarian cancer tissues, where it correlates with poor prognosis^{6,7}. However, the comprehensive role of *MKNK2* in pan-cancer progression and its underlying mechanisms are not yet fully understood, warranting further investigation.

The nexus between cancer stem cells (CSCs) and the tumor microenvironment (TME) has increasingly become a focal point in cancer research. The TME, which fosters cancer growth and metastasis, is intricately connected with CSCs, emphasizing the importance of understanding the interplay between these components in cancer development and progression^{8,9}. MKNK2's association with the maintenance of stemness in multiple cancers suggests a complex interaction within the TME.

The presence of immune cells within the TME, such as B cells, tumor-associated macrophages, regulatory T cells, and natural killer cells, is increasingly recognized for its role in cancer formation and immune evasion^{10,11}. Suppressive molecules like programmed cell death ligand-1 and cytotoxic T-lymphocyte associated antigen-4 are pivotal in modulating anti-tumor immunity. Despite the promise of cancer immunotherapy, there is a pressing need to refine existing approaches and explore new targets to enhance the efficacy of immunotherapeutic interventions.

In this study, we delve into the expression patterns of MKNK2 across diverse non-cancerous and cancerous tissues and cell lines, aiming to systematically elucidate its incidence and prognostic value in various cancer types. By integrating data from authoritative databases, such as The Cancer Genome Atlas (TCGA), Genotype-Tissue Expression (GTEx), and Tumor Immune Estimation Resource (TIMER), we have conducted a comprehensive evaluation of MKNK2's role in immune responses and its prognostic implications in a spectrum of cancers. The potential interplay between MKNK2, tumor mutational burden (TMB), microsatellite instability (MSI), immune cell infiltration, stemness, and genetic immune checkpoints was scrutinized. Furthermore, Gene Set Enrichment Analysis (GSEA) was utilized to explore the biological pathways and functions related to MKNK2. Our findings reveal a significant association between MKNK2 expression and patient survival, particularly emphasizing its prognostic value in glioblastoma multiforme lowgrade gliomas (GBMLGG), liver hepatocellular carcinoma (LIHC), and acute myeloid leukemia (LAML). These insights underscore the potential

of *MKNK2* as a generalized predictor for cancer diagnosis and prognosis and highlight its therapeutic potential in the realm of immunotherapy.

Materials and Methods

Data Collection and Processing

A pan-cancer dataset was downloaded from the TARGET, GTEx, UCSC (https://xenabrowser. net/) database, and TCGA Pan-Cancer (PANCAN, N=10,535, G=60,499). Then, the data of MKNK2 gene levels [(ENSG00000099875 (MKNK2)] of each sample, along with the data of primary solid tumor, normal tissue, and primary blood-derived cancer-bone marrow, were extracted. The specimens with 0 expression level were filtered along with studies with <3 samples of single cancer, followed by the determination of log2(x+0.001) transformation per expression value. Finally, the expression data of 26 cancer species from UCSC and TCGA and 34 from TARGET and GTEx were selected. Cell line gene expression matrix of multiple cancer species was obtained from CCLE (https:// portals.broadinstitute.org/ccle/about) data set and transformed by log2 (TPM +1) using R package of "rma" in an R environment (R version: 3.6.1).

Genetic Expression Levels in All Tumor Stages

The R software (version 3.6.4) was utilized to determine the variability in the genetic expression of each sample during different tumor stages. The unpaired Student's *t*-test was performed to evaluate the significant difference of inter-pair analysis. Variance analysis was conducted to determine the difference in multiple sample groups.

Cox Regression and Kaplan-Meier Survival Analyses

Data from a previous TCGA high-quality prognostic investigations published in the cell and that from the UCSC cancer browser (https://xenabrowser.net/datapages/), along with the follow-up data from TARGET (as a supplement) were also obtained¹². Cox regression analysis was carried out to evaluate *MKNK2* expression association with patients' overall survival (OS), disease-specific survival (DSS), disease-free interval (DFI), and Progression Free Interval (PFI) in each kind of cancer from GTEx and TCGA samples in the R environment. The Kaplan-Meier assessment was applied to obtain pan-cancer patients' survival curves after separating the patients into

high and low *MKNK2* expression cohorts based on the chief separation strategy.

MKNK2 Expression Association with Immune Cells and Checkpoint

With the help of TIMER, the correlation of immune cell infiltration with MKNK2 expression was determined. It provided an important assessment of immune cell integration for TCGA samples' RNA sequencing. These cells involve: B, dendritic, macrophages, CD4+ T, neutrophils, CD8+ T, and cells. More than 60 common immune checkpoint genes were selected, and the link between the expression of these genes and MKNK2 was evaluated¹³. CIBERSORT algorithm and EPIC algorithm of immunity score were calculated using the deconvo CIBERSOR method and deconvo epic method of R software package IOBR, respectively. Algorithm and EPIC algorithm of immunity score were calculated using the deconvo CIBERSOR method and deconvo epic method of R software package IOBR, respectively. The R software package used in this document can be downloaded from the CRAN package repository (Available at: https://cran.r-project.org/web/packages/available packages by name.html).

Function Landscape and Stemness Scores Analyses

With the help of the R software package maftools (version 2.8.05), the function of TMB for each tumor was determined. The MSI scores of each tumor were also evaluated. The MSI and TMB scores were acquired from TCGA. Correlation analysis of TMB/MSI and MKNK2 expression was carried out using Spearman's protocol. The horizontal axis in the figure depicts the correlation coefficient between TMB/MSI and MKNK2; the vertical axis shows various cancer types, and the bar color depth represents the *p*-value significance. The relationship between tumor cell stemness and gene expression was analyzed by methylated signature and mRNA expression¹⁴. The DNA methylation-based stemness scores (DNAss) and the RNA-based stemness scores (RNAss) reflect epigenetic characteristics and gene expression characteristics of stem cells, respectively.

Gene Set Enrichment Analysis

The "gmt" file of the hallmark gene set (h.all. v7.4.symbols.gmt), which contains 50 hallmark gene sets, was downloaded from the website of Molecular Signatures Database (MSigDB, Available at: https://www.gsea-msigdb.org/gsea/index.jsp) and

used to calculate the normalized enrichment score (NES) and false discovery rate (FDR) of the DEGs between low- and high-*MKNK2* expression cancer groups for each biological process in each cancer type. The GSEA was conducted using the R package "clusterProfiler" (the clusterProfiler package is released under Artistic-2.0 License within Bioconductor project). The source code and vignette are freely available at http://bioconductor.org/packages/release/bioc/html/clusterProfiler.html, and the results were summarized in the bubble plot depicted by the R package "ggplot2".

Statistical Analysis

Data analysis was conducted using R (3.6.1 & 3.6.4 version) and SPSS 27.0 (Statistical Package for the Social Sciences; IBM Corp., Armonk, NY, USA); download at: https://www.ibm.com/cn-zh/ spss. To determine the association of MKNK2 expression with aimed targets, including immune cell infiltration scores (six aforementioned immune cell types), TMB, MSI, and methylation transferase genes, the Spearman Correlation test was applied. The intergroup comparison of MKNK2 expression levels or between tumor and normal tissues was determined by the paired or the simple *t*-test, based on whether the samples were paired. Multiple group comparisons were conducted using One-way analysis of variance, while Bonferroni adjustment was used for pairwise comparisons. A *p*-value <0.05 was deemed significant.

Results

Expression Levels of MKNK2 in Pan-Cancer

By employing the R software, we conducted a meticulous analysis to assess the variability in MKNK2 expression between tumor and normal samples across a diverse range of cancer types. Through the application of unpaired Wilcoxon Rank Sum and Signed Rank Tests, we meticulously evaluated the statistical significance of observed expression differences. This rigorous approach revealed that MKNK2 expression significantly diverged in 17 of the 26 examined cancer types. Particularly, we identified substantial upregulation in cancers such as glioblastoma (GBM), glioblastoma multiforme low-grade gliomas (GBMLGG), cervical squamous cell carcinoma (CESC), prostate adenocarcinoma (PRAD), uterine corpus endometrial carcinoma (UCEC), liver hepatocellular carcinoma (LIHC), kidney chromophobe (KICH), and cholangiocarcinoma (CHOL), along with nine additional tumor categories, including lung adenocarcinoma (LUAD), colon adenocarcinoma (COAD), stomach adenocarcinoma (STAD), colorectal adenocarcinoma with readthrough (COADREAD), esophageal squamous cell carcinoma (STES), lung squamous cell carcinoma (LUSC), head and neck squamous cell carcinoma (HNSC), thyroid carcinoma (THCA), and rectal adenocarcinoma (READ) (Figure 1A).

By expanding our analysis to encompass 34 cancer types, we observed a generally consistent expression pattern. However, certain cancers, including prostate adenocarcinoma (PRAD) and liver hepatocellular carcinoma (LIHC), stood out with significant upregulation, in contrast to the marked downregulation observed in cancers, such as breast invasive carcinoma (BRAC), esophageal carcinoma (ESCA), wild-type skin cutaneous melanoma (WT SKCM), pancreatic adenocarcinoma (PAAD), testicular germ cell tumors (TGCT), and uterine carcinosarcoma (UCS) (Figure 1B). These findings highlight the considerable variability in MKNK2 expression levels between tumor and non-tumor tissues and underscore the complexity of its role across different cancer types.

Further investigation of MKNK2 expression across all tumor stages (I-IV) in patients revealed notable differences in 11 tumor types, including cervical squamous cell carcinoma (CESC), esophageal carcinoma (ESCA), stomach squamous cell carcinoma (STES), kidney papillary cell carcinoma (KIPAN), uterine corpus endometrial carcinoma (UCEC), thymoma (THYM), thyroid carcinoma (THCA), testicular germ cell tumors (TGCT), bladder urothelial carcinoma (BLCA), adenocarcinoma of the cervix (ACC), and kidney chromophobe (KICH) (Figure 1C). An analysis of MKNK2 expression in a variety of cancer cell lines, based on data from the Cancer Cell Line Encyclopedia (CCLE), exposed significant heterogeneity among the cell lines. Advanced tumors, particularly those of the cervical squamous cell carcinoma (CESC), thymoma (THYM), thyroid carcinoma (THCA), and kidney chromophobe (KICH), generally showed upregulation of MKNK2, whereas downregulation was observed in advanced tumors of the stomach squamous cell carcinoma (STES), uterine corpus endometrial carcinoma (UCEC), and bladder urothelial carcinoma (BLCA) (Figure 1D). These insights contribute to a nuanced understanding of MKNK2's potential as a biomarker and therapeutic target in cancer progression and heterogeneity.

Prognostic Analysis of MKNK2 in Pan-Cancer

The existing body of research hints at a connection between *MKNK2* levels and specific cancer types, including gastric and glioma cancers; however, the broader prognostic implications of this gene are not yet fully understood. Our study undertook a comprehensive assessment, evaluating the impact of *MKNK2* on overall survival (OS), disease-specific survival (DSS), disease-free interval (DFI), and Progression-Free Interval (PFI) across a diverse spectrum of 33 cancer types.

Through rigorous Cox regression analysis, we identified a significant correlation between *MKNK2* expression and OS in eight distinct cancer types, specifically TCGA-designated cancers, such as GBMLGG, LAML, LGG, ACC, LIHC, STES, KIPAN, and STAD. The Kaplan-Meier survival curves provided further validation, illustrating a particularly strong association between elevated *MKNK2* levels and diminished OS in cancers like CGA-GBM, CGA-LGG, TC-GA-LGG, TCGA-LIHC, TCGA-LAML, and TCGA-ACC. Conversely, in cancers such as TC-GA-STES, TCGA-KIPAN, and TCGA-STAD, lower *MKNK2* levels were associated with poorer outcomes (Figure 2).

Our examination of the relationship between DSS, DFI, PFI, and *MKNK2* expression levels revealed that high expression in cancers like TC-GA-GBMLGG, TCGA-LGG, TCGA-PRAD, and TCGA-ACC was linked to a poorer prognosis. Additionally, high *MKNK2* levels in TCGA-ACC and low levels in TCGA-STAD were correlated with adverse DFI outcomes. The PFI analysis mirrored these findings, with high expression in TC-GA-GBMLGG, TCGA-LGG, and TCGA-ACC, and low expression in TCGA-BLCA, indicating a poorer prognosis (Figures 3-5).

The synthesis of these analyses reinforces MKNK2's prognostic relevance in a subset of cancers. These findings not only point to *MKNK2*'s potential as an oncogene but also underscore its significance in the prognostic evaluation of cancer, offering valuable insights that may inform future therapeutic strategies and patient management.

Immune Cells Infiltration and Checkpoint Analyses

Emerging evidence has underscored the prognostic relevance of *MKNK2* across a spectrum of cancer types, with particular significance in glioblastoma multiforme low-grade gliomas (GBMLGG), liver hepatocellular car-

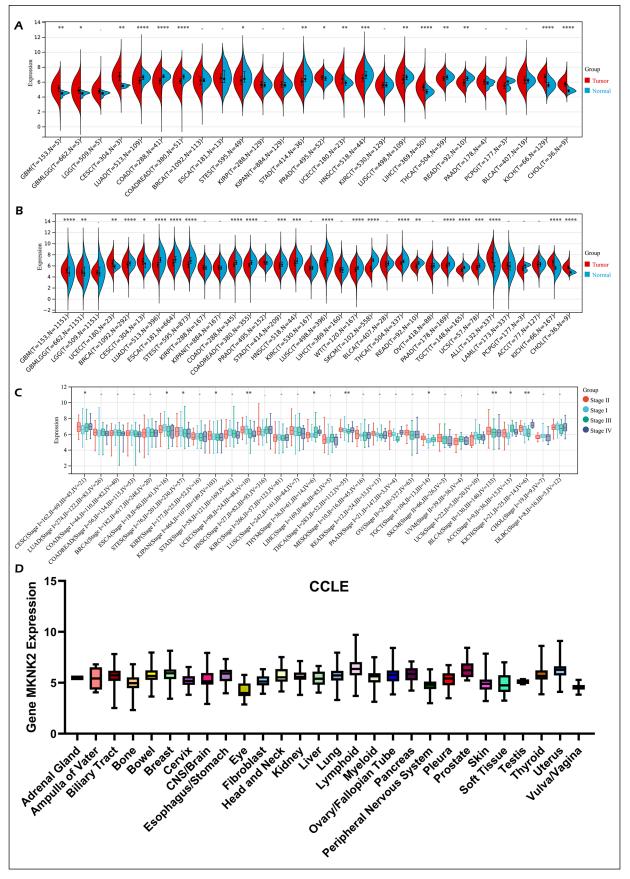


Figure 1. Differential expression of MKNK2. **A**, The expression level of MKNK2 in different cancer types from TCGA. **B**, The expression level of MKNK2 in different cancer types from TCGA and GTEx. **C**, Correlations between the MKNK2 expression and the major pathological stages of multiple cancer types were investigated based on the TCGA data. **D**, MKNK2 expression in various tumor cell lines based on the Cancer Cell Line Encyclopedia (CCLE) database. *p<0.05, **p<0.01, ***p<0.001. ns, not statistically significant.

cinoma (LIHC), and adenocarcinoma of the cervix (ACC). The immune microenvironment, modulated by the infiltration of immune cells, is increasingly recognized for its influence on cancer prognosis. Notably, research has established that *MKNK2* plays a regulatory role in

the anti-inflammatory phenotype of macrophages, a key component of the tumor's immune landscape¹⁶. Despite this, the extent to which *MKNK2* may influence the recruitment of immune cells into the tumor microenvironment remains an open question.

A					
CancerCode	pvalue		Hazard Ratio(95%CI)		
TCGA-GBMLGG(N=619)	2.4e-12	⊦• +	2.27 (1.81, 2.84)		
TCGA-LGG(N=474)	9.9e-9	} -	2.58 (1.87, 3.57)		
TCGA-ACC(N=77)	4.4e-3	 	2.22 (1.27, 3.86)		
TCGA-LIHC(N=341)	7.6e-3	I ● I	1.47 (1.11, 1.96)		
TCGA-LAML(N=144)	0.03]●	1.38 (1.03, 1.85)		
TCGA-THYM(N=117)	0.11	[2.16 (0.84, 5.56)		
TCGA-GBM(N=144)	0.18	I∳∙H	1.20 (0.92, 1.55)		
TCGA-PCPG(N=170)	0.22	-	2.48 (0.59, 10.38)		
TCGA-PRAD(N=492)	0.25		2.44 (0.52, 11.38)		
TCGA-KICH(N=64)	0.34	∤	1.71 (0.56, 5.21)		
TCGA-UVM(N=74)	0.36	[\	1.45 (0.65, 3.20)		
TCGA-DLBC(N=44)	0.45		1.50 (0.51, 4.39)		
TCGA-PAAD(N=172)	0.52	<mark>;● </mark>	1.13 (0.78, 1.64)		
TCGA-UCS(N=55)	0.60	 	1.13 (0.70, 1.83)		
TCGA-KIRP(N=276)	0.72	I	1.10 (0.66, 1.83)		
TCGA-MESO(N=84)	0.78	 -	1.05 (0.74, 1.49)		
TCGA-THCA(N=501)	0.81		1.14 (0.40, 3.25)		
TCGA-SKCM-P(N=97)	0.83		1.07 (0.60, 1.89)		
TCGA-COAD(N=278)	0.86	<mark>-</mark>	1.03 (0.72, 1.49)		
TCGA-COADREAD(N=368)	0.88	} -	1.03 (0.73, 1.43)		
TCGA-SKCM(N=444)	0.90	I-•∙I	1.01 (0.83, 1.24)		
TCGA-OV(N=407)	0.90	F∳H	1.01 (0.87, 1.17)		
TCGA-CESC(N=273)	0.91	[<mark>-</mark>	1.02 (0.75, 1.38)		
TCGA-STES(N=547)	1.6e-3	I- ⊕- I .	0.75 (0.63, 0.90)		
TCGA-STAD(N=372)	2.5e-3	}- 0 -1	0.71 (0.56, 0.88)		
TCGA-KIPAN(N=855)	0.04	I- ●- į	0.81 (0.66, 0.99)		
TCGA-ESCA(N=175)	0.08	[- ∮	0.78 (0.59, 1.03)		
TCGA-HNSC(N=509)	0.08	ŀ⊕-j	0.85 (0.71, 1.02)		
TCGA-UCEC(N=166)	0.21	I•-∳I	0.78 (0.53, 1.15)		
TCGA-KIRC(N=515)	0.48	<mark></mark>	0.91 (0.70, 1.18)		
TCGA-LUSC(N=468)	0.60	H <mark>♦</mark> H	0.96 (0.82, 1.12)		
TCGA-BRCA(N=1044)	0.62	I- <mark>•</mark> -1	0.95 (0.77, 1.17)		
TCGA-TGCT(N=128)	0.68	l	0.62 (0.06, 6.05)		
TCGA-LUAD(N=490)	0.76	I- • −1	0.97 (0.79, 1.19)		
TCGA-SARC(N=254)	0.78	<mark>-</mark>	0.96 (0.74, 1.26)		
TCGA-READ(N=90)	0.79	I	0.88 (0.36, 2.19)		
TCGA-SKCM-M(N=347)	0.93	ŀ•∳·l	0.99 (0.80, 1.23)		
TCGA-BLCA(N=398)	0.98	I-∳-I	1.00 (0.82, 1.21)		
TCOA-BLCA(N-396)			0.99 (0.34, 2.87)		

Figure 2. Association between the *MKNK2* expression and the OS of cancer patients. **A**, A forest plot of hazard ratios of *MKNK2* in different cancer types from TCGA. **B-I**, Kaplan-Meier survival curves of OS for patients stratified by the different expressions of *MKNK2* in GBMLGG, LGG, ACC, LIHC, LAML, STES, STAD, and KIPAN.

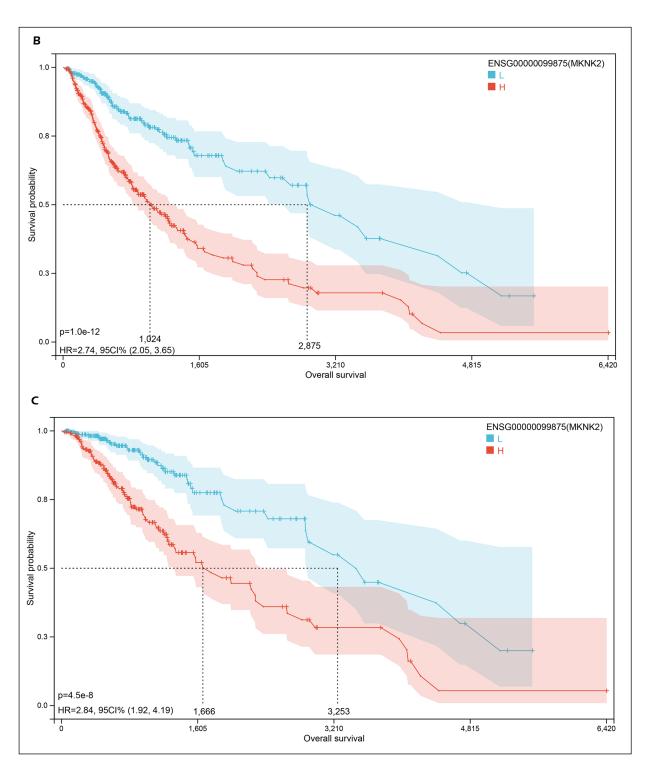


Figure 2 *(Continued).* Association between the *MKNK2* expression and the OS of cancer patients. **A**, A forest plot of hazard ratios of *MKNK2* in different cancer types from TCGA. **B-I**, Kaplan-Meier survival curves of OS for patients stratified by the different expressions of *MKNK2* in GBMLGG, LGG, ACC, LIHC, LAML, STES, STAD, and KIPAN.

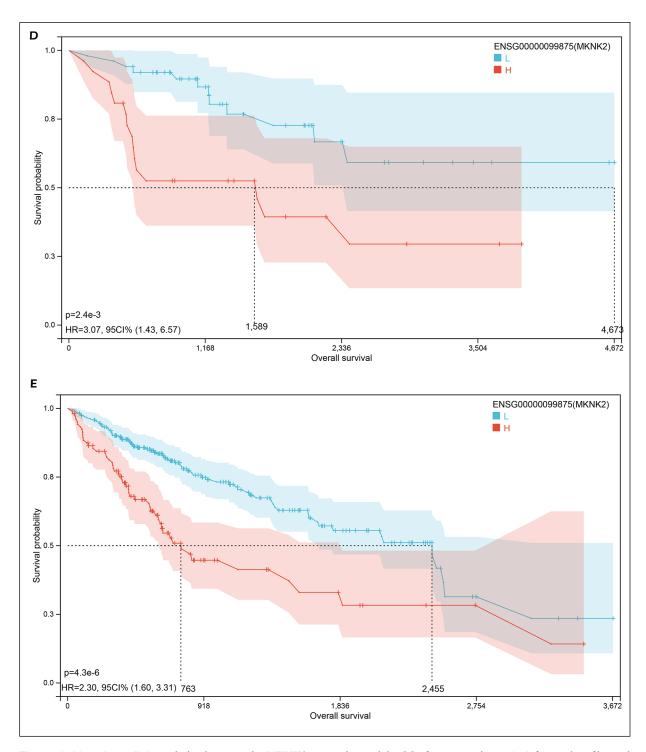


Figure 2 *(Continued).* Association between the *MKNK2* expression and the OS of cancer patients. **A**, A forest plot of hazard ratios of *MKNK2* in different cancer types from TCGA. **B-I**, Kaplan-Meier survival curves of OS for patients stratified by the different expressions of *MKNK2* in GBMLGG, LGG, ACC, LIHC, LAML, STES, STAD, and KIPAN.

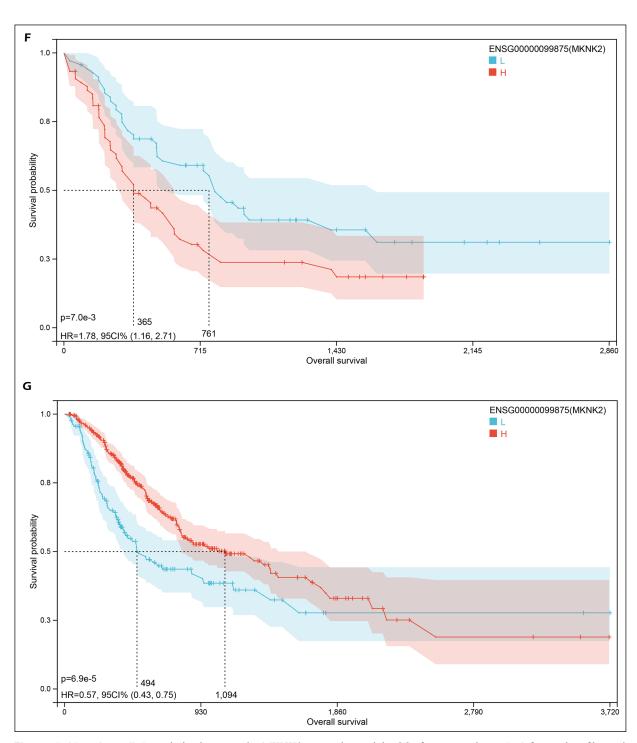


Figure 2 *(Continued).* Association between the *MKNK2* expression and the OS of cancer patients. **A**, A forest plot of hazard ratios of *MKNK2* in different cancer types from TCGA. **B-I**, Kaplan-Meier survival curves of OS for patients stratified by the different expressions of *MKNK2* in GBMLGG, LGG, ACC, LIHC, LAML, STES, STAD, and KIPAN.

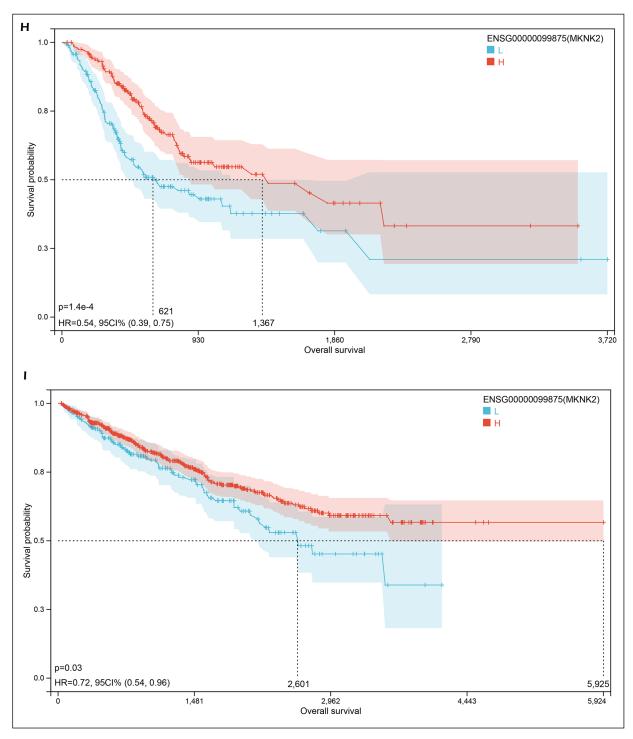


Figure 2 *(Continued).* Association between the *MKNK2* expression and the OS of cancer patients. **A**, A forest plot of hazard ratios of *MKNK2* in different cancer types from TCGA. **B-I**, Kaplan-Meier survival curves of OS for patients stratified by the different expressions of *MKNK2* in GBMLGG, LGG, ACC, LIHC, LAML, STES, STAD, and KIPAN.

In this study, we meticulously evaluated the relationship between *MKNK2* expression and immune cell infiltration across various cancers,

leveraging data from the TIMER database. The Spearman correlation coefficient was employed to quantify the association between gene expression and immune cell infiltration scores within each tumor context. Utilizing the R package psych (version 2.1.6) and its corr.test function, we discerned a significant correlation between *MKNK2* levels and immune infiltration in 31 distinct cancer types, including TCGA-designated cancers, such as BLCA, BRCA, CESC, and others. Con-

versely, no significant correlation was detected in cancers like MESO, DLBC, KICH, PAAD, and ACC (Figure 6A).

The advent of cancer immunotherapy has brought forth a new era in cancer treatment, with immune checkpoints emerging as pivotal regulators of the tumor-immune interface.

TCGA-LGG(N=466) 6.4e-9	Hazard Ratio(95%CI)	pvalue	CancerCode
TCGA-PRAD(N=490) 1.9e-3 TCGA-ACC(N=75) 3.7e-3 TCGA-THYM(N=117) 0.13 TCGA-PCPG(N=170) 0.13 TCGA-PCPG(N=170) 0.13 TCGA-UVM(N=74) 0.26 TCGA-GBM(N=131) 0.27 TCGA-BCHIC(N=333) 0.27 TCGA-LIHC(N=333) 0.27 TCGA-BCO(N=377) 0.31 TCGA-MESO(N=64) 0.36 TCGA-UCS(N=53) 0.52 TCGA-KICH(N=64) 0.36 TCGA-UCS(N=53) 0.52 TCGA-RESC(N=269) 0.60 TCGA-CACCS(N=269) 0.60 TCGA-READ(N=84) 0.67 TCGA-STES(N=524) 0.06 TCGA-STES(N=524) 0.06 TCGA-STES(N=524) 0.06 TCGA-STES(N=524) 0.06 TCGA-STES(N=524) 0.06 TCGA-STES(N=524) 0.06 TCGA-UCS(N=173) 0.08 TCGA-UCS(N=18) 0.17 TCGA-STAD(N=840) 0.16 TCGA-UCSC(N=18) 0.17 TCGA-STAD(N=840) 0.16 TCGA-STAD(N=840) 0.16 TCGA-TGCT(N=128) 0.26 TCGA-TGCT(N=128) 0.26 TCGA-STAD(N=385) 0.35 TCGA-STAC(N=485) 0.52 TCGA-SKCM-N=485) 0.52 TCGA-SKCM-N=485) 0.56 TCGA-SKCM-N=77) 0.69 TCGA-SKCM-N=97) 0.69 TCGA-SKCM-N=97) 0.69 TCGA-SCACN=1025) 0.87	2.20 (1.73, 2.79)	1.5e-10	TCGA-GBMLGG(N=598)
TCGA-ACC(N=75) 3.7e-3 TCGA-THYM(N=117) 0.13 TCGA-PCPG(N=170) 0.13 TCGA-PCPG(N=170) 0.13 TCGA-GBM(N=131) 0.27 TCGA-GBM(N=131) 0.27 TCGA-LIHC(N=333) 0.27 TCGA-ACO(N=77) 0.31 TCGA-BESO(N=64) 0.33 TCGA-MESO(N=64) 0.36 TCGA-UCS(N=53) 0.52 TCGA-CESC(N=269) 0.60 TCGA-CESC(N=269) 0.65 TCGA-CADREAD(N=84) 0.67 TCGA-CADREAD(N=84) 0.67 TCGA-CADREAD(N=84) 0.67 TCGA-CADREAD(N=84) 0.67 TCGA-CADREAD(N=84) 0.67 TCGA-STES(N=524) 0.06 TCGA-STES(N=524) 0.06 TCGA-STES(N=524) 0.06 TCGA-STES(N=524) 0.06 TCGA-UCC(N=164) 0.13 TCGA-UCC(N=164) 0.13 TCGA-UCCC(N=164) 0.13 TCGA-UCCC(N=164) 0.16 TCGA-STAD(N=840) 0.16 TCGA-TGCT(N=128) 0.26 TCGA-TGCT(N=128) 0.26 TCGA-TGCT(N=128) 0.26 TCGA-STAD(N=385) 0.35 TCGA-STAD(N=385) 0.35 TCGA-STAD(N=385) 0.56 TCGA-SKCM-M(N=341) 0.61	2.71 (1.93, 3.79)	6.4e-9	TCGA-LGG(N=466)
TCGA-THYM(N=117) TCGA-PCPG(N=170) TCGA-UVM(N=74) TCGA-QBM(N=131) TCGA-BAD(N=166) TCGA-UCS(N=377) TCGA-BEAD(N=64) TCGA-UCS(N=53) TCGA-UCS(N=53) TCGA-ESCA(N=172) TCGA-COAD(N=263) TCGA-COAD(N=263) TCGA-STES(N=524) TCGA-SECS(N=173) TCGA-UCS(N=173) TCGA-UCS(N=173) TCGA-UCS(N=173) TCGA-UCS(N=173) TCGA-UCS(N=173) TCGA-UCS(N=164) TCGA-UCS(N=173) TCGA-UCS(N=164) TCGA-UCS(N=164) TCGA-UCS(N=173) TCGA-UCS(N=164) TCGA-UCS(N=18) TCGA-UCS(N	193.75 (7.29, 5147.79)	1.9e-3	TCGA-PRAD(N=490)
TCGA-PCPG(N=170) 0.13	2.30 (1.30, 4.09)	3.7e-3	TCGA-ACC(N=75)
TCGA-UVM(N=74) TCGA-GBM(N=131) TCGA-LIHC(N=333) 0.27 TCGA-PAAD(N=166) TCGA-PAAD(N=166) TCGA-V(N=377) TCGA-WESO(N=64) TCGA-WESO(N=64) TCGA-UCS(N=53) TCGA-UCS(N=53) TCGA-CESC(N=269) TCGA-CESC(N=269) TCGA-READ(N=84) TCGA-COADREAD(N=347) TCGA-COADREAD(N=347) TCGA-STES(N=524) TCGA-STES(N=524) TCGA-SECA(N=173) TCGA-UCEC(N=164) TCGA-UCEC(N=164) TCGA-LUSC(N=48) TCGA-TGCT(N=128) TCGA-TGCT(N=128) TCGA-SARC(N=485) TCGA-SARC(N=485) TCGA-SKCM-M(N=341) TCGA-SKCM-M(N=341) TCGA-SKCM-P(N=97) TCGA-SUCCN-SCAN TCGA-SCA(N=173) TCGA-SCA(N=173) TCGA-SKCM-P(N=97) TCGA-SKCM-P(N=97) TCGA-SKCM-P(N=97) TCGA-SCA(N=1025) TCGA-BCA(N=1025) TCGA-BC	2.60 (0.74, 9.06)	0.13	TCGA-THYM(N=117)
TCGA-GBM(N=131) TCGA-LIHC(N=333) 0.27 TCGA-PAAD(N=166) TCGA-PAAD(N=166) TCGA-OV(N=377) TCGA-MESO(N=64) TCGA-KICH(N=64) TCGA-CSC(N=53) TCGA-CSC(N=269) TCGA-CSC(N=269) TCGA-CACAD(N=347) TCGA-CAD(N=347) TCGA-CAD(N=263) TCGA-CAD(N=263) TCGA-CAD(N=263) TCGA-CAD(N=263) TCGA-STES(N=524) TCGA-STES(N=524) TCGA-UCEC(N=164) TCGA-LUCC(N=164) TCGA-LUCC(N=188) TCGA-TGCA(N=385) TCGA-TGCT(N=128) TCGA-STCC(N=288) TCGA-SARC(N=288) TCGA-SARCA(N=288) TCGA-SARCA(N	4.21 (0.66, 26.92)	0.13	TCGA-PCPG(N=170)
TCGA-LIHC(N=333) 0.27	1.64 (0.69, 3.87)	0.26	TCGA-UVM(N=74)
TCGA-PAAD(N=166) 0.27 TCGA-PAAD(N=166) 0.31 TCGA-MESO(N=64) 0.33 TCGA-KICH(N=64) 0.36 TCGA-CESC(N=53) 0.52 TCGA-CESC(N=269) 0.60 TCGA-KIRP(N=272) 0.65 TCGA-CACACACACACACACACACACACACACACACACACA	1.16 (0.89, 1.52)	0.27	TCGA-GBM(N=131)
TCGA-OV(N=377) TCGA-MESO(N=64) TCGA-MESO(N=64) TCGA-KICH(N=64) TCGA-CSC(N=53) TCGA-CSC(N=269) TCGA-CSC(N=269) TCGA-READ(N=84) TCGA-COADREAD(N=347) TCGA-COADREAD(N=347) TCGA-STES(N=524) TCGA-STES(N=524) TCGA-STES(N=173) TCGA-UCEC(N=164) TCGA-LUSC(N=418) TCGA-STAD(N=351) TCGA-TGCT(N=128) TCGA-TGCT(N=128) TCGA-SARC(N=485) TCGA-SARC(N=485) TCGA-SKCM-M(N=341) TCGA-SKCM-M(N=341) TCGA-SKCM-M(N=37) TCGA-SKCM-M(N=37) TCGA-SKCM-M(N=37) TCGA-SKCM-M(N=37) TCGA-SKCM-M(N=37) TCGA-SKCM-M(N=37) TCGA-SKCM-M(N=37) TCGA-SKCM-M(N=37) TCGA-SCANCN=105) TCGA-SCANCN=105) TCGA-SCANCN=105 TCGA-SCANCN=105	1.22 (0.85, 1.76)	0.27	TCGA-LIHC(N=333)
TCGA-MESO(N=64) TCGA-MESO(N=64) TCGA-KICH(N=64) TCGA-UCS(N=53) TCGA-CSC(N=269) TCGA-CSC(N=269) TCGA-CSC(N=269) TCGA-READ(N=84) TCGA-COADREAD(N=347) TCGA-COADREAD(N=347) TCGA-COADREAD(N=263) TCGA-STES(N=524) TCGA-STES(N=524) TCGA-STES(N=524) TCGA-UCEC(N=164) TCGA-LUSC(N=418) TCGA-KIPAN(N=840) TCGA-STAD(N=351) TCGA-TGCT(N=128) TCGA-TGCT(N=128) TCGA-TGCT(N=128) TCGA-KIRC(N=504) TCGA-KIRC(N=504) TCGA-SARC(N=485) TCGA-SARC(N=485) TCGA-SARC(N=485) TCGA-SKCM-M(N=341) TCGA-SKCM-M(N=341) TCGA-SKCM-P(N=97) TCGA-BCA-CN=1025) TCGA-BCA-CN=1025) TCGA-BCA-CN=1025) TCGA-BCA-CN=1025) TCGA-BCA-CN=1025) TCGA-BCA-CN=1025) TCGA-BCA-CN=1025) TCGA-BCA-CN=1025) TCGA-BCA-CN=1025)	1.27 (0.83, 1.94)	0.27	TCGA-PAAD(N=166)
TCGA-KICH(N=64) TCGA-UCS(N=53) TCGA-CESC(N=269) TCGA-CESC(N=269) TCGA-READ(N=84) TCGA-COADREAD(N=347) TCGA-COADREAD(N=263) TCGA-STES(N=524) TCGA-STES(N=524) TCGA-UCE(N=164) TCGA-UCEC(N=164) TCGA-LUSC(N=418) TCGA-STAD(N=351) TCGA-TGCT(N=128) TCGA-TGCT(N=128) TCGA-KIRC(N=504) TCGA-KIRC(N=504) TCGA-KIRC(N=504) TCGA-SARC(N=485) TCGA-SARC(N=485) TCGA-SKCM-M(N=341) TCGA-SKCM-M(N=341) TCGA-SKCM-P(N=97) TCGA-LUAD(N=457) TCGA-BCA(N=1025) TCGA	1.09 (0.92, 1.30)	0.31	TCGA-OV(N=377)
TCGA-UCS(N=53) TCGA-CESC(N=269) TCGA-KIRP(N=272) TCGA-READ(N=84) TCGA-COADREAD(N=347) TCGA-COADREAD(N=347) TCGA-STES(N=524) TCGA-STES(N=524) TCGA-UCSC(N=164) TCGA-UCSC(N=164) TCGA-KIPAN(N=840) TCGA-LUSC(N=418) TCGA-STAD(N=351) TCGA-TGCT(N=128) TCGA-TGCT(N=128) TCGA-KIRC(N=504) TCGA-KIRC(N=504) TCGA-KIRC(N=485) TCGA-SARC(N=485) TCGA-SKCM-M(N=341) TCGA-SKCM-M(N=341) TCGA-SKCM-P(N=97) TCGA-LUAD(N=457) TCGA-BLCA(N=1025) TCGA-BCA(N=1025)	1.26 (0.80, 1.98)	0.33	TCGA-MESO(N=64)
TCGA-CESC(N=269) 0.60 TCGA-KIRP(N=272) 0.65 TCGA-READ(N=84) 0.67 TCGA-COADREAD(N=347) 0.76 TCGA-COADREAD(N=263) 0.97 TCGA-STES(N=524) 0.06 TCGA-STES(N=524) 0.08 TCGA-UCEC(N=164) 0.13 TCGA-UCEC(N=164) 0.16 TCGA-LUSC(N=418) 0.17 TCGA-STAD(N=351) 0.18 TCGA-TGCT(N=128) 0.26 TCGA-BLCA(N=385) 0.35 TCGA-KIRC(N=504) 0.47 TCGA-HNSC(N=485) 0.52 TCGA-SARC(N=248) 0.56 TCGA-SKCM-M(N=341) 0.61 TCGA-SKCM-M(N=341) 0.61 TCGA-SKCM-M(N=341) 0.61 TCGA-SKCM-P(N=97) 0.69 TCGA-LUAD(N=457) 0.83 TCGA-BRCA(N=1025) 0.87	1.79 (0.50, 6.40)	0.36	TCGA-KICH(N=64)
TCGA-KIRP(N=272) 0.65	1.17 (0.72, 1.90)	0.52	TCGA-UCS(N=53)
TCGA-READ(N=84) TCGA-COADREAD(N=347) TCGA-COADREAD(N=347) TCGA-COAD(N=263) TCGA-STES(N=524) TCGA-STES(N=524) TCGA-ESCA(N=173) TCGA-UCEC(N=164) TCGA-LUCEC(N=164) TCGA-LUSC(N=418) TCGA-LUSC(N=418) TCGA-TGCT(N=128) TCGA-TGCT(N=128) TCGA-BLCA(N=385) TCGA-KIRC(N=504) TCGA-KIRC(N=504) TCGA-SARC(N=485) TCGA-SARC(N=248) TCGA-SKCM-M(N=341) TCGA-SKCM-M(N=341) TCGA-SKCM-M(N=341) TCGA-SKCM-P(N=97) TCGA-LUAD(N=457) TCGA-BRCA(N=1025) 0.87	1.10 (0.78, 1.54)	0.60	TCGA-CESC(N=269)
TCGA-COADREAD(N=347) TCGA-COAD(N=263) TCGA-STES(N=524) TCGA-STES(N=524) TCGA-ESCA(N=173) TCGA-UCEC(N=164) TCGA-KIPAN(N=840) TCGA-LUSC(N=418) TCGA-STAD(N=351) TCGA-TGCT(N=128) TCGA-BLCA(N=385) TCGA-BLCA(N=385) TCGA-KIRC(N=504) TCGA-KIRC(N=485) TCGA-SARC(N=248) TCGA-SARC(N=248) TCGA-SKCM-M(N=341) TCGA-SKCM-M(N=341) TCGA-SKCM-M(N=341) TCGA-SKCM-P(N=97) TCGA-LUAD(N=457) TCGA-BCA(N=1025)	1.16 (0.62, 2.15)	0.65	TCGA-KIRP(N=272)
TCGA-COAD(N=263) 0.97	1.39 (0.30, 6.45)	0.67	TCGA-READ(N=84)
TCGA-STES(N=524) TCGA-ESCA(N=173) TCGA-ESCA(N=173) TCGA-UCEC(N=164) TCGA-KIPAN(N=840) TCGA-LUSC(N=418) TCGA-TGCT(N=128) TCGA-TGCT(N=128) TCGA-BLCA(N=385) TCGA-KIRC(N=504) TCGA-KIRC(N=504) TCGA-SARC(N=248) TCGA-SARC(N=248) TCGA-SKCM-M(N=341) TCGA-SKCM-M(N=341) TCGA-SKCM-P(N=97) TCGA-LUAD(N=457) TCGA-BCCA(N=1025)	1.08 (0.67, 1.74)	0.76	TCGA-COADREAD(N=347)
TCGA-ESCA(N=173) 0.08 TCGA-UCEC(N=164) 0.13 TCGA-KIPAN(N=840) 0.16 TCGA-LUSC(N=418) 0.17 TCGA-STAD(N=351) 0.18 TCGA-GA-GCT(N=128) 0.26 TCGA-BLCA(N=385) 0.35 TCGA-BLCA(N=385) 0.47 TCGA-KIRC(N=504) 0.47 TCGA-HNSC(N=485) 0.52 TCGA-SARC(N=248) 0.56 TCGA-SKCM-M(N=341) 0.61 TCGA-SKCM-M(N=341) 0.61 TCGA-SKCM-P(N=97) 0.69 TCGA-LUAD(N=457) 0.83 TCGA-BRCA(N=1025) 0.87	1.01 (0.62, 1.63)	0.97	TCGA-COAD(N=263)
TCGA-UCEC(N=164) TCGA-UCEC(N=164) TCGA-KIPAN(N=840) TCGA-LUSC(N=418) TCGA-STAD(N=351) TCGA-TGCT(N=128) TCGA-BLCA(N=385) TCGA-BLCA(N=385) TCGA-KIRC(N=504) TCGA-HNSC(N=485) TCGA-HNSC(N=485) TCGA-SARC(N=248) TCGA-SKCM-M(N=341) TCGA-SKCM-M(N=341) TCGA-SKCM-P(N=97) TCGA-LUAD(N=457) TCGA-BCA(N=1025) 0.13 1-1-1 1-1 1-1 1-1-1 1-	0.81 (0.65, 1.01)	0.06	TCGA-STES(N=524)
TCGA-KIPAN(N=840) TCGA-LUSC(N=418) TCGA-STAD(N=351) TCGA-TGCT(N=128) TCGA-BLCA(N=385) TCGA-BLCA(N=385) TCGA-KIRC(N=504) TCGA-HNSC(N=485) TCGA-SARC(N=248) TCGA-SARC(N=248) TCGA-SKCM-M(N=341) TCGA-SKCM-M(N=341) TCGA-SKCM-P(N=97) TCGA-SKCM-P(N=97) TCGA-LUAD(N=457) 0.83	0.74 (0.52, 1.04)	0.08	TCGA-ESCA(N=173)
TCGA-LUSC(N=418) TCGA-STAD(N=351) TCGA-TGCT(N=128) TCGA-BLCA(N=385) TCGA-KIRC(N=504) TCGA-HNSC(N=485) TCGA-SARC(N=248) TCGA-SARC(N=248) TCGA-SKCM-M(N=341) TCGA-SKCM-M(N=438) TCGA-SKCM-P(N=97) TCGA-LUAD(N=457) TCGA-BRCA(N=1025) 0.17 0.18 0.17 0.18 0.19 10 10 11 11 11 11 11 11 11	0.70 (0.45, 1.11)	0.13	TCGA-UCEC(N=164)
TCGA-STAD(N=351) TCGA-TGCT(N=128) TCGA-BLCA(N=385) TCGA-KIRC(N=504) TCGA-HNSC(N=485) TCGA-SARC(N=248) TCGA-SKCM-M(N=341) TCGA-SKCM-M(N=341) TCGA-SKCM-P(N=97) TCGA-LUAD(N=457) TCGA-BRCA(N=1025) 0.18 10 11 11 11 11 11 11 11 11	0.83 (0.65, 1.07)	0.16	TCGA-KIPAN(N=840)
TCGA-TGCT(N=128) TCGA-BLCA(N=385) TCGA-KIRC(N=504) TCGA-HNSC(N=485) TCGA-SARC(N=248) TCGA-SARC(N=248) TCGA-SKCM-M(N=341) TCGA-SKCM(N=438) TCGA-SKCM-P(N=97) TCGA-LUAD(N=457) TCGA-BRCA(N=1025) 0.26 0.35 0.47 0.61 0.61 0.61 0.61 0.61 0.61 0.61 0.61 0.63 0.64 0.64 0.69 0.69 0.69 0.69 0.69 0.69	0.85 (0.67, 1.07)	0.17	TCGA-LUSC(N=418)
TCGA-BLCA(N=385) TCGA-KIRC(N=504) TCGA-KIRC(N=485) TCGA-SARC(N=248) TCGA-SARC(N=248) TCGA-SKCM-M(N=341) TCGA-SKCM(N=438) TCGA-SKCM-P(N=97) TCGA-LUAD(N=457) TCGA-BRCA(N=1025) 0.35 0.35 0.35 0.35 0.35 0.35 0.35 0.35 0.35 0.37	0.82 (0.61, 1.09)	0.18	TCGA-STAD(N=351)
TCGA-KIRC(N=504) 0.47 TCGA-HNSC(N=485) 0.52 TCGA-SARC(N=248) 0.56 TCGA-SKCM-M(N=341) 0.61 TCGA-SKCM(N=438) 0.64 TCGA-SKCM-P(N=97) 0.69 TCGA-LUAD(N=457) 0.83 TCGA-BRCA(N=1025) 0.87	0.12 (2.7e-3, 5.16)	0.26	TCGA-TGCT(N=128)
TCGA-HNSC(N=485) 0.52 TCGA-SARC(N=248) 0.56 TCGA-SKCM-M(N=341) 0.61 TCGA-SKCM(N=438) 0.64 TCGA-SKCM-P(N=97) 0.69 TCGA-LUAD(N=457) 0.83 TCGA-BRCA(N=1025) 0.87	0.89 (0.71, 1.13)	0.35	TCGA-BLCA(N=385)
TCGA-SARC(N=248) 0.56 TCGA-SKCM-M(N=341) 0.61 TCGA-SKCM(N=438) 0.64 TCGA-SKCM-P(N=97) 0.69 TCGA-LUAD(N=457) 0.83 TCGA-BRCA(N=1025) 0.87	0.89 (0.64, 1.23)	0.47	TCGA-KIRC(N=504)
TCGA-SKCM-M(N=341) 0.61 TCGA-SKCM(N=438) 0.64 TCGA-SKCM-P(N=97) 0.69	0.93 (0.74, 1.17)	0.52	TCGA-HNSC(N=485)
TCGA-SKCM(N=438) 0.64 TCGA-SKCM-P(N=97) 0.69	0.92 (0.68, 1.23)	0.56	TCGA-SARC(N=248)
TCGA-SKCM-P(N=97) 0.69 1-0-1 TCGA-LUAD(N=457) 0.83 10-1 TCGA-BRCA(N=1025) 0.87	0.94 (0.75, 1.18)	0.61	TCGA-SKCM-M(N=341)
TCGA-LUAD(N=457) 0.83 IIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIII	0.95 (0.77, 1.18)	0.64	TCGA-SKCM(N=438)
TCGA-BRCA(N=1025) 0.87	0.87 (0.44, 1.73)	0.69	TCGA-SKCM-P(N=97)
100H BREA(IV 1023)	0.97 (0.75, 1.25)	0.83	TCGA-LUAD(N=457)
	0.98 (0.73, 1.30)	0.87	TCGA-BRCA(N=1025)
1CGN-DEBC(17 +4) 0.50	0.91 (0.23, 3.69)	0.90	TCGA-DLBC(N=44)
TCGA-THCA(N=495) 0.96	0.96 (0.20, 4.63)	0.96	TCGA-THCA(N=495)
TCGA-CHOL(N=32) 0.97	0.98 (0.32, 2.97)	0.97	TCGA-CHOL(N=32)

Figure 3. Association between the *MKNK2* expression and DSS in cancer patients. **A**, A forest plot of hazard ratios of *MKNK2* in different cancer types from TCGA. **B-E**, Kaplan-Meier survival curves of DSS for patients stratified by the different expressions of *MKNK2* in GBMLGG, LGG, ACC, and PRAD.

Figure continued

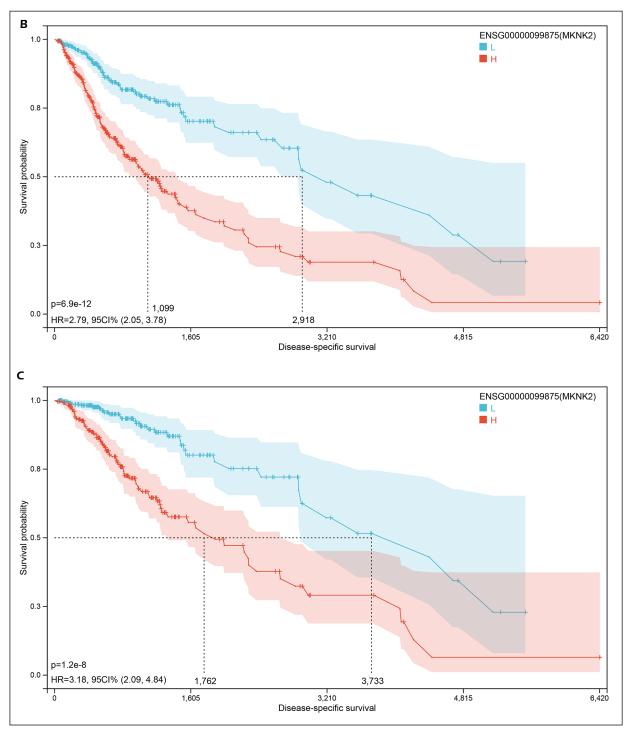


Figure 3 *(Continued).* Association between the *MKNK2* expression and DSS in cancer patients. **A**, A forest plot of hazard ratios of *MKNK2* in different cancer types from TCGA. **B-E**, Kaplan-Meier survival curves of DSS for patients stratified by the different expressions of *MKNK2* in GBMLGG, LGG, ACC, and PRAD. *Figure continued*

Our analysis revealed a broad association between *MKNK2* expression and immune checkpoint status across all pan-cancers (Figure 6B). These findings are corroborated by the results

obtained from the CIBERSORT (Figure 6C) and EPIC (Figure 6D) algorithms, which estimate the immune score based on gene expression profiles.

Moreover, recent studies have suggested that the pharmacological inhibition of *MKNK2* can impede *eIF4E*-regulated translation, thereby suppressing the growth of stem cells and decelerat-

ing tumor growth *in vivo*¹⁷⁻¹⁹. These observations collectively point towards a potential role for *MKNK2* in modulating the density of immune cells within tumors, offering a promising avenue

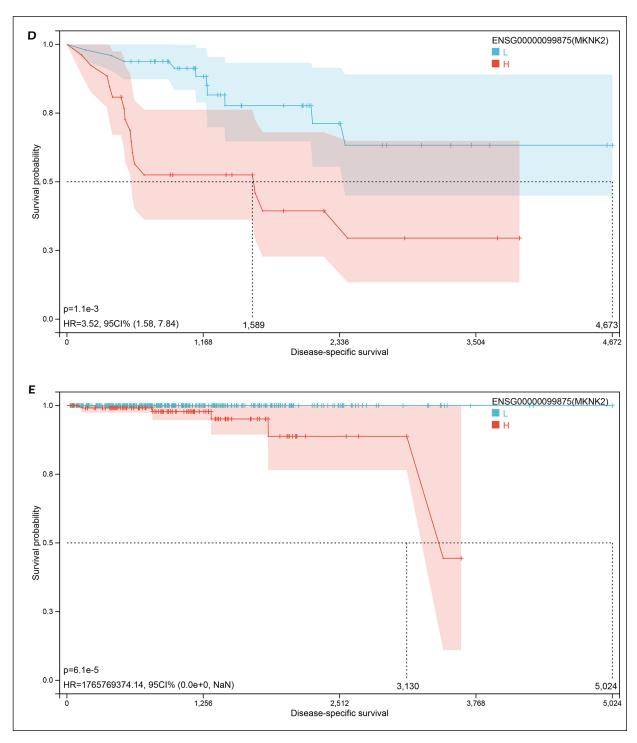


Figure 3 *(Continued).* Association between the *MKNK2* expression and DSS in cancer patients. **A**, A forest plot of hazard ratios of *MKNK2* in different cancer types from TCGA. **B-E**, Kaplan-Meier survival curves of DSS for patients stratified by the different expressions of *MKNK2* in GBMLGG, LGG, ACC, and PRAD.

for the development of therapies targeting refractory tumors (Figure 6).

Tumor Mutation Burden and Microsatellite Instability Analyses

Microsatellite instability (MSI), associated with defects in DNA mismatch repair, stands as a pivotal clinical marker in oncology. Both tumor mutational burden (TMB) and MSI have emerged as crucial factors that influence the genesis and progression of tumors. Their significance extends to the realm of immunotherapy, where they are recognized as burgeoning biomarkers that predict patient responses to immune checkpoint inhibitors.

Our investigation reveals a significant correlation between *MKNK2* expression levels and TMB across a wide array of tumor types. This association is evident in malignancies such as glioblastoma (GBM), glioblastoma multiforme low-grade gliomas (GBMLGG), cervical squamous cell carcinoma (CESC), colorectal adenocarcinoma with readthrough (COADREAD), colorectal adenocarcinoma (COAD), sarcoma (SARC), uterine corpus endometrial carcinoma (UCEC), stomach adenocarcinoma (STAD), head and neck squamous cell carcinoma (HNSC), liver hepatocellular carcinoma (LIHC), thymoma (THYM), kidney chromophobe (KICH), breast invasive carcinoma

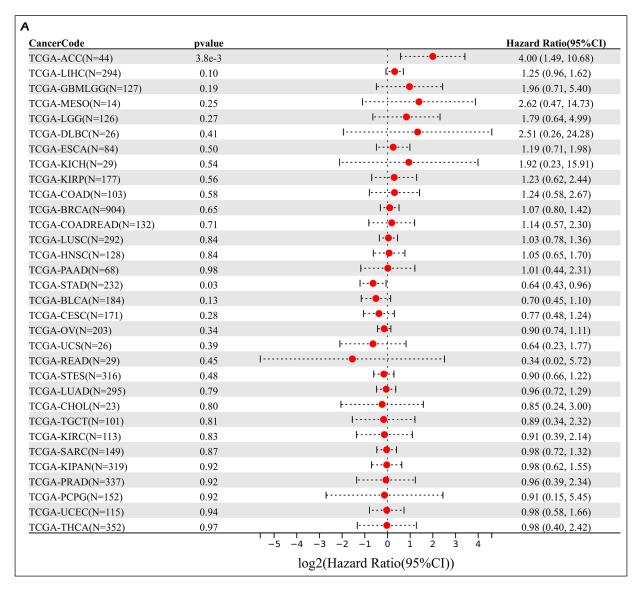


Figure 4. Association between the *MKNK2* expression and DFI in cancer patients. **A**, A forest plot of hazard ratios of *MKNK2* in different cancer types from TCGA. **B-C**, Kaplan-Meier survival curves of DFI for patients stratified by the different expressions of *MKNK2* in ACC and STAD.

Figure continued

(BRCA), brain lower grade glioma (LGG), and kidney renal papillary cell carcinoma (KIRP), as well as in kidney renal clear cell carcinoma (KIPAN) (Figure 7A).

Furthermore, an assessment of the relationship between *MKNK2* expression and MSI across 11 cancer types uncovered a markedly positive correlation in tumors such as lung adenocarcinoma

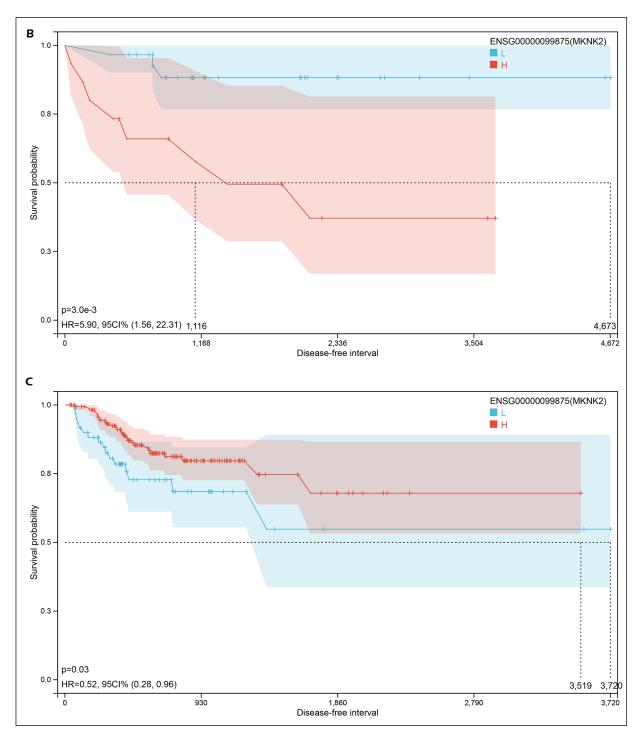


Figure 4 *(Continued).* Association between the *MKNK2* expression and DFI in cancer patients. **A**, A forest plot of hazard ratios of *MKNK2* in different cancer types from TCGA. **B-C**, Kaplan-Meier survival curves of DFI for patients stratified by the different expressions of *MKNK2* in ACC and STAD.

G G I	_		TT 1D 1 10 20 1 20 20 1
CancerCode	pvalue	0	Hazard Ratio(95%CI)
TCGA-GBMLGG(N=616)	9.2e-11	1	1.96 (1.60, 2.40)
TCGA-LGG(N=472)	2.9e-8		2.09 (1.61, 2.72)
TCGA-ACC(N=76)	2.3e-4		2.51 (1.53, 4.11)
TCGA-LIHC(N=340)	0.14		1.19 (0.95, 1.51)
TCGA-UVM(N=73)	0.18		1.63 (0.79, 3.38)
TCGA-DLBC(N=43)	0.18		1.98 (0.73, 5.36)
TCGA-KICH(N=64)	0.19	 	1.95 (0.70, 5.38)
TCGA-MESO(N=82)	0.24		1.25 (0.86, 1.83)
TCGA-BRCA(N=1043)	0.39	F÷	1.10 (0.89, 1.37)
TCGA-KIRP(N=273)	0.47	F -	1.18 (0.75, 1.87)
TCGA-PAAD(N=171)	0.60	I;•I	1.10 (0.76, 1.61)
TCGA-UCS(N=55)	0.67	F	1.10 (0.70, 1.75)
TCGA-THYM(N=117)	0.83		1.08 (0.53, 2.19)
TCGA-LUSC(N=467)	0.93	I <mark>∳</mark> I	1.01 (0.84, 1.21)
TCGA-STAD(N=375)	0.03	-	0.77 (0.61, 0.97)
TCGA-BLCA(N=397)	0.04	} - ∤	0.81 (0.66, 0.99)
TCGA-COADREAD(N=363)	0.07	1	0.79 (0.60, 1.02)
TCGA-READ(N=88)	0.08	 -	0.46 (0.19, 1.10)
TCGA-KIPAN(N=845)	0.12	मन••∳	0.85 (0.69, 1.05)
TCGA-CHOL(N=33)	0.16	ł	0.50 (0.19, 1.31)
TCGA-UCEC(N=166)	0.20	I - <mark>-</mark> I	0.81 (0.58, 1.12)
TCGA-STES(N=548)	0.22	<mark></mark> -¦	0.90 (0.75, 1.07)
TCGA-COAD(N=275)	0.23	F <mark>-</mark>	0.84 (0.63, 1.11)
TCGA-SARC(N=250)	0.26	F -	0.88 (0.70, 1.10)
TCGA-KIRC(N=508)	0.30	I•-∳-I	0.86 (0.66, 1.14)
TCGA-SKCM-P(N=96)	0.40		0.80 (0.48, 1.34)
TCGA-LUAD(N=486)	0.46	I <mark>•</mark> I	0.93 (0.76, 1.13)
TCGA-PRAD(N=492)	0.51	<u> </u>	0.85 (0.53, 1.37)
TCGA-TGCT(N=126)	0.55	ļ	0.77 (0.33, 1.81)
TCGA-ESCA(N=173)	0.62	} - I	0.93 (0.71, 1.22)
TCGA-SKCM(N=434)	0.64	<mark>-</mark>	0.96 (0.81, 1.14)
TCGA-CESC(N=273)	0.73	-	0.95 (0.71, 1.27)
TCGA-GBM(N=143)	0.75	-	0.96 (0.74, 1.25)
TCGA-PCPG(N=168)	0.76	ļ	0.88 (0.38, 2.02)
TCGA-SKCM-M(N=338)	0.70		0.98 (0.81, 1.18)
TCGA-THCA(N=499)	0.83		0.94 (0.51, 1.72)
TCGA-OV(N=406)	0.83		0.94 (0.31, 1.72)
TCGA-HNSC(N=508)	0.90	11	1.00 (0.83, 1.20)
	0.99	· 🕌 '	1.00 (0.85, 1.20)

Figure 5. Association between the *MKNK2* expression and PFI in cancer patients. **A**, A forest plot of hazard ratios of *MKNK2* in different cancer types from TCGA. **B-F**, Kaplan-Meier survival curves of PFI for patients stratified by the different expressions of *MKNK2* in GBMLGG, LGG, ACC, STAD, and BLCA.

(LUAD), kidney renal papillary cell carcinoma (KIPAN), stomach adenocarcinoma (STAD), uterine corpus endometrial carcinoma (UCEC), lung squamous cell carcinoma (LUSC), liver he-

patocellular carcinoma (LIHC), testicular germ cell tumors (TGCT), and kidney chromophobe (KICH). In contrast, a negative correlation was observed in glioblastoma multiforme low-grade

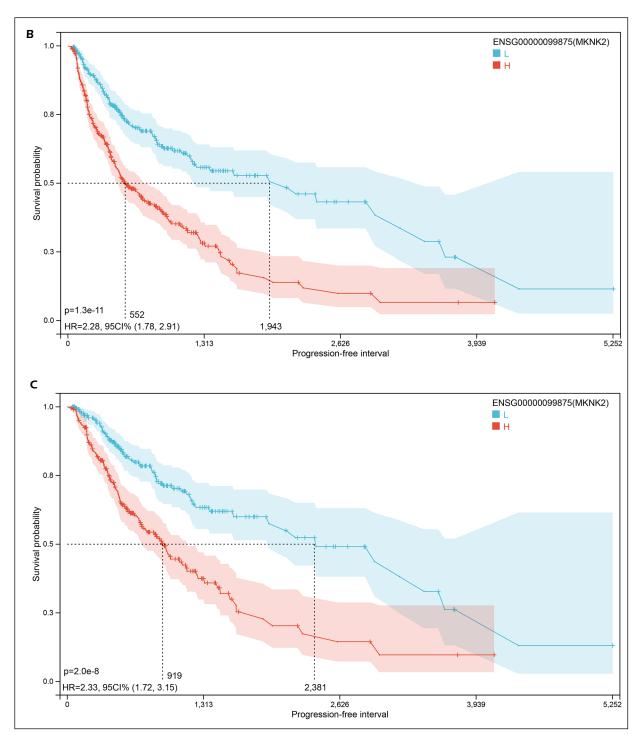


Figure 5 *(Continued).* Association between the *MKNK2* expression and PFI in cancer patients. **A**, A forest plot of hazard ratios of *MKNK2* in different cancer types from TCGA. **B-F**, Kaplan-Meier survival curves of PFI for patients stratified by the different expressions of *MKNK2* in GBMLGG, LGG, ACC, STAD, and BLCA.

gliomas (GBMLGG), breast invasive carcinoma (BRCA), and diffuse large B-cell lymphoma (DLBC) (Figure 7B).

These findings not only highlight the intricate relationship between *MKNK2*, TMB, and MSI but also emphasize the potential of *MKNK2* as a bio-

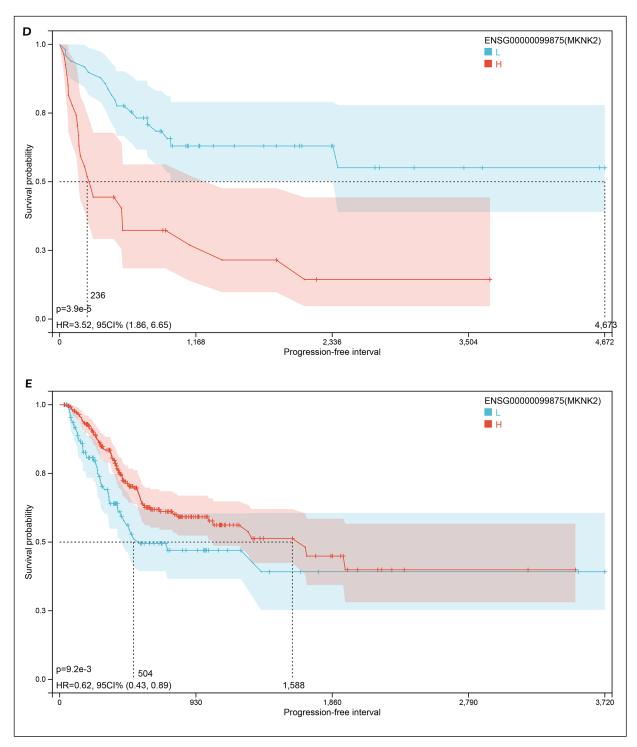


Figure 5 *[Continued].* Association between the *MKNK2* expression and PFI in cancer patients. **A**, A forest plot of hazard ratios of *MKNK2* in different cancer types from TCGA. **B-F**, Kaplan-Meier survival curves of PFI for patients stratified by the different expressions of *MKNK2* in GBMLGG, LGG, ACC, STAD, and BLCA.

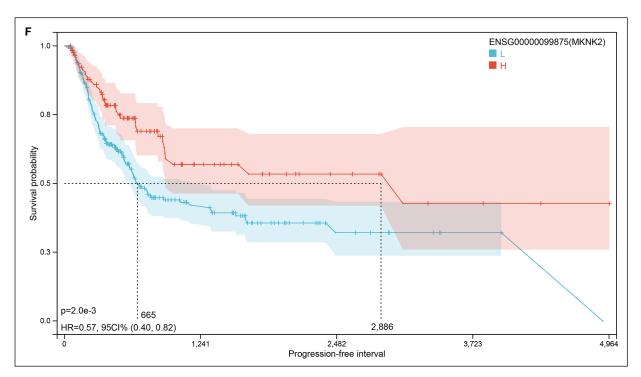


Figure 5 *(Continued).* Association between the *MKNK2* expression and PFI in cancer patients. **A**, A forest plot of hazard ratios of *MKNK2* in different cancer types from TCGA. **B-F**, Kaplan-Meier survival curves of PFI for patients stratified by the different expressions of *MKNK2* in GBMLGG, LGG, ACC, STAD, and BLCA.

marker that could inform therapeutic strategies, particularly in the context of immunotherapy.

The DNA Methylation-Based and the RNA-Based Stemness Scores

It is widely recognized that tumor stem cells play a pivotal role in the initiation of tumors and the establishment of the tumor microenvironment (TME). In our analysis, we calculated stemness scores and determined the Spearman correlation for each type of tumor, uncovering a significant association in 16 different malignancies. Notably, we identified a robust positive correlation in nine of these tumors, including glioblastoma multiforme low-grade gliomas (GBMLGG), lower-grade glioma (LGG), cervical squamous cell carcinoma (CESC), kidney renal papillary cell carcinoma (KIRP), kidney pancreatic cancer (KIPAN), thymoma (THYM), penile cancer (PCPG), uveal melanoma (UVM), and cholangiocarcinoma (CHOL). Conversely, a significant negative correlation was observed in seven tumors, comprising lung adenocarcinoma (LUAD), colorectal adenocarcinoma (COAD), colorectal adenocarcinoma with readthrough (COADREAD), breast invasive carcinoma (BRCA), thymic carcinoma (THCA), rectal adenocarcinoma (READ), and testicular germ cell tumors (TGCT) (Figure 7C).

The *MNK* signaling pathway has been identified as essential for the preservation of stem cell populations across various cancers (Figure 7C-D). Specifically, in blast crisis chronic myeloid leukemia, the *MNK-eIF4E* signaling cascade is known to sustain the leukemic precursors¹⁹. Additionally, *MKNK1* has been reported to positively regulate the levels of *Sema3C* and growth-supporting factors such as $TGF-\beta$, which are implicated in the maintenance and progression of glioma stem cells^{20,21}. The $TGF-\beta$ signaling pathway is also recognized for its role in promoting the growth, invasion, and immune evasion of mesenchymal stem cells within tumors.

Given these insights, further research is both necessary and promising, potentially leading to significant clinical applications in tumor therapy. To gain a comprehensive understanding of the molecular processes regulated by *MKNK2* in different types of cancer, we conducted Gene Set Enrichment Analysis (GSEA) and Kyoto Encyclopedia of Genes and Genomes (KEGG) analyses. The collective results from these analyses suggest that elevated *MKNK2* expression is predominantly associated with immune response and stemness in cancer, offering valuable clues for the

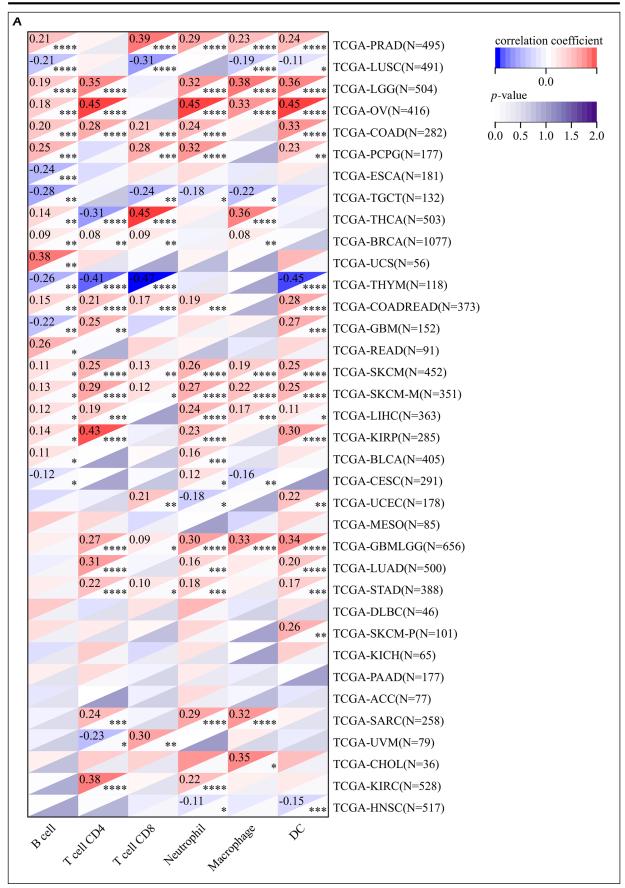


Figure 6. Different algorithms were used to explore the potential correlation between the expression level of the MKNK2 gene and the level of immune cell infiltration in all types of cancer in TCGA. **A**, The MKNK2 expression significantly correlated with the infiltration levels of various immune cells in the TIMER database. **B**, Correlation between MKNK2 expression and immune checkpoint. **C**, CIBERSORT predicts that MKNK2 expression is correlated with immunocytes. **D**, EPIC predicts that MKNK2 expression is correlated with immunocytes. *Indicates statistical significance, where *p<0.05, *p<0.01, ***p<0.001.

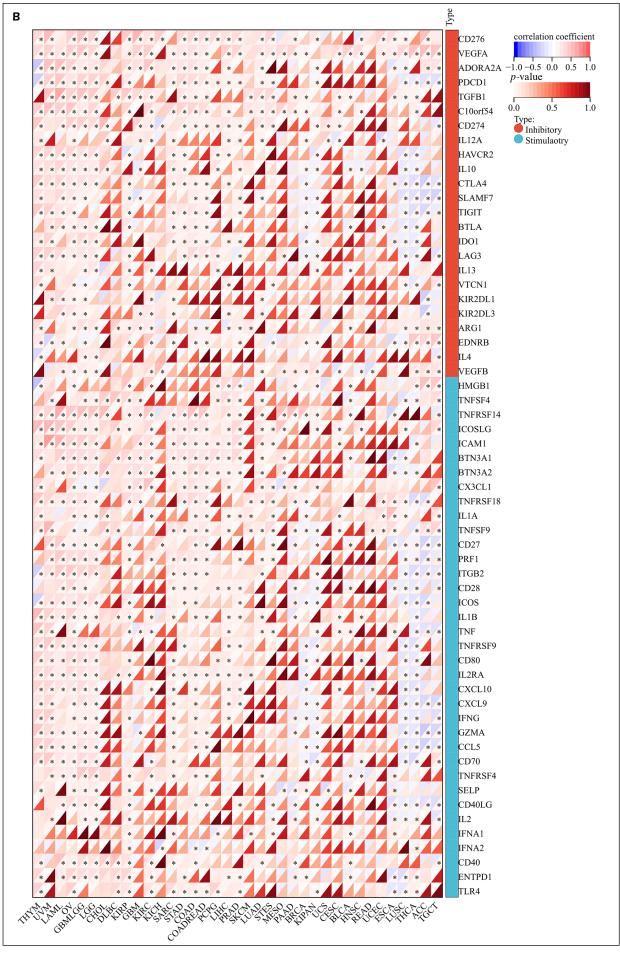


Figure 6 (*Continued*). Different algorithms were used to explore the potential correlation between the expression level of the MKNK2 gene and the level of immune cell infiltration in all types of cancer in TCGA. **A**, The MKNK2 expression significantly correlated with the infiltration levels of various immune cells in the TIMER database. **B**, Correlation between MKNK2 expression and immune checkpoint. **C**, CIBERSORT predicts that MKNK2 expression is correlated with immunocytes. **D**, EPIC predicts that MKNK2 expression is correlated with immunocytes. *Indicates statistical significance, where *p<0.05, **p<0.01, ***p<0.001.

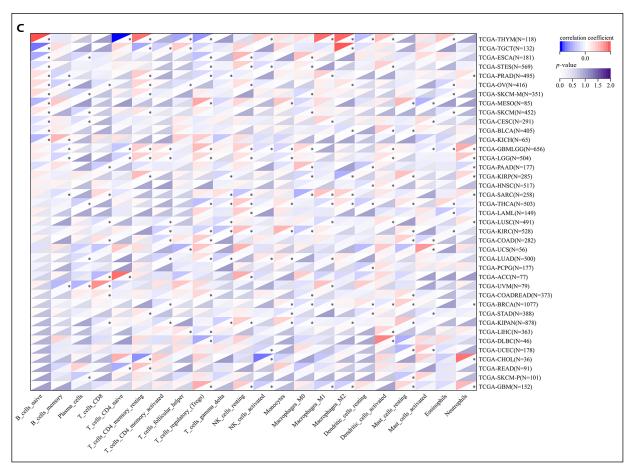


Figure 6 *(Continued).* Different algorithms were used to explore the potential correlation between the expression level of the MKNK2 gene and the level of immune cell infiltration in all types of cancer in TCGA. **A**, The MKNK2 expression significantly correlated with the infiltration levels of various immune cells in the TIMER database. **B**, Correlation between MKNK2 expression and immune checkpoint. **C**, CIBERSORT predicts that MKNK2 expression is correlated with immunocytes. **D**, EPIC predicts that MKNK2 expression is correlated with immunocytes. *Indicates statistical significance, where *p<0.05, **p<0.01, ***p<0.001.

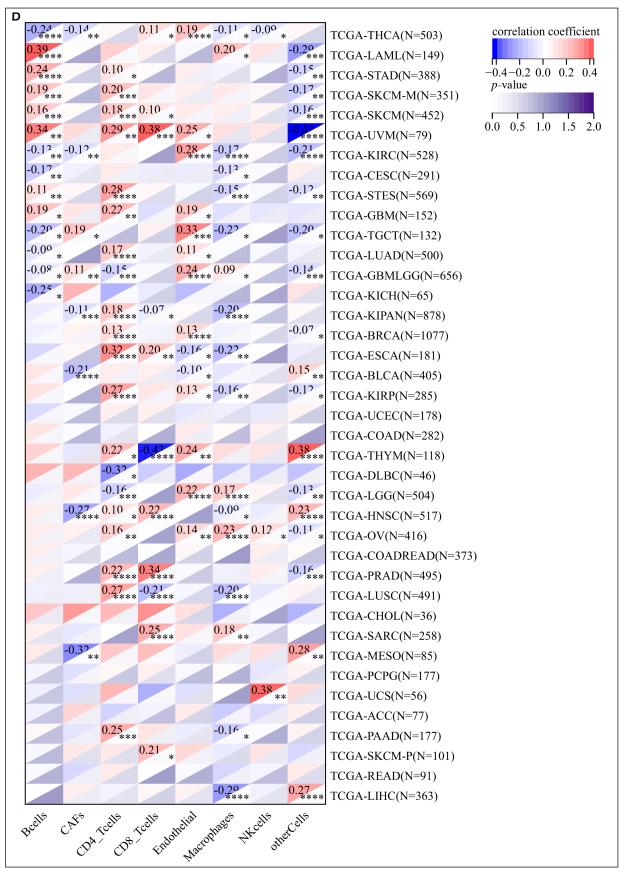


Figure 6 *(Continued).* Different algorithms were used to explore the potential correlation between the expression level of the MKNK2 gene and the level of immune cell infiltration in all types of cancer in TCGA. **A**, The MKNK2 expression significantly correlated with the infiltration levels of various immune cells in the TIMER database. **B**, Correlation between MKNK2 expression and immune checkpoint. **C**, CIBERSORT predicts that MKNK2 expression is correlated with immunocytes. **D**, EPIC predicts that MKNK2 expression is correlated with immunocytes. *Indicates statistical significance, where *p<0.05, **p<0.01, ***p<0.001.

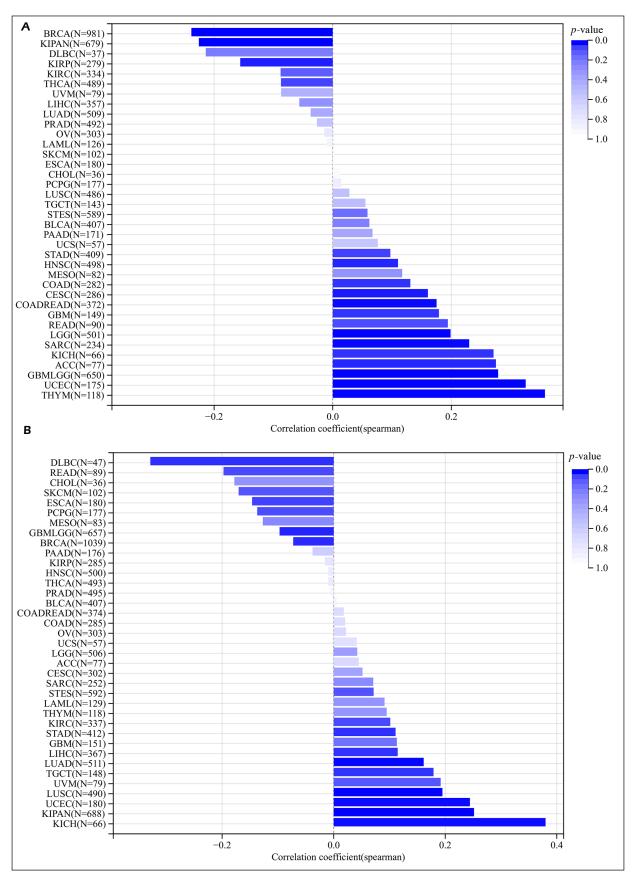


Figure 7. Relationship between MKNK2 expression and tumor mutation burden (**A**), microsatellite Instability (**B**), and Stemness Scores DNAss (**C**), RNAss (**D**) in pan-cancer.

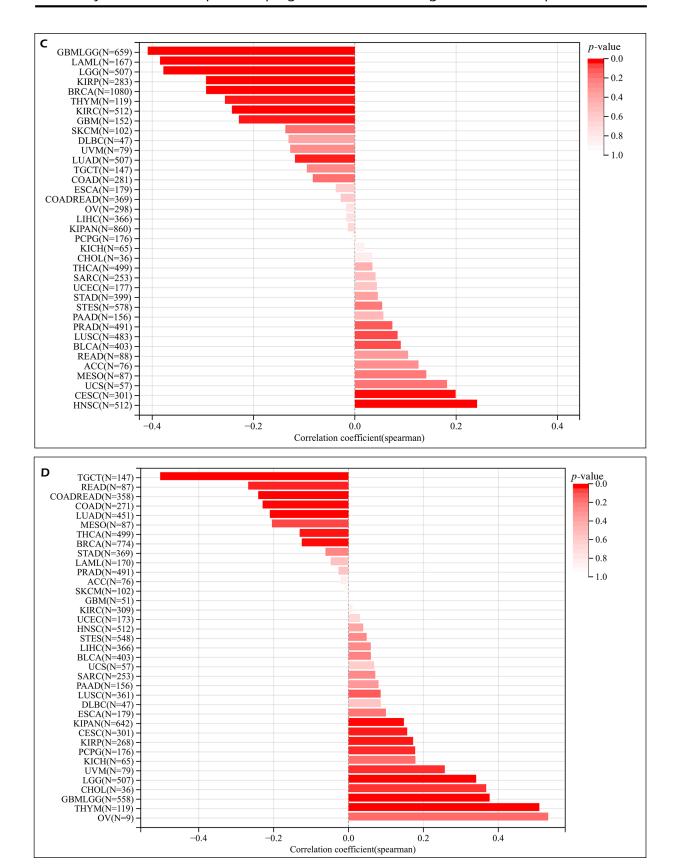


Figure 7 *(Continued).* Relationship between MKNK2 expression and tumor mutation burden (A), microsatellite Instability (B), and Stemness Scores DNAss (C), RNAss (D) in pan-cancer.

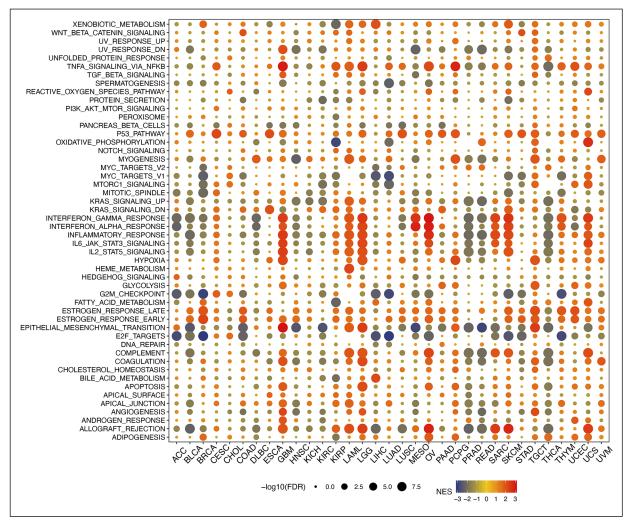


Figure 8. The hallmarks gene set enrichment analysis of *MKNK2* in pan-cancer. The size of the circle represents the FDR value of the enriched term in each cancer, and the color indicates the normalized enrichment score (NES) of each term.

development of targeted therapies that leverage the immunological and stemness-related roles of *MKNK2*.

GSEA of MKNK2 in Pan-Cancer

To delve deeper into the molecular underpinnings of MKNK2's role in cancer, we conducted Gene Set Enrichment Analysis (GSEA) utilizing the differentially expressed genes (DEGs) between subgroups characterized by low and high MKNK2 expression within each cancer type. This approach was instrumental in identifying cancer hallmarks associated with MKNK2. Our analysis revealed a striking correlation between MKNK2 expression and a spectrum of immune-related pathways. Notably, these included $TNF-\alpha$ signaling via NF-κB, IFN- α response, IFN- γ response, inflammatory

response, *IL6-JAK-STAT3* signaling, *IL-2-STAT5* signaling, and allograft-rejection pathways, with particularly strong associations observed in cancers such as ESCA, KIRP, LAML, MESO, SARC, and UCEC (Figure 8). The pathways $TNF-\alpha$ signaling via NF- κ B and JAK-STAT3 signaling, which are implicated in our findings, have been well-documented in the literature for their critical role in sustaining the stemness of tumor stem cells²².

Moreover, our study also identified a tight association between *MKNK2* expression and key cancer-related processes such as epithelial-mesenchymal transition (EMT) and the *p53* pathway. EMT is recognized for its contribution to cancer metastasis, while the *p53* pathway is a key regulator of cellular responses to DNA damage and a well-known tumor suppressor.

In conclusion, the collective results of our analyses suggest that elevated *MKNK2* expression is intricately linked to the immune-activation status of various cancers. These findings not only shed light on the multifaceted roles of *MKNK2* in the immune landscape of cancers but also provide a foundation for further research into the functions and roles of *MKNK2* in the initiation and progression of cancer. Understanding these mechanisms could pave the way for novel therapeutic strategies that leverage the immunomodulatory effects of *MKNK2*.

Discussion

Multiple investigations prove the association of MKNK2 with tumor progression, prognosis, and treatment response^{4,23}. The MKNK2 kinase, which is encoded by the gene MKNK2, is important for MAPK signaling and is linked with oncogenesis^{20,24}. Many well-known kinase networks, such as EGFR, MAPKs, and c-Src also take part in tumor pathogenesis²⁵⁻²⁸. The MAP kinase-interacting serine/threonine-protein kinases (MNKS) is a downstream protein of MAPKs that can phosphorylate eIF4E and increase oncogenic mRNA translation^{23,29}. The literature suggests that MKNK2 not only modulates some physiological processes but also regulates tumor growth and immune infiltration (Figure 6), but studies reporting its importance in the cancer field are still missing²⁷. Recently, a relationship between MKNK2 expression and tumor cell progression (mainly in GBM and Gastric Cancer) was acknowledged^{5,27}. Therefore, it can be speculated that MKNK2 expression may affect a patient's survival via tumor cell progression (Figures 2 and 3). Consistent with previous research, this study also noted that MKNK2 has diverse activity in pan-cancers²⁸⁻³⁰. The differential MKNK2 expression in pan-cancers and their matched para-cancer normal tissues was evaluated. Results may differ because of the variability in data sources, collection approach, and the number of cancers in the investigation cohort. Nonetheless, the three databases revealed consistently poor prognostic value with MKNK2 expression in GBMLGG, ACC, and STAD (Figures 2 and 3).

In association with *MKNK2*'s influence on tumor immunosuppressive microenvironment and antitumor immune response *via* the *MNK1/2-eIF4E* axis, the systematic analysis further correlated *MKNK2* and tumor immunity³¹. The relationship between *MKNK2* expression and tumor immune

cell infiltration levels is mostly positively associated. Based on TIMER analysis, it was found that MKNK2 levels were substantially correlated with the degree of CD4+ T, CD8+ T, B, neutrophils, macrophages, and dendritic cell infiltration (Figure 6A). Tumor-associated macrophages (TAMs) are the main part of the immune microenvironment of many tumors, which is closely linked with tumor growth, invasion, and drug resistance³². The available research results show that MNK2 can govern the macrophage anti-inflammatory phenotype¹⁶. Analysis results of immune cell infiltration show that MKNK2 is significantly associated with macrophages in many tumors, suggesting that it induces metabolic reprogramming and regulates the function of TAMs. Although the high MKNK2 expression is associated with a substandard prognosis of ACC, the MKNK2 expression correlation with immune cell infiltration in ACC is very low, suggesting that MKNK2 may affect the prognosis of ACC through non-immune pathways. MKNK2 expression was also widely associated with immune checkpoints. MNK1/2-eIF4E axis is considered a potential therapeutic index in melanoma³³. Dysregulated mRNA translation is crucial for tumorigenesis. MAPK interacting kinases (MNK)1/2 are key mRNA translation regulators, integrating oncogenic and immune pathways signals *via* phosphorylation of eIF4E and other mRNA binding proteins³⁴. Although the expression of MKNK2 is linked with immune infiltration and patient survival, it has not been demonstrated that MKNK2 affects patient's survival through immune infiltration.

The correlation of MKNK2 with TMB, MSI, and Stemness in various cancer types was also investigated in our study (Figure 7). It was reported that inhibition of MNKs with small molecule inhibitors or knockdown of MKNK1 and MKNK2 disrupts the growth of tumor cells and prevents tumor growth in vivo35. TMB levels affect immunogenic peptide generation, thus affecting the patient's response to immune checkpoint inhibitors³⁶. MSI is a vital index for predicting tumorigenesis and development³⁷. We found that MKNK2 expression is highly related to TMB, MSI, and Stemness in most cancer types. GBM-LGG, KIPAN, STAD, UCEC, BRCA, and other cancer species showed a significant correlation in TMB and MSI scores (Figure 7A-B).

The GSEA result suggests that *PDIA3* is closely associated with immune-activated processes, such as $TNF-\alpha$ signaling via NF- κ B, IFN- α response, IFN- γ response, inflammatory-response,

IL6-JAK-STAT3 signaling, *IL-2-STAT5* signaling, and allograft-rejection pathways, but completely opposite results were observed in distinct cancer types. For example, these processes were mostly significantly enriched in high-*MKNK2* cancer subgroups, but reversed results were found in ACC, BLCA, PCPG, PRAD, and TGCT (Figure 8). This indicated that *PDIA3* might play different roles in distinct cancer types.

Recent studies demonstrated that pharmacological inhibition of *MNKs* could block *eIF4E*-mediated translation and thus suppress tumor cell proliferation and slow down tumor growth *in vivo*^{27,31}. *eIF4E*-mediated mRNA translation plays an important role in oncogenic transformation, and an elevated level of *eIF4E* expression/activity has been observed in many tumors¹⁷. *MNK* activity also plays an important role in regulating the innate and adaptive immune systems^{38,39}. Interference with the reprogramming of immune cells in the immunosuppressive microenvironment and the stemness maintenance of tumor stem cells could be an underlying regulatory mechanism associated with *MKNK2* and bad prognosis^{16,27}.

The present study unveiled a complicated role of *MKNK2* aberrant expression in cancer progression and patient outcome that warrants further investigation. Since our study was based on bioinformatics and relies on public databases, there are major limitations. First, the quality of data collection and the method used to generate the data could be inconsistent depending on the sources. This could affect the results of different analyses. Secondly, the results and the conclusions are not experimentally or prospectively confirmed in the laboratory or clinic. A future investigation is required to validate *MKNK2* expression and function *in vivo* and *in vitro*.

Altogether, this pan-cancer analysis indicated the clinical importance of *MKNK2* in prognosis, stemness, immune cell infiltration, and immunity indices such as TMB and MSI in understanding its activity in tumorigenesis.

Conclusions

This investigation comprehensively evaluated the prognostic significance and immunological aspects of *MKNK2* in pan-cancer. *MKNK2* is differentially expressed in many tumors, and its abnormal expression is linked with tumor progression, immune checkpoint genes, immune cell infiltration, MSI, TMB, and stemness, especially

in GBMLGG. Therefore, it may be a potent prognostic physiological marker.

This article only describes the correlation between *MKNK2* expression and tumor immunity from the perspective of bioinformatic analysis. Future prospective and experimental studies on *MKNK2* expression and immune cell infiltration in different cancer populations may provide more ideas and channels for the development of tumor mechanisms and therapeutic strategies targeting *MKNK2* to improve the therapeutic effect of immunotherapy.

Conflicts of Interest

All authors declare no conflict of interest.

Ethics Approval and Informed Consent

The data used in this article belongs to public databases. The patients involved in the databases have received ethical approval with informed consent, which could be downloaded for free to conduct research and publish relevant articles. This study is based on open resource data, so there are no ethical issues or other conflicts of interest.

Data Availability

The data that support the findings of this study are openly available in https://www.cancer.gov/about-nci/organization/ccg/research/structural-genomics/tcga and https://portal.gdc.cancer.gov/. Stemness indices across all tumor samples/types can be accessed at https://bioinformaticsfmrp.github.io/Pan-CanStem_Web. TCGA, GTEx, and CCLE belong to public databases. The patients involved in the database have obtained ethical approval. Users can download relevant data for free for research and publish relevant articles.

Acknowledgments

We acknowledge the Cancer Genome Atlas (TCGA), Genotype-Tissue Expression (GTEx), and Cancer Cell Line Encyclopedia (CCLE) database for providing their platforms and contributors for uploading their meaningful datasets.

Authors' Contributions

Yiming Zhang: writing original draft editing, original draft, methodology, validation, and visualization. Jikang Fan: original draft, methodology, validation. Xuya Wang: original draft, methodology, validation. Jie Liu: methodology, validation, and visualization. Xisen Wang and Tao Li: methodology and visualization. Xuejun Yang: original draft and supervision.

Funding

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors. The authors acknowledge the support from their respective institutions for the publication of this research.

Al Disclosure

In conducting this study, we have employed artificial intelligence tools for data analysis and visualization where appropriate. The use of AI has been confined to tasks that enhance the efficiency and accuracy of our research without replacing the intellectual input and decision-making processes of the research team. The authors thoroughly reviewed and validated all AI-generated outputs to ensure the integrity and reliability of the study's findings.

ORCID ID

Yiming Zhang: 0009-0009-1285-8112

References

- Deo SVS, Sharma J, Kumar S. GLOBOCAN 2020 Report on Global Cancer Burden: Challenges and Opportunities for Surgical Oncologists. Ann Surg Oncol 2022; 29: 6497-6500.
- Srivastava S, Hanash S. Pan-Cancer Early Detection: Hype or Hope? Cancer Cell 2020; 38: 23-24.
- Hou J, Lam F, Proud C, Wang S. Targeting Mnks for cancer therapy. Front Oncotarget 2012; 3: 118-131.
- 4) Xie J, Shen K, Jones AT, Yang J, Tee AR, Shen MH, Yu M, Irani S, Wong D, Merrett JE, Lenchine RV, De Poi S, Jensen KB, Trim PJ, Snel MF, Kamei M, Martin SK, Fitter S, Tian S, Wang X, Butler LM, Zannettino ACW, Proud CG. Reciprocal signaling between mTORC1 and MNK2 controls cell growth and oncogenesis. Cell Mol Life Sci 2021; 78: 249-270.
- Qin M, Liang Z, Qin H, Huo Y, Wu Q, Yang H, Tang G. Novel Prognostic Biomarkers in Gastric Cancer: CGB5, MKNK2, and PAPPA2. Front Oncol 2021; 11: 683582.
- 6) Liu B, Sun Y, Tang M, Liang C, Huang CP, Niu Y, Wang Z, Chang C. The miR-361-3p increases enzalutamide (Enz) sensitivity via targeting the ARv7 and MKNK2 to better suppress the Enz-resistant prostate cancer. Cell Death Dis 2020; 11: 807.
- Mogilevsky M, Shimshon O, Kumar S, Mogilevsky A, Keshet E, Yavin E, Heyd F, Karni R. Modulation of MKNK2 alternative splicing by splice-switching oligonucleotides as a novel approach for glioblastoma treatment. Nucleic Acids Res 2018; 46: 11396-11404.
- Bejarano L, Jordão MJC, Joyce JA. Therapeutic Targeting of the Tumor Microenvironment. Cancer Discov 2021; 11: 933-959.
- 9) He X, Smith SE, Chen S, Li H, Wu D, Meneses-Giles PI, Wang Y, Hembree M, Yi K, Zhao X, Guo F, Unruh JR, Maddera LE, Yu Z, Scott A, Perera A, Wang Y, Zhao C, Bae K, Box A, Haug JS, Tao F, Hu D, Hansen DM, Qian P, Saha S, Dixon D, Anant S, Zhang D, Lin EH, Sun W, Wiedemann LM, Li

- L. Tumor-initiating stem cell shapes its microenvironment into an immunosuppressive barrier and pro-tumorigenic niche. Cell Rep 2021; 36: 109674.
- Lei X, Lei Y, Li JK, Du WX, Li RG, Yang J, Li J, Li F, Tan HB. Immune cells within the tumor microenvironment: Biological functions and roles in cancer immunotherapy. Cancer Lett 2020; 470: 126-133.
- Li B, Chan HL, Chen P. Immune Checkpoint Inhibitors: Basics and Challenges. Curr Med Chem 2019; 26: 3009-3025.
- 12) Liu J, Lichtenberg T, Hoadley KA, Poisson LM, Lazar AJ, Cherniack AD, Kovatich AJ, Benz CC, Levine DA, Lee AV, Omberg L, Wolf DM, Shriver CD, Thorsson V; Cancer Genome Atlas Research Network; Hu H. An Integrated TCGA Pan-Cancer Clinical Data Resource to Drive High-Quality Survival Outcome Analytics. Cell 2018; 173: 400-416.e11.
- 13) Thorsson V, Gibbs DL, Brown SD, Wolf D, Bortone DS, Ou Yang TH, Porta-Pardo E, Gao GF, Plaisier CL, Eddy JA, Ziv E, Culhane AC, Paull EO, Sivakumar IKA, Gentles AJ, Malhotra R, Farshidfar F, Colaprico A, Parker JS, Mose LE, Vo NS, Liu J, Liu Y, Rader J, Dhankani V, Reynolds SM, Bowlby R, Califano A, Cherniack AD, Anastassiou D, Bedognetti D, Mokrab Y, Newman AM, Rao A, Chen K, Krasnitz A, Hu H, Malta TM, Noushmehr H, Pedamallu CS, Bullman S, Ojesina AI, Lamb A, Zhou W, Shen H, Choueiri TK, Weinstein JN, Guinney J, Saltz J, Holt RA, Rabkin CS; Cancer Genome Atlas Research Network; Lazar AJ, Serody JS, Demicco EG, Disis ML, Vincent BG, Shmulevich I. The Immune Landscape of Cancer. Immunity 2018; 48: 812-830.e14. Erratum in: Immunity 2019; 51: 411-412.
- 14) Malta TM, Sokolov A, Gentles AJ, Burzykowski T, Poisson L, Weinstein JN, Kamińska B, Huelsken J, Omberg L, Gevaert O, Colaprico A, Czerwińska P, Mazurek S, Mishra L, Heyn H, Krasnitz A, Godwin AK, Lazar AJ; Cancer Genome Atlas Research Network; Stuart JM, Hoadley KA, Laird PW, Noushmehr H, Wiznerowicz M. Machine Learning Identifies Stemness Features Associated with Oncogenic Dedifferentiation. Cell 2018; 173: 338-354.e15.
- Yu G, Wang LG, Han Y, He QY. clusterProfiler: an R package for comparing biological themes among gene clusters. OMICS 2012; 16: 284-287.
- 16) Bartish M, Tong D, Pan Y, Wallerius M, Liu H, Ristau J, de Souza Ferreira S, Wallmann T, van Hoef V, Masvidal L, Kerzel T, Joly AL, Goncalves C, Preston SEJ, Ebrahimian T, Seitz C, Bergh J, Pietras K, Lehoux S, Naldini L, Andersson J, Squadrito ML, Del Rincón SV, Larsson O, Rolny C. MNK2 governs the macrophage antiinflammatory phenotype. Proc Natl Acad Sci U S A 2020; 117: 27556-27565.
- 17) Huang B, Guo S, Zhang Y, Lin P, Lin C, Chen M, Zhu S, Huang L, He J, Zhang L, Zheng Y, Wen Z. MiR-223-3p alleviates trigeminal neuropathic pain in the male mouse by targeting MKNK2 and MAPK/ ERK signaling. Brain Behav 2022; 12: e2634.
- 18) Liu H, Gong Z, Li K, Zhang Q, Xu Z, Xu Y. SRPK1/2 and PP1α exert opposite functions by modulating SR-

- SF1-guided MKNK2 alternative splicing in colon adenocarcinoma. J Exp Clin Cancer Res 2021; 40: 75.
- 19) Lim S, Saw TY, Zhang M, Janes MR, Nacro K, Hill J, Lim AQ, Chang CT, Fruman DA, Rizzieri DA, Tan SY, Fan H, Chuah CT, Ong ST. Targeting of the MNK-eIF4E axis in blast crisis chronic myeloid leukemia inhibits leukemia stem cell function. Proc Natl Acad Sci U S A 2013; 110: E2298-E2307.
- 20) Grzmil M, Morin P Jr, Lino MM, Merlo A, Frank S, Wang Y, Moncayo G, Hemmings BA. MAP kinase-interacting kinase 1 regulates SMAD2-dependent TGF-β signaling pathway in human glioblastoma. Cancer Res 2011; 71: 2392-2402.
- 21) Beier CP, Kumar P, Meyer K, Leukel P, Bruttel V, Aschenbrenner I, Riemenschneider MJ, Fragoulis A, Rümmele P, Lamszus K, Schulz JB, Weis J, Bogdahn U, Wischhusen J, Hau P, Spang R, Beier D. The cancer stem cell subtype determines immune infiltration of glioblastoma. Stem Cells Dev 2012; 21: 2753-2761.
- 22) Wang T, Fahrmann JF, Lee H, Li YJ, Tripathi SC, Yue C, Zhang C, Lifshitz V, Song J, Yuan Y, Somlo G, Jandial R, Ann D, Hanash S, Jove R, Yu H. JAK/ STAT3-Regulated Fatty Acid β-Oxidation Is Critical for Breast Cancer Stem Cell Self-Renewal and Chemoresistance. Cell Metab 2018; 27: 136-150.e5.
- 23) Wang J, Da C, Su Y, Song R, Bai Z. MKNK2 enhances chemoresistance of ovarian cancer by suppressing autophagy via miR-125b. Biochem Biophys Res Commun 2021; 556: 31-38.
- 24) Tan B, Yang G, Su L, Zhou J, Wu Y, Liang C, Lai Y. MiR-125b targeted regulation of MKNK2 inhibits multiple myeloma proliferation and invasion. Am J Transl Res 2024; 16: 3366-3375.
- 25) Li Z, Zhao PL, Gao X, Li X, Meng YQ, Li ZQ, Zhai KR, Wei SL, Feng HM, Huang HR, Li B. DUS4L suppresses invasion and metastasis in LUAD via modulation of PI3K/AKT and ERK/MAPK signaling through GRB2. Int Immunopharmacol 2024; 142(Pt A): 113043.
- 26) Shiers S, Sahn JJ, Price TJ. MNK1 and MNK2 Expression in the Human Dorsal Root and Trigeminal Ganglion. Neuroscience 2023; 515: 96-107.
- 27) Bell JB, Eckerdt FD, Alley K, Magnusson LP, Hussain H, Bi Y, Arslan AD, Clymer J, Alvarez AA, Goldman S, Cheng SY, Nakano I, Horbinski C, Davuluri RV, James CD, Platanias LC. MNK Inhibition Disrupts Mesenchymal Glioma Stem Cells and Prolongs Survival in a Mouse Model of Glioblastoma. Mol Cancer Res 2016; 14: 984-993.
- 28) Wang J, Da C, Su Y, Song R, Bai Z. MKNK2 enhances chemoresistance of ovarian cancer by suppressing autophagy via miR-125b. Biochem Biophys Res Commun 2021; 556: 31-38.
- 29) Konicek BW, Stephens JR, McNulty AM, Robichaud N, Peery RB, Dumstorf CA, Dowless MS, Iversen PW, Parsons S, Ellis KE, McCann DJ, Pelletier J, Furic L, Yingling JM, Stancato LF, Sonenberg N, Graff JR. Therapeutic inhibition of MAP kinase interacting kinase blocks eukaryotic initiation factor 4E phosphorylation and suppres-

- ses outgrowth of experimental lung metastases. Cancer Res 2011; 71: 1849-1857.
- 30) Dreas A, Mikulski M, Milik M, Fabritius CH, Brzozka K,Rzymski T. Mitogen-activated Protein Kinase (MAPK) Interacting Kinases 1 and 2 (MNK1 and MNK2) as Targets for Cancer Therapy: Recent Progress in the Development of MNK Inhibitors. Curr Med Chem 2017; 24: 3025-3053.
- 31) Huang F, Gonçalves C, Bartish M, Rémy-Sarrazin J, Issa ME, Cordeiro B, Guo Q, Emond A, Attias M, Yang W, Plourde D, Su J, Gimeno MG, Zhan Y, Galán A, Rzymski T, Mazan M, Masiejczyk M, Faber J, Khoury E, Benoit A, Gagnon N, Dankort D, Journe F, Ghanem GE, Krawczyk CM, Saragovi HU, Piccirillo CA, Sonenberg N, Topisirovic I, Rudd CE, Miller WH Jr, Del Rincón SV. Inhibiting the MNK1/2-eIF4E axis impairs melanoma phenotype switching and potentiates antitumor immune responses. J Clin Invest 2021; 131: e140752.
- Vitale I, Manic G, Coussens LM, Kroemer G, Galluzzi L. Macrophages and Metabolism in the Tumor Microenvironment. Cell Metab 2019; 30: 36-50.
- 33) Prabhu SA, Moussa O, Miller WH Jr, Del Rincón SV. The MNK1/2-elF4E Axis as a Potential Therapeutic Target in Melanoma. Int J Mol Sci 2020; 21: 4055.
- 34) Reich SH, Sprengeler PA, Chiang GG, Appleman JR, Chen J, Clarine J, Eam B, Ernst JT, Han Q, Goel VK, Han EZR, Huang V, Hung INJ, Jemison A, Jessen KA, Molter J, Murphy D, Neal M, Parker GS, Shaghafi M, Sperry S, Staunton J, Stumpf CR, Thompson PA, Tran C, Webber SE, Wegerski CJ, Zheng H, Webster KR. Structure-based Design of Pyridone-Aminal eFT508 Targeting Dysregulated Translation by Selective Mitogen-activated Protein Kinase Interacting Kinases 1 and 2 (MNK1/2) Inhibition. J Med Chem 2018; 61: 3516-3540.
- 35) Ueda T, Sasaki M, Elia AJ, Chio II, Hamada K, Fukunaga R, Mak TW. Combined deficiency for MAP kinase-interacting kinase 1 and 2 (Mnk1 and Mnk2) delays tumor development. Proc Natl Acad Sci U S A 2010; 107: 13984-13990.
- 36) Wu HX, Wang ZX, Zhao Q, Chen DL, He MM, Yang LP, Wang YN, Jin Y, Ren C, Luo HY, Wang ZQ, Wang F. Tumor mutational and indel burden: a systematic pan-cancer evaluation as prognostic biomarkers. Ann Transl Med 2019; 7: 640.
- Li K, Luo H, Huang L, Luo H, Zhu X. Microsatellite instability: a review of what the oncologist should know. Cancer Cell Int 2020; 20: 16.
- Pham TND, Spaulding C, Munshi HG. Controlling TIME: How MNK Kinases Function to Shape Tumor Immunity. Cancers (Basel) 2020; 12: 2096.
- 39) Sharma S, Singh M, Chiranjivi AK, Dadwal A, Ahmed S, Asthana S, Das S. Structural insights into trypanosomatid Mnk kinase orthologues (kMnks) suggest altered mechanism in the kinase domain. Int J Biol Macromol. 2024 Oct;277(Pt 3):134428. doi: 10.1016/j.ijbiomac.2024.134428. Epub 2024 Aug 7. Erratum in: Int J Biol Macromol 2024; 279: 135530.