

Critical genes of hepatocellular carcinoma revealed by network and module analysis of RNA-seq data

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Abstract. – **OBJECTIVE:** RNA-seq data of hepatocellular carcinoma (HCC) was analyzed to identify critical genes related to the pathogenesis and prognosis.

MATERIALS AND METHODS: Three RNA-seq datasets of HCC (GSE69164, GSE63863 and GSE55758) were downloaded from Gene Expression Omnibus (GEO), while another dataset including 54 HCC cases with survival time was obtained from The Cancer Genome Atlas (TCGA). Differentially expressed genes (DEGs) were identified by significant analysis of microarrays (SAM) method using package samr of R. As followed, we constructed a protein-protein interaction (PPI) network based on the information in Human Protein Reference Database (HPRD). Modules in the PPI network were identified with MCODE method using plugin clusterViz of Cytoscape. Gene Ontology (GO) enrichment analysis and pathway enrichment analysis were performed with DAVID. The difference in survival curves was analyzed with Kaplan-Meier (K-M) method using package survival.

RESULTS: A total of 2572 DEGs were identified in the 3 datasets from GEO (GSE69164, GSE63863 and GSE55758). The PPI network was constructed including 660 nodes and 1008 edges, and 4 modules were disclosed in the network. Module A (containing 244 DEGs) was found to related to HCC closely, which genes were involved in transcription factor binding, protein metabolism as well as regulation of apoptosis. Nine hub genes were identified in the module A, including PRKCA, YWHAZ, KRT18, NDRG1, HSPA1A, HSP90AA1, HSF1, IKGKB and UBE21. The network provides the protein-protein interaction of these critical genes, which were implicated in the pathogenesis of HCC. Survival analysis showed that there is a significant difference between two groups classified by the genes in module A. Further Univariate Cox regression analysis showed that 72 genes were associated with survival

time significantly, such as NPM1, PRKDC, SPARC, HMGA1, COL1A1 and COL1A2.

CONCLUSIONS: Nine critical genes related to the pathogenesis and 72 potential prognostic markers were revealed in HCC by the network and module analysis of RNA-seq data. These findings could improve the understanding of the pathogenesis and provide valuable information to further investigate the prognostic markers of HCC.

Key Words:

Hepatocellular carcinoma, Differentially expressed genes, Protein-protein interaction network, Hub genes, Survival analysis.

Introduction

Hepatocellular carcinoma (HCC) is one of the most common and lethal malignancies worldwide, which has a poor prognosis partly because of the extreme variability of the clinical outcome. Previous studies indicated that several biological pathways attributed to this heterogeneous clinical behavior.

A variety of pathways has been implicated in the pathogenesis HCC or related to the patients' survival time. Some pathways contribute to the progression and metastasis of HCC, such as Wnt/ β -catenin pathway^{1,2}, DNMT1/PTEN/Akt pathway³, ERK/c-Myc pathway⁴. Other pathways are associated with chemoresistance, such as Akt/PKB survival pathway⁵ and IGF-1R signaling pathway⁶. Therapeutic targets are also found in signaling pathways, such as RalA signaling pathway⁷ and Rac signaling pathway⁸. Meanwhile, some biomarkers have been revealed. MicroRNA-145 is regarded as a promising biomark-

er in HCC⁹. Alpha-Methylacyl-CoA racemase (AMACR) can serve as a prognostic biomarker for the early recurrence/metastasis of HCC¹⁰. Progressive increase of SCCA-IgM immune complexes in cirrhotic patients is associated with the development of hepatocellular carcinoma¹¹. NDRG1 is found as a biomarker for metastasis, recurrence and of poor prognosis in HCC¹².

In a word, many researchers have revealed the important pathways and biomarkers in the HCC, some of which have been applied to the clinical stage. However, the improvement of cancer pathogenesis understanding and bioinformatics methods provide the possibility to further study the HCC pathogenesis by novel analysis method. Although many high throughputs sequencing-based transcriptome analysis were carried out to disclose critical genes in HCC¹³⁻¹⁷, they focused on metastasis or promoter methylation. In the present study, we try to analyze the data from several RNA-seq datasets with network and module analysis method, to mine valuable information about HCC.

Differentially expressed genes (DEGs) were identified from RNA-seq data of HCC and then protein-protein interaction (PPI) network analysis was performed to unveil critical genes. Besides, genes of prognostic effect were also revealed. These findings could not only supplement the understanding of the pathogenesis of HCC, but also provide potential prognostic markers for HCC.

Materials and Methods

Gene Expression Data

Three RNA-seq datasets (GSE69164, GSE63863 and GSE55758) were downloaded from Gene Expression Omnibus (GEO). The RNA-seq data of all three datasets were acquired using Illumina's platforms. Dataset GSE69164 included 11 paired normal and HCC samples. Dataset GSE63863 contained 12 paired normal and HCC samples. Dataset GSE55758 consisted of 8 paired normal and HCC samples. The survival time of HCC cases was included. Another RNA-seq dataset was obtained from The Cancer Genome Atlas (TCGA) and 54 HCC cases with survival time were selected out as the valid dataset.

Genes with missing value more than 20% were excluded, and others were filled with average value. Log₂ transformation was then applied to the following analysis.

Screening of DEGs

DEGs were identified by significant analysis of microarrays (SAM) method¹⁸ using package *samR*¹⁹ of *R*. This method has the advantage in control false discovery rate (FDR) in multiple testing. The relative difference (*d*) was calculated as following formula:

$$d = \frac{X_1 - X_2'}{S + S_0}$$

Where X_1' and X_2' represent average expression levels of certain gene in two different status, and *s* represents the variance of the gene.

FDR < 0.05 and fold change > 2 were set as the threshold to screen out DEGs.

Construction of Protein-Protein Interaction (PPI) Network

Protein-protein interactions (PPIs) are the basis of biological processes. Investigations on PPIs can provide clues to understand the molecular mechanism of diseases. In the present study, PPI network was constructed based on the information from Human Protein Reference Database (HPRD)²⁰, which contained the largest number of proteins in the currently available databases.

Identification of modules

Modules were identified with MCODE method²¹ using plugin *clusterViz* of CytoScape²². The method consisted of three steps: (1) calculate weights for nodes; (2) predict complex; (3) filter complex according to certain criteria. Clustering coefficient (*C_i*) was chosen as the weight of node:

$$C_i = 2 \times n / K_i \times (K_i - 1)$$

Where *K_i* represents the number of nodes directly connected to node *i*; *n* represents number of edges among the *K_i* nodes.

Node with maximum weight was regarded as seed and then node *j* with weight ratio (W_j/W_{seed}) greater than threshold was included in the module.

Functional Enrichment Analysis

Gene Ontology (GO) enrichment analysis and pathway enrichment analysis were performed with DAVID (Database for Annotation, Visualization and Integration Discovery, <http://david.abcc.ncifcrf.gov/>)²³, an enrichment test based upon hypergeometric distribution.

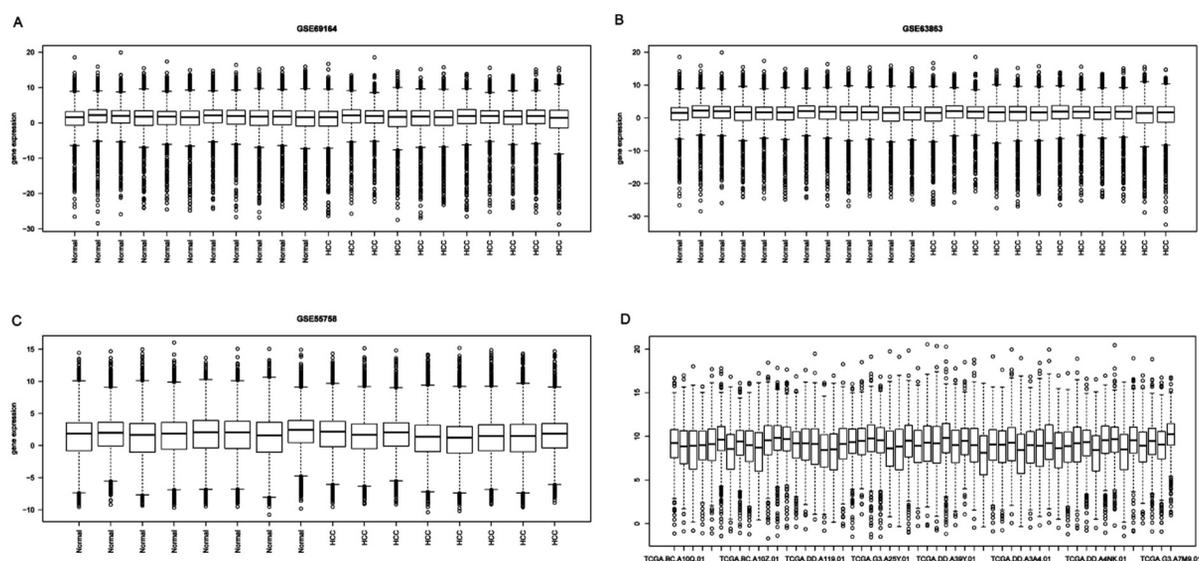


Figure 1. Box plots of pre-treated RNA-seq data. Dataset GSE69164 (A), Dataset GSE63863 (B), Dataset GSE55758 (C) and Dataset from TCGA (D).

Survival Analysis

Difference in survival curves was analyzed with Kaplan-Meier (K-M) method using package *survival*. Univariate Cox regression analysis was performed to disclose genes associated with survival time. p -value < 0.05 was set as the cut-off.

Results

Pre-treated Gene Expression Data

After pre-treatment, dataset GSE69164, GSE63863 and GSE55758 contained 17059, 17110 and 17750 non-redundant genes, respectively (Figure 1), while dataset from TCGA contained 17781 non-redundant genes (Figure 1D). According to the box plots, a good performance of normalization was achieved for all the 4 datasets.

Differentially Expressed Genes

A total of 2439 DEGs were identified in dataset GSE69164, and 2424 DEGs were identified in dataset GSE63863, and 410 DEGs were identified in dataset GSE55857. The 3 sets of DEGs were combined to a union set, which contained 2572 DEGs.

To validate the reliability of the DEGs, principal component analysis (PCA) was performed. As shown in Figure 2, we found that the 3 sets of DEGs have good distinguish effect to separate

HCC samples from normal samples, suggesting these DEGs were related to HCC actually.

PPI Network and Modules

A total of 39240 PPI interactions were obtained from HPRD. The 2572 DEGs were mapped into the network and interactions between two DEGs were selected out. Thus, a HCC-related PPI network was constructed, which contained 660 nodes and 1008 pair of interactions.

Four modules were unveiled in the HCC-related PPI network using plugin *clusterViz* of Cytoscape. After careful checking, we found that HCC-related genes were enriched in Module A (including a total of 244 DEGs, Figure 3). The genes were involved in transcription factor binding, protein metabolism as well as regulation of apoptosis (Table I). Therefore, we focus on the genes in module A in the following analysis.

Nine hub genes (top 5% genes ranked by degree) were revealed and 8 of them had been implicated in HCC. They are protein kinase C alpha (PRKCA), tyrosine 3-monooxygenase/tryptophan 5-monooxygenase activation protein zeta (YWHAZ), keratin 18 (KRT18), N-myc downstream regulated 1 (NDRG1), heat shock protein family A member 1A (HSPA1A), heat shock protein 90kDa alpha family class A member 1 (HSP90AA1), heat shock transcription factor 1 (HSF1), and inhibitor of kappa light polypeptide

Critical genes of HCC revealed by RNA-seq data analysis

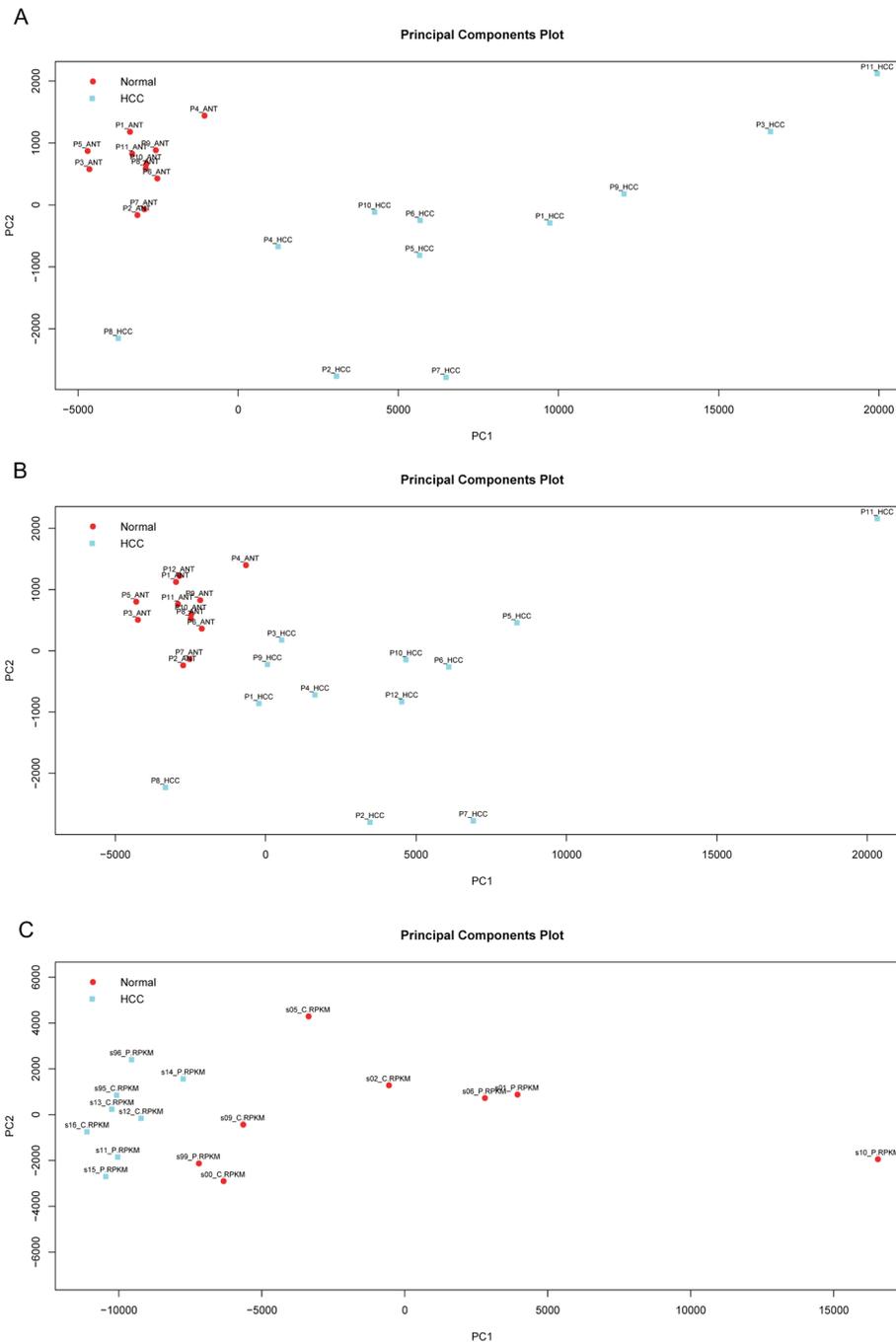


Figure 2. Principle component analysis result using DEGs from datasets. GSE69164 (A), GSE63868 (B) and GSE55857 (C). Green circles represent HCC samples and red circles represent normal samples.

gene enhancer in B-cells, kinase beta (IKGKB). Ubiquitin-conjugating enzyme E2I (UBE2I) was identified as a hub gene in module A, which has not yet been reported related to hepatocellular carcinoma. However, it has been reported associated with ovarian cancer²⁴.

Potential prognostic genes

The 54 cases with survival time obtained from TCGA were clustered into two groups using genes from module A (Figure 4 A). A significant difference was observed between the two groups in survival analysis (Figure 4 B). Univariate Cox

Table I. Top 10 (ranked by FDR) biological processes over-represented in module A.

Category	Term	FDR
GOTERM_MF_FAT	GO:0008134~transcription factor binding	1.09E-04
GOTERM_MF_FAT	GO:0019904~protein domain specific binding	1.58E-04
GOTERM_BP_FAT	GO:0043933~macromolecular complex subunit organization	4.24E-04
GOTERM_BP_FAT	GO:0043067~regulation of programmed cell death	2.77E-03
GOTERM_MF_FAT	GO:0005524~ATP binding	2.81E-03
GOTERM_BP_FAT	GO:0010941~regulation of cell death	3.00E-03
GOTERM_MF_FAT	GO:0032553~ribonucleotide binding	5.18E-03
GOTERM_BP_FAT	GO:0042981~regulation of apoptosis	6.30E-03
GOTERM_BP_FAT	GO:0070271~protein complex biogenesis	2.01E-02
GOTERM_BP_FAT	GO:0006461~protein complex assembly	2.01E-02

FDR: false discovery rate.

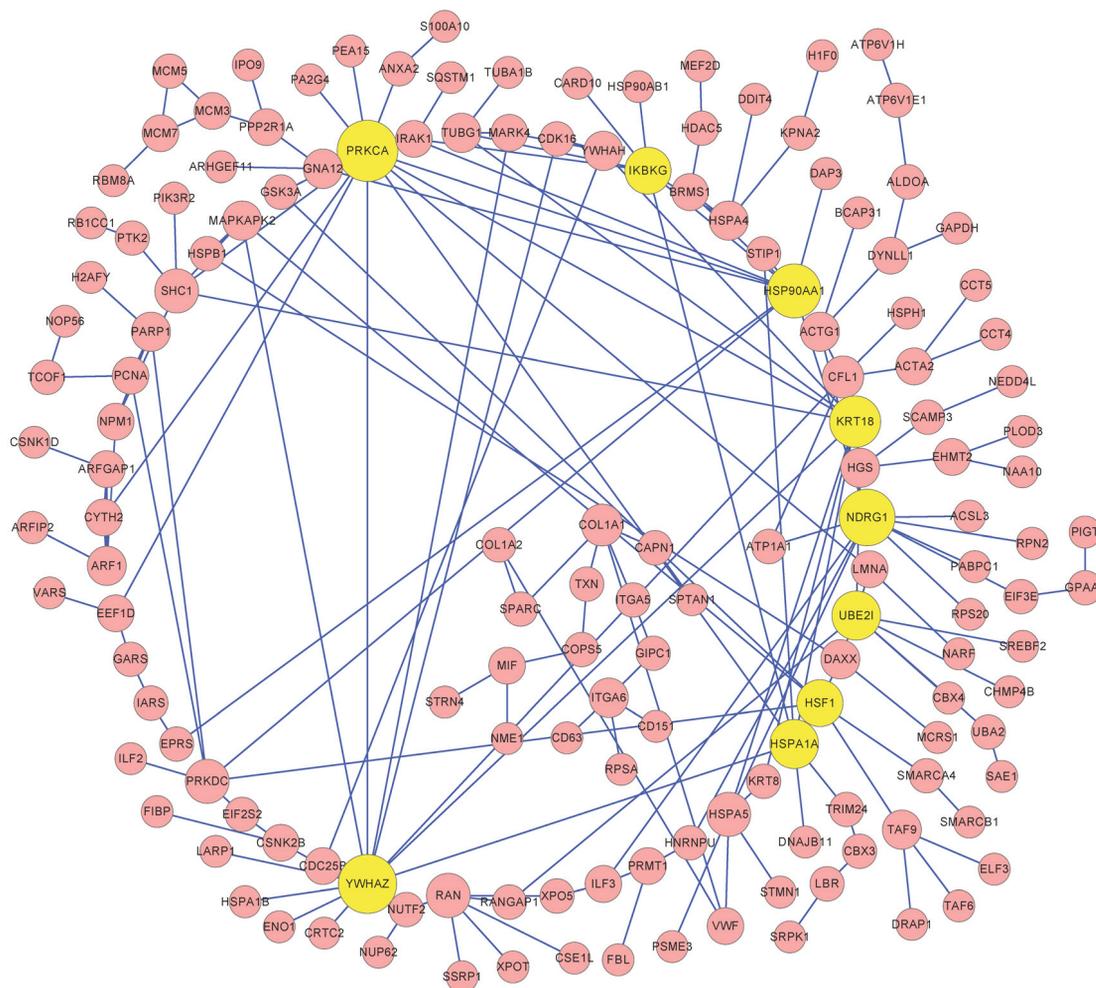


Figure 3. Module A identified from the HCC-related PPI network. Yellow circles represent hub genes.

regression analysis indicated that 72 genes were significantly associated with survival time (Table II). Top 15 genes are listed in Table II, such as nucleophosmin (NPM1), protein kinase, DNA-

activated, catalytic polypeptide (PRKDC), secreted protein, acidic, cysteine-rich (SPARC), high mobility group AT-hook 1 (HMGA1), collagen type I alpha 1 (COL1A1) and COL1A2.

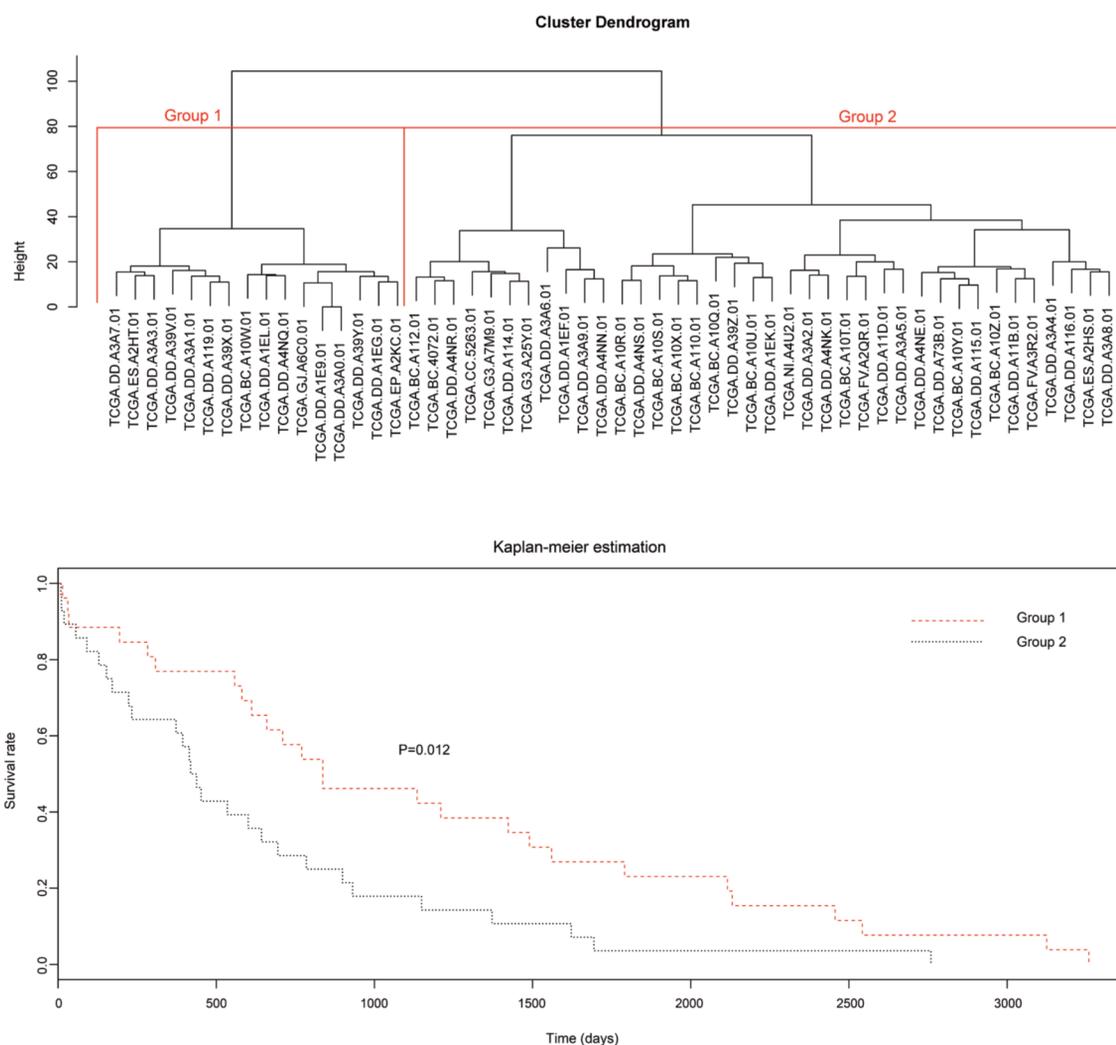


Figure 4. Clustering of the 54 HCC cases (A) and K-M analysis result of survival curve (B).

Discussion

In the present study, 2572 DEGs were identified from 3 RNA-seq datasets. A PPI network including 660 nodes and 1008 edges was constructed, from which 4 modules were obtained. Genes from module A were involved in transcription factor binding, protein metabolism as well as regulation of apoptosis, which were closely associated with cancers. Further, 9 hub genes were disclosed and 8 of them have been implicated in HCC.

PRKCA plays a role in many different cellular processes, such as cell adhesion, cell transformation, cell cycle checkpoint, and cell volume control. Reduction of PRKCA decreases cell proliferation, migration, and invasion of HCC cells²⁵. YWHAZ

Table II. Top 15 genes significantly correlated to survival time.

Gene Symbol	p-value	Coefficient
CACYBP	0.0001	0.7952
IARS	0.0002	0.8284
ACTA2	0.0002	-0.5025
PARP1	0.0003	0.7045
NPM1	0.0004	0.7824
PRKDC	0.0008	0.5632
ACSL3	0.0009	0.7448
ACACA	0.0010	0.6593
AATF	0.0017	0.8597
CBX3	0.0018	0.9965
EPRS	0.0020	0.7482
SPARC	0.0024	-0.4561
COL1A1	0.0027	-0.2546
HMGA1	0.0029	0.3609
COL1A2	0.0030	-0.2886

mediates signal transduction by binding to phosphoserine-containing proteins, which were upregulated in HepG2 with doxorubicin treatment²⁶. KRT18 is part of type I intermediate filament chain. Dysregulation of KRT18 is observed in HCC²⁷. NDRG1 is involved in stress responses, hormone responses, cell growth, and differentiation. It's necessary for p53-mediated caspase activation and apoptosis. NDRG1 promotes growth of HCC cells by directly interacting with GSK-3 β and Nur77 to prevent β -catenin degradation²⁸. It's also responsible for doxorubicin and retinoic acid resistance in hepatocellular carcinoma cells²⁸. Therefore, it's regarded as a biomarker for metastasis, recurrence and of poor prognosis in HCC¹². HSPA1A is a member of the heat shock protein 70 family. It is overexpressed in HCC²⁹ and may be related to aggressiveness and prognosis of HCC. HSP90AA1 aids in the proper folding of specific target protein. It's found to be up-regulated in HCC³⁰. HSF1 is a heat-shock transcription factor. It's considered as a key determinant of HCC development by regulating hepatic steatosis and metabolic syndrome³¹. It activates miR-135b expression, consequently enhancing HCC cell motility³². It may have prognostic value in patients receiving resection of HCC³³. IKK β is a key molecule in signaling to the transcription factor NF- κ B³⁴. Suppression of IKK- β by miR-451 inhibits HCC cell proliferation³⁵.

Survival analysis indicated that genes from module A could separate HCC cases with significantly different survival time. A total of 72 survival-related genes were unveiled. Some of them have been regarded as potential prognostic markers in HCC, like HMGA1³⁶ and COL1A1³⁷. Besides, many genes play roles in the development of HCC. NPM1 promotes degradation of activating transcription factor 5 (ATF5) and thus regulates proliferation and survival of HCC³⁸. PRKDC is the catalytic subunit of the DNA-dependent protein kinase (DNAPK). The study of Cornell et al³⁹ indicate that DNAPK is a candidate driver of hepatocarcinogenesis and tissue biomarker that predicts response to treatment and survival. The expression of SPARC correlates with tumor angiogenesis in HCC⁴⁰. COL1A2 is involved in HCC cell migration⁴¹. More researches on these genes are necessary to uncover their prognostic value.

Conclusions

Overall, network and module analysis of RNA-seq data revealed nine critical genes in

HCC. Besides, 72 potential prognostic genes were obtained. These findings could advance the understanding of the pathogenesis and provide valuable information to further investigate the prognostic markers of HCC.

Conflict of Interest

The Authors declare that they have no conflict of interests.

References

- 1) YU H, SHEN H, ZHANG Y, ZHONG F, LIU Y, QIN L, YANG P. CAV1 promotes HCC cell progression and metastasis through Wnt/ β -catenin pathway. *PLoS One* 2014; 9: e106451.
- 2) TANG J, LI L, HUANG W, SUI C, YANG Y, LIN X, HOU G, CHEN X, FU J, YUAN S, LI S, WEN W, TANG S, CAO D, WU M, CHEN L, WANG H. MiR-429 increases the metastatic capability of HCC via regulating classic Wnt pathway rather than epithelial-mesenchymal transition. *Cancer Lett* 2015; 364: 33-43.
- 3) QADIR XV, HAN C, LU D, ZHANG J, WU T. miR-185 inhibits hepatocellular carcinoma growth by targeting the DNMT1/PTEN/Akt pathway. *Am J Pathol* 2014; 184: 2355-2364.
- 4) CHEN Y, LIN C, LIU Y, JIANG Y. HMGB1 promotes HCC progression partly by downregulating p21 via ERK/c-Myc pathway and upregulating MMP-2. *Tumor Biol* 2016; 37: 4399-4408.
- 5) MA S, LEE T, ZHENG B, CHAN K, GUAN X. CD133+ HCC cancer stem cells confer chemoresistance by preferential expression of the Akt/PKB survival pathway. *Oncogene* 2008; 27: 1749-1758.
- 6) BODZIN AS, WEI Z, HURTT R, GU T, DORIA C. Gefitinib resistance in HCC mahlavu cells: upregulation of CD133 expression, activation of IGF-1R signaling pathway, and enhancement of IGF-1R nuclear translocation. *J Cell Physiol* 2012; 227: 2947-2952.
- 7) EZZELDIN M, BORREGO-DIAZ E, TAHA M, ESFANDYARI T, WISE AL, PENG W, ROUYANIAN A, ASVADI Kermani A, SOLEIMANI M, PATRAD E, LIALYTE K, WANG K, WILLIAMSON S, ABDULKARIM B, OLYAEE M, FARASSATI F. RalA signaling pathway as a therapeutic target in hepatocellular carcinoma (HCC). *Mol Oncol* 2014; 8: 1043-1053.
- 8) LEE TK, MAN K, HO JW, WANG XH, POON RT, SUN CK, NG KT, NG IO, XU R, FAN ST. Significance of Rac signaling pathway in HCC cell motility: Implication for a new therapeutic treatment. *Carcinogenesis* 2005; 26: 681-687.
- 9) DUAN X, HU J, WANG Y, GAO J, PENG D, XIA L. MicroRNA-145: a promising biomarker for hepatocellular carcinoma (HCC). *Gene* 2014; 541: 67-68.
- 10) XU B, CAI Z, ZENG Y, CHEN L, DU X, HUANG A, LIU X, LIU J. α -Methylacyl-CoA racemase (AMACR) serves as a prognostic biomarker for the early recurrence/metastasis of HCC. *J Clin Pathol* 2014; 67: 974-979.

- 11) PONTISSO P, QUARTA S, CABERLOTTO C, BENEDEUCE L, MARINO M, BERNARDINELLO E, TONO N, FASSINA G, CAVALLETTO L, GATTA A, CHEMELLO L. Progressive increase of SCCA-IgM immune complexes in cirrhotic patients is associated with development of hepatocellular carcinoma. *Int J Cancer* 2006; 119: 735-740.
- 12) CHENG J, XIE HY, XU X, WU J, WEI X, SU R, ZHANG W, LV Z, ZHENG S, ZHOU L. NDRG1 as a biomarker for metastasis, recurrence and of poor prognosis in hepatocellular carcinoma. *Cancer Lett* 2011; 310: 35-45.
- 13) ZHANG H, YE J, WENG X, LIU F, HE L, ZHOU D, LIU Y. Comparative transcriptome analysis reveals that the extracellular matrix receptor interaction contributes to the venous metastases of hepatocellular carcinoma. *Cancer Genet* 2015; 208: 482-491.
- 14) ZHANG H, WENG X, YE J, HE L, ZHOU D, LIU Y. Promoter hypermethylation of TERT is associated with hepatocellular carcinoma in the Han Chinese population. *Clin Res Hepatol Gastroenterol* 2015; 39: 600-609.
- 15) WANG Y, LI J, CHEN J, LIU L, PENG Z, DING J, DING K. From cirrhosis to hepatocellular carcinoma in HCV-infected patients: genes involved in tumor progression. *Eur Rev Med Pharmacol Sci* 2012; 16: 995-1000.
- 16) HE TL, ZHENG KL, LI G, SONG B, ZHANG YJ. Identification of typical miRNAs and target genes in hepatocellular carcinoma by DNA microarray technique. *Eur Rev Med Pharmacol Sci* 2014; 18: 108-116.
- 17) WANG SY, FENG LY, MENG ZQ. Bicluster and pathway enrichment analysis related to tumor progression of hepatocellular carcinoma. *Eur Rev Med Pharmacol Sci* 2015; 19: 1191-1197.
- 18) GRACE C, NACHEVA EP. Significance analysis of microarrays (SAM) offers clues to differences between the genomes of adult philadelphia positive ALL and the lymphoid blast transformation of CML. *Cancer Inform* 2012; 11: 173-183.
- 19) TAN H, TIAN Y, YANG H, LIU G, NIE L. A novel *Streptomyces* gene, *samR*, with different effects on differentiation of *Streptomyces ansochromogenes* and *Streptomyces coelicolor*. *Arch Microbiol* 2002; 177: 274-278.
- 20) FRANCESCHINI A, SZKLARCZYK D, FRANKILD S, KUHN M, SIMONOVIC M, ROTH A, LIN J, MINGUEZ P, BORK P, VON MERING C, JENSEN LJ. STRING v9.1: protein-protein interaction networks, with increased coverage and integration. *Nucleic Acids Res* 2013; 41: D808-815.
- 21) BANDETTINI WP, KELLMAN P, MANCINI C, BOOKER OJ, VASU S, LEUNG SW, WILSON JR, SHANBHAG SM, CHEN MY, ARAI AE. MultiContrast Delayed Enhancement (MCOE) improves detection of subendocardial myocardial infarction by late gadolinium enhancement cardiovascular magnetic resonance: a clinical validation study. *J Cardiovasc Magn Reson* 2012; 14: 83.
- 22) SHANNON P, MARKIEL A, OZIER O, BALIGA NS, WANG JT, RAMAGE D, AMIN N, SCHWIKOWSKI B, IDEKER T. Cytoscape: a software environment for integrated models of biomolecular interaction networks. *Genome Res* 2003; 13: 2498-2504.
- 23) DENNIS G, JR., SHERMAN BT, HOSACK DA, YANG J, GAO W, LANE HC, LEMPICKI RA. DAVID: Database for Annotation, Visualization, and Integrated Discovery. *Genome Biol* 2003; 4: P3.
- 24) GUO Y, FU P, ZHU H, REED E, REMICK SC, PETROS W, MUELLER MD, YU JJ. Correlations among ERCC1, XPB, UBE2I, EGF, TAL2 and ILF3 revealed by gene signatures of histological subtypes of patients with epithelial ovarian cancer. *Oncol Rep* 2012; 27: 286-292.
- 25) WU TT, HSIEH YH, HSIEH YS, LIU JY. Reduction of PKCa decreases cell proliferation, migration, and invasion of human malignant hepatocellular carcinoma. *J Cell Biochem* 2008; 103: 9-20.
- 26) WANG J, CHAN JY, FONG CC, TZANG CH, FUNG KP, YANG M. Transcriptional analysis of doxorubicin-induced cytotoxicity and resistance in human hepatocellular carcinoma cell lines. *Liver Int* 2009; 29: 1338-1347.
- 27) LEE CF, LING ZQ, ZHAO T, FANG SH, CHANG WC, LEE SC, LEE KR. Genomic-wide analysis of lymphatic metastasis-associated genes in human hepatocellular carcinoma. *World J Gastroenterol* 2009; 15: 356-365.
- 28) LU WJ, CHUA MS, WEI W, SO SK. NDRG1 promotes growth of hepatocellular carcinoma cells by directly interacting with GSK-3 β and Nur77 to prevent β -catenin degradation. *Oncotarget* 2015; 6: 29847-29859.
- 29) YANG Z, ZHUANG L, SZATMARY P, WEN L, SUN H, LU Y, XU Q, CHEN X. Upregulation of heat shock proteins (HSPA12A, HSP90B1, HSPA4, HSPA5 and HSPA6) in tumour tissues is associated with poor outcomes from HBV-related early-stage hepatocellular carcinoma. *Int J Med Sci* 2015; 12: 256-263.
- 30) NEGRONI L, TAOUI S, ARMA D, PALLARES-LUPON N, LEONG K, BEAUSANG LA, LATTERICH M, BOSSÉ R, BALABAUD C, SCHMITTER JM, BIOULAC-SAGE P, ZUCMAN-ROSSI J, ROSENBAUM J, CHEVET E. Integrative quantitative proteomics unveils proteostasis imbalance in human hepatocellular carcinoma developed on nonfibrotic livers. *Mol Cell Proteomics* 2014; 13: 3473-3483.
- 31) JIN X, MOSKOPHIDIS D, MIVECHI N. Heat shock transcription factor 1 is a key determinant of HCC development by regulating hepatic steatosis and metabolic syndrome. *Cell Metab* 2011; 14: 91-103.
- 32) LI Y, XU D, BAO C, ZHANG Y, CHEN D, ZHAO F, DING J, LIANG L, WANG Q, LIU L, LI J, YAO M, HUANG S, HE X. MicroRNA-135b, a HSF1 target, promotes tumor invasion and metastasis by regulating RECK and EVI5 in hepatocellular carcinoma. *Oncotarget* 2015; 6: 2421-2433.
- 33) ZHANG JB, GUO K, SUN HC, ZHU XD, ZHANG B, LIN ZH, ZHANG BH, LIU YK, REN ZG, FAN J. Prognostic value of peritumoral heat-shock factor-1 in patients receiving resection of hepatocellular carcinoma. *Br J Cancer* 2013; 109: 1648-1656.

- 34) SCHMID JA, BIRBACH A. IkappaB kinase beta (IKKbeta/IKK2/IKBKB)—a key molecule in signaling to the transcription factor NF-kappaB. *Cytokine Growth Factor Rev* 2008; 19: 157-165.
- 35) LI HP1, ZENG XC, ZHANG B, LONG JT, ZHOU B, TAN GS, ZENG WX, CHEN W, YANG JY. miR-451 inhibits cell proliferation in human hepatocellular carcinoma through direct suppression of IKK- β . *Carcinogenesis* 2013; 34: 2443-2451.
- 36) CHANG ZG, YANG LY, WANG W, PENG JX, HUANG GW, TAO YM, DING X. Determination of high mobility group A1 (HMGA1) expression in hepatocellular carcinoma: a potential prognostic marker. *Dig Dis Sci* 2005; 50: 1764-1770.
- 37) HAYASHI M, NOMOTO S, HISHIDA M, INOKAWA Y, KANDA M, OKAMURA Y, NISHIKAWA Y, TANAKA C, KOBAYASHI D, YAMADA S, NAKAYAMA G, FUJII T, SUGIMOTO H, KOIKE M, FUJIWARA M, TAKEDA S, KODERA Y. Identification of the collagen type 1 alpha 1 gene (COL1A1) as a candidate survival-related factor associated with hepatocellular carcinoma. *BMC Cancer* 2014; 14: 108.
- 38) LIU X1, LIU D, QIAN D, DAI J, AN Y, JIANG S, STANLEY B, YANG J, WANG B, LIU X, LIU DX. Nucleophosmin (NPM1/B23) interacts with activating transcription factor 5 (ATF5) protein and promotes proteasome- and caspase-dependent ATF5 degradation in hepatocellular carcinoma cells. *J Biol Chem* 2012; 287: 19599-19609.
- 39) CORNELL L, MUNCK JM, ALSINET C, VILLANUEVA A, OGLE L, WILLOUGHBY CE, TELEVANTOU D, THOMAS HD, JACKSON J, BURT AD, NEWELL D, ROSE J, MANAS DM, SHAPIRO GI, CURTIN NJ, REEVES HL. DNA-PK-A candidate driver of hepatocarcinogenesis and tissue biomarker that predicts response to treatment and survival. *Clin Cancer Res* 2014; 21: 925-933.
- 40) LAU CP, POON RT, CHEUNG ST, YU WC, FAN ST. SPARC and Hevin expression correlate with tumour angiogenesis in hepatocellular carcinoma. *J Pathol* 2006; 210: 459-468.
- 41) JI J, ZHAO L, BUDHU A, FORGUES M, JIA HL, QIN LX, YE OH, YU J, SHI X, TANG ZY, WANG XW. Let-7g targets collagen type I α 2 and inhibits cell migration in hepatocellular carcinoma. *J Hepatol* 2010; 52: 690-697.