

# MiR-524 inhibits cell proliferation and induces cell apoptosis in thyroid cancer via targeting SPAG9

Z. ZHEN<sup>1</sup>, F. DONG<sup>2</sup>, H. SHEN<sup>1</sup>, Q.-G. WANG<sup>1</sup>, L. YANG<sup>3</sup>, J. HU<sup>4</sup>

<sup>1</sup>Department of Otolaryngology, Head and Neck Surgery, Peking University First Hospital, Beijing, China

<sup>2</sup>Department of Breast and Thyroid Surgery, Union Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan, China

<sup>3</sup>Department of Emergency Medicine, Tongji Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan, China

<sup>4</sup>Department of Otolaryngology-Head and Neck Surgery, Tongji Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan, China

**Abstract. – OBJECTIVE:** To investigate the effect of miR-524 on the proliferation of thyroid cancer and its underlying mechanism.

**PATIENTS AND METHODS:** MiR-524 expression levels in thyroid cancer samples and para-cancer tissues were tested by quantitative Real-time polymerase chain reaction (qRT-PCR). Cells proliferative ability and apoptosis were evaluated through methyl thiazolyl tetrazolium (MTT) and apoptosis assays, respectively. Luciferase reporter assay was used to confirm regulatory mechanism.

**RESULTS:** MiR-524 expression was reduced in thyroid cancer specimen. Over-regulated miR-524 expression inhibited the proliferative ability and enhanced cell apoptosis in thyroid cancer cells. SPAG9 was target of miR-524, and was reverse regulated by miR-524.

**CONCLUSIONS:** MiR-524 inhibits thyroid cancer cell proliferation and induces cell apoptosis via targeting SPAG9.

**Key Words:** miR-524, thyroid cancer, Proliferation, Apoptosis.

## Introduction

Thyroid cancer is the most common malignant tumor of endocrine origin, accounting for 94.5% of all endocrine tumors and 2.6% of all malignant tumors with the highest incidence of head and neck malignancies. At the beginning of the last century, the incidence of thyroid cancer exhibited a gradual upward trend<sup>1,2</sup>. Papillary thyroid cancer (PTC) is the most common type of thyroid cancer, accounting for about 90% of all pathological types<sup>3</sup>. At present, the treatment of papillary thyroid carcinoma

includes surgical treatment, thyroid hormone suppression therapy, isotope iodine 131 treatment and adjuvant radiotherapy. With effective and reasonable treatment, papillary thyroid carcinoma generally has a good prognosis; the 5-year survival rate is about 90% and 10-year survival rate is also above 80%<sup>4</sup>. However, some papillary cancers have the tendency of dedifferentiation and eventually develop into poorly differentiated thyroid cancer or undifferentiated carcinoma, which results in the decline of survival rate and quality of life<sup>5</sup>. Therefore, it is crucial to look for differentially expressed genes, predictive biomarkers and target molecules for intervention therapy, which could further increase the cure rate and survival rate of patients.

Previous researchers<sup>6-11</sup> have found that miRNAs are implicated in cell proliferation, apoptosis, invasion and angiogenesis in the different diseases. In thyroid cancer, miRNAs are differentially expressed in the serum of patients<sup>12</sup>. MiR-146a polymorphism is significantly associated with PTC risk<sup>13</sup>.

Upregulation of microRNA-524-5p enhances the cisplatin sensitivity of gastric cancer cells by modulating proliferation and metastasis via targeting SOX9<sup>14</sup>. miRNA-106b-mediated down-regulation of Clorf 24 expression induces apoptosis and suppresses invasion of thyroid cancer<sup>15</sup>. MiR-524-5p may function as a novel tumor suppressor gene, and suppresses the growth and invasive abilities of gastric cancer cells<sup>16</sup>. However, there are no relevant reports on the mechanism of action of miR-524 in PTC. The primary purpose of this study was to investigate the effect of miR-524 on the proliferation of thyroid cancer and its underlying mechanism.

## Patients and Methods

### Clinical Specimens

We collected fresh cancer tissues and para-cancer tissues from thyroid cancer patients in our hospital. All specimens were pathological confirmed and quickly placed in inactivated RNase cryopreservation tube in liquid nitrogen cryopreservation during 15 minutes, and then into 80°C refrigerator to be reserved. All patients did not receive the treatment of preoperative iodine 131 and thyroid hormone suppression therapy. Patients with other malignant tumors, systemic serious infection, and other serious systemic diseases were excluded. This study was approved by the Ethics Committee of Tongji Hospital, Tongji Medical College, Huazhong University of Science and Technology. Signed written informed consents were obtained from all participants before the study.

### Cell Culture

Thyroid cancer cell lines WRO [DMEM (Dulbecco's Modified Eagle Medium)] and TPC1 [RPMI (Roswell Park Memorial Institute)] medium were supplemented with 10% fetal bovine serum (FBS; Life Technologies, Grand Island, NY, USA). Both of them were maintained in a 5% CO<sub>2</sub> and 37°C humidified incubator.

### Cell Transfection

MiR-524 mimics and miR-524 mimics NC (normal control) were synthesized and purified by Shanghai Jima Co., Ltd. (Shanghai, China). Cells were seeded into 6-well plate at a density of  $2.5 \times 10^5$ /mL. After 48 h, miR-524 mimics and miR-524 mimics NC were transfected into WRO and TPC1 cell lines with Lipofectamine 2000 (Invitrogen, Carlsbad, CA, USA) according to the manufacturer's protocol. The mimics concentration was 50 nM.

### Quantitative Real-Time Polymerase Chain Reaction (qRT-PCR)

Total RNA was extracted from the cell lines and tissues using the TRIzol reagent kit (Life Technologies, Grand Island, NY, USA). Then, all RNA was synthesized into complementary DNA (cDNA) via using MuLV reverse transcriptase (Fisher Scientific, Pittsburgh, PA, USA). Real-time PCR was conducted using SYBR Green mix (TaKaRa, Dalian, China). The detection system condition was

94°C for 3 min, followed by 35 cycles of 94°C for 30 s and 60°C for 30 s. MiR-524 expression was normalized to U6 RNA. Primer sequences used in this study were as follows: miR-524, F: 5'-GCTGTGACCCTACAAAGGGA-3', R: 5'-AGCATCAACTTCAACGCTG-3'; SPAG9, F: 5'-TCCTGAGCTGGATATGCAAGA-3', R: 5'-GCCTGAGCCAGCTCAAGG-3'; U6, F: 5'-CTCGCTTCGGCAGCAACA-3', R: 5'-GCTTCACGAATTTGG-3'.

### Methyl Thiazolyl Tetrazolium (MTT) Assay

Cells ( $1 \times 10^4$  cells/well) were seeded into a 96-well plate and incubated for 2, 3, 4, 5, and 6 days. MTT reagent (20 µL) was added to each well for 4 h incubation. Then, dimethyl sulfoxide (DMSO) (150 µL) was added to each well. The absorbance at 540 nm wavelength was detected.

### Cell Apoptosis Detection

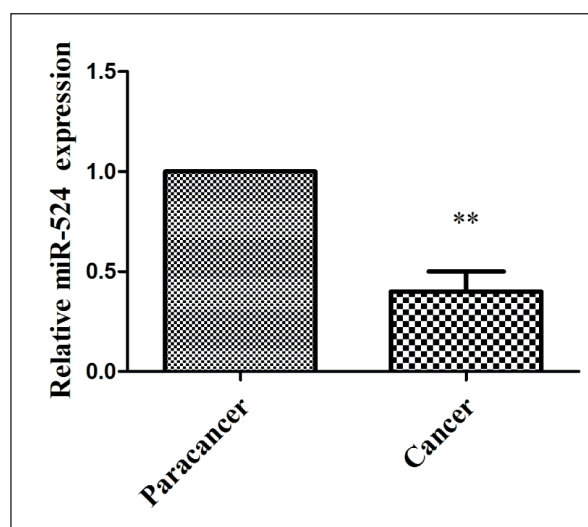
Cells were re-suspended in the binding buffer (200 µL) and incubated with FITC Annexin V (5 µL) and propidium iodide solution (1 µL). Finally, cell apoptosis was analyzed by Calibur Flow Cytometer (Becton, Dickinson and Company, Franklin Lakes, NJ, USA).

### Luciferase Reporter Assay

With PCR Amplification kit (TaKaRa, Dalian, China), SPAG9 3'UTR including the predicted binding site was amplified, and then cloned into the psiCHECK-2 reporter vector (Promega, Madison, WI, USA). By using Lipofectamine 2000 (Invitrogen, Carlsbad, CA, USA), psiCHECK-2-SPAG9-wild (200 ng) or psiCHECK-2-SPAG9-mutant (200 ng) and miR-524 mimics (100 nmol/L) were co-transfected into WRO cells. The reporter activity was examined by a Dual Luciferase Reporter Assay kit (Promega, Madison, WI, USA).

### Statistical Analysis

The results were carried out in triplicate. Graphpad Prism v5.0 (GraphPad Software, La Jolla, CA, USA) was used for analyses. The comparisons were examined using the Student's *t*-test. A *p* value of <0.05 was regarded as statistically significant.



**Figure 1.** The relative miR-524 expression was detected by qRT-PCR between paracancer tissues and cancer tissues (\*\* $p < 0.01$ ).

## Results

### *MiR-524 Was Down-Regulation In Thyroid Cancer Samples*

To investigate the unknown functions of miR-524 in thyroid cancer, we firstly detected the expression of miR-524 in cancer tissues and para-cancer tissues *via* using qRT-PCR method. The analysis exhibited that lower expression of miR-524 was found in cancer tissues compared with the para-cancer tissues (Figure 1). This finding demonstrated that dysregulation of miR-524 is implicated in the progression of thyroid cancer.

### *MiR-524 Overexpression Could Reversely Regulate Cell Proliferative Ability*

Furthermore, to explore the influence of miR-524 down-regulation on cell proliferative ability, MTT method was used. Interestingly, the result of MTT (Figure 2A and 2B) showed that the proliferative ability of cells transfected with miR-524 mimics was decreased as compared to the control group. This suggested that miR-524 overexpression could suppress cell proliferative ability.

### *Up-regulation of miR-524 Could Promote Cell Apoptosis*

We further investigated the changes of cell apoptosis responding to up-regulation of miR-524 through an apoptosis assay. The results of apoptosis assay showed that more cell apoptosis was detected in over-expressing miR-524 group compared with the control group (Figure

2C and 2D). These results indicated that up-regulation of miR-524 could promote cell apoptosis.

### *SPAG9 Was a Target Gene of miR-524*

Various studies<sup>6-9</sup> have reported that miRNAs influences the different cancer development by modulating the target genes. To find the target gene of miR-524, we used three miRNA bioinformatics predictive software (miRDB, RNA22 and TargetScan). The intersection of the three bioinformatics predictive software were assessed (Figure 3A). A total of 10 potential target genes of miR-524 were shown in Figure 3B. Additionally, according to the research of Wang et al<sup>17</sup>, SPAG9 was highly expressed in thyroid cancer, and knockdown of SPAG9 expression in cell significantly reduced cellular growth and colony formation. Therefore, we focused on SPAG9.

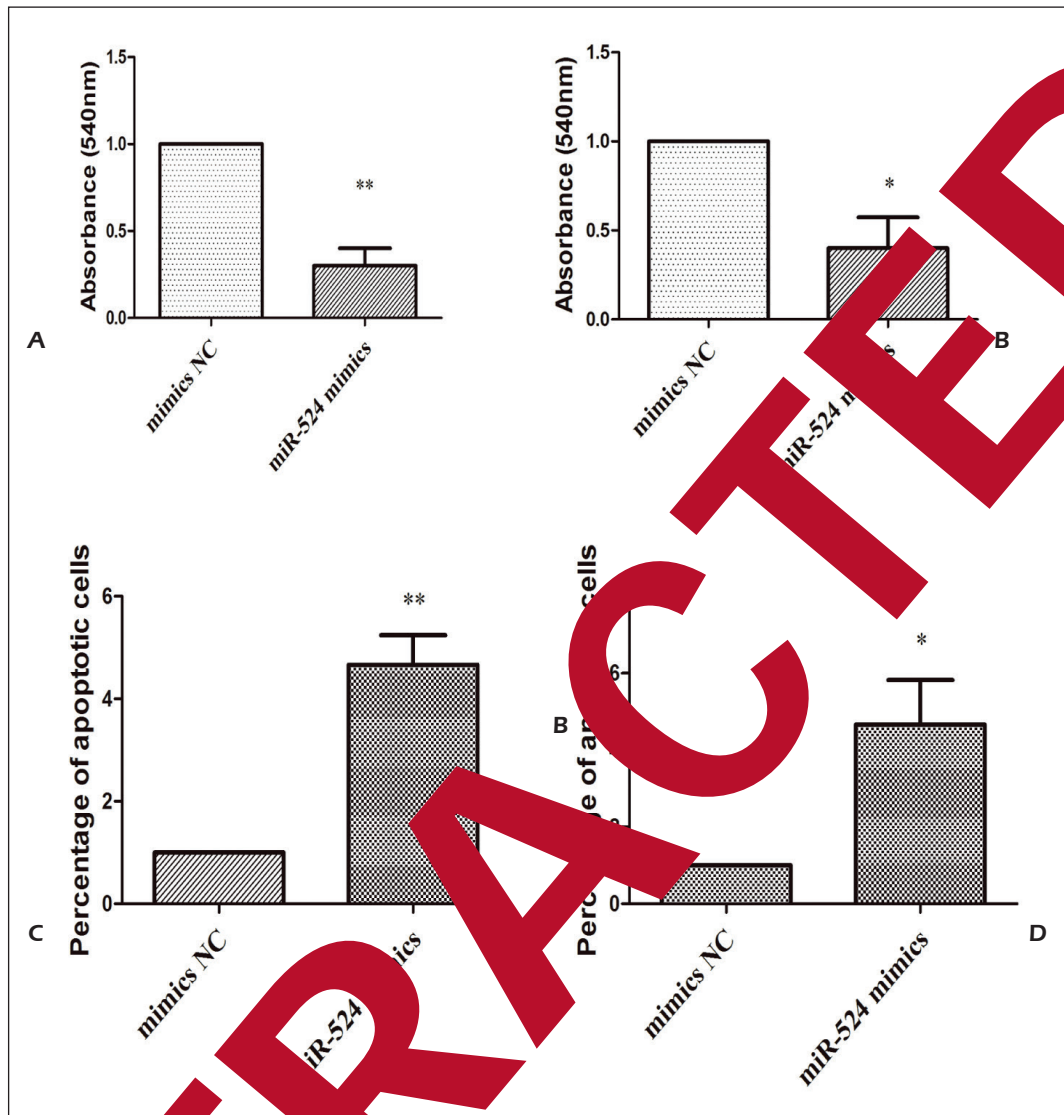
### *MiR-524 Could Reversely Regulate SPAG9 Through Bounding to SPAG9 3'UTR*

To explore the binding site of miR-524 to SPAG9 3'UTR, we showed in Figure 4A. To confirm it, we planned a luciferase reporter assay. The result displayed that the luciferase activity was significantly less in psiCHECK-2-SPAG9-Wild-mimics (Figure 4B), which demonstrated that miR-524 could bound to SPAG9 3'UTR.

Furthermore, we used qRT-PCR method to detect the regulatory correlation between miR-524 and SPAG9. This test displayed that declined SPAG9 was expressed when miR-524 was up-regulation (Figure 4C and 4D). All these results indicated that miR-524 could reversely regulate SPAG9 through bound to SPAG9 3'UTR.

## Discussion

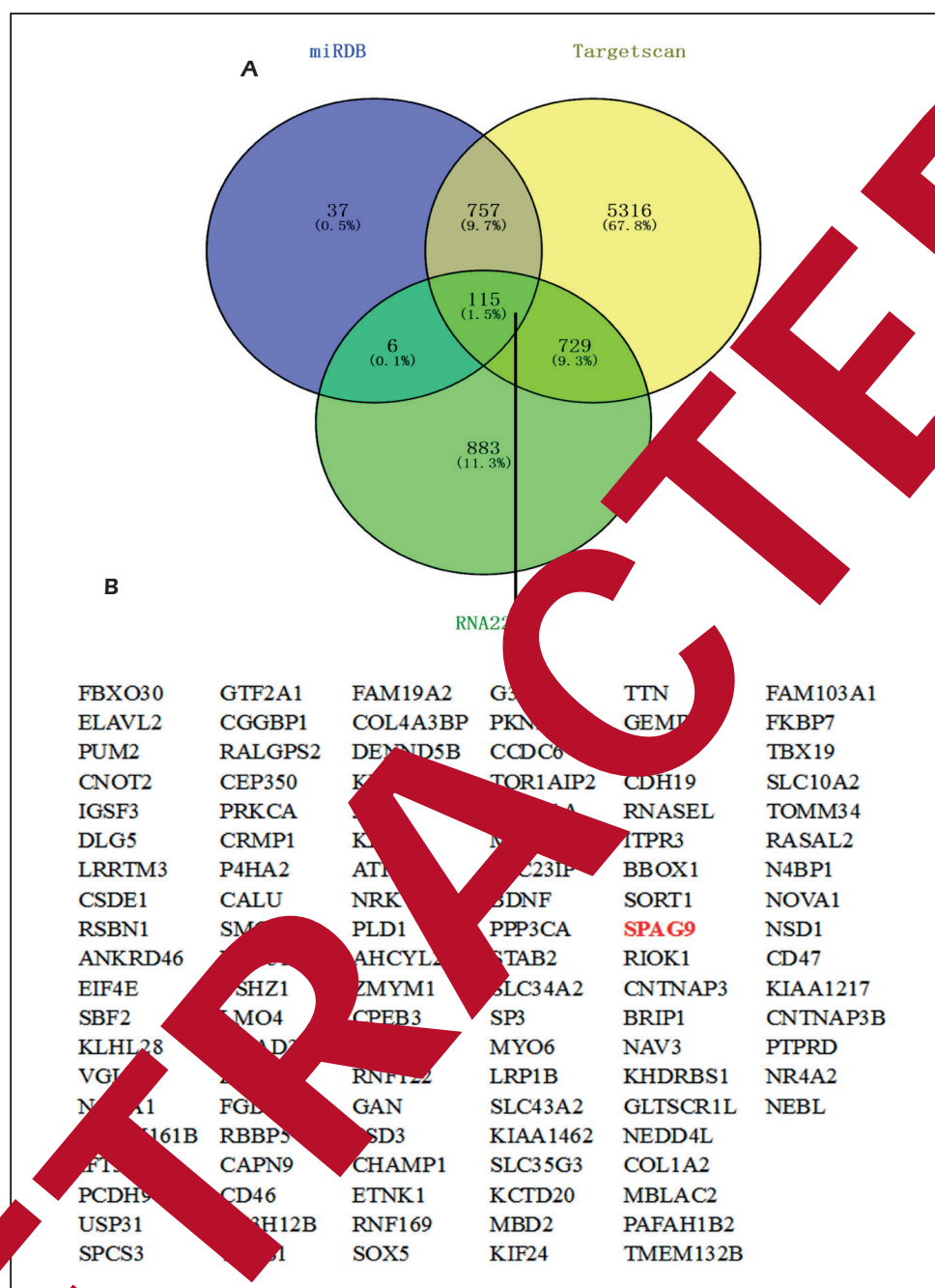
MiRNAs, as small single-chain non-coding RNAs with 19-22 nt in length, can regulate the expression of specific mRNAs at the transcriptional level, ultimately resulting in functional changes in cells or tissue. Evidence had indicated that abnormal up-regulation or down-regulation of miRNAs is associated with the development, invasion, and metastasis of various tumors<sup>18,19</sup>. Our work found that there is a significant difference in the expression of miR-524 between thyroid cancer and para-cancerous tissues. Through the previous functional tests, we discovered that miR-524 acts as a tumor suppressor gene in thyroid cancer. Ac-



**Figure 2.** MiR-524 mimics expression inhibited cell proliferation and promoted cell apoptosis. MTT assessment of up-regulated miR-524 on cell proliferation, absorbance (540nm) was obtained in WRO (A) and TPC1 (B); cell apoptosis assay was performed in WRO (C) and TPC1 (D) (\* $p < 0.05$ , \*\* $p < 0.01$ ).

According to the mechanism of miRNA on downstream target genes, it can inhibit the translational expression of target gene mRNA and reduce the protein level through complete or incomplete complementation with the target gene. Therefore, target genes of miR-524 should meet the following conditions: firstly, the target genes are oncogenes, which are involved in promoting the development and progression of tumors. Secondly, there is complementation of miRNAs with their target sites. Thirdly, there are conserved types of miRNA target sites between different species. Fourthly, there is the thermal stability between miRNA and mRNA double-stranded. Fifthly, the

miRNA target site should not have complex secondary structure. We chose three miRNAs bioinformatics predictive software (miRDB, RNA22, and TargetScan) together to predict the target genes of miR-524 according to the above conditions. The intersection of the three bioinformatics predictive software was assessed. Then, to verify whether SPAG9 was a downstream gene of miR-524, we detected SPAG9 mRNA expression via qRT-PCR. We found that miR-524 could up-regulate the expression of SPAG9. Additionally, miR-524 was further verified to act specifically on SPAG9 3'UTR by dual luciferase reporter gene system.

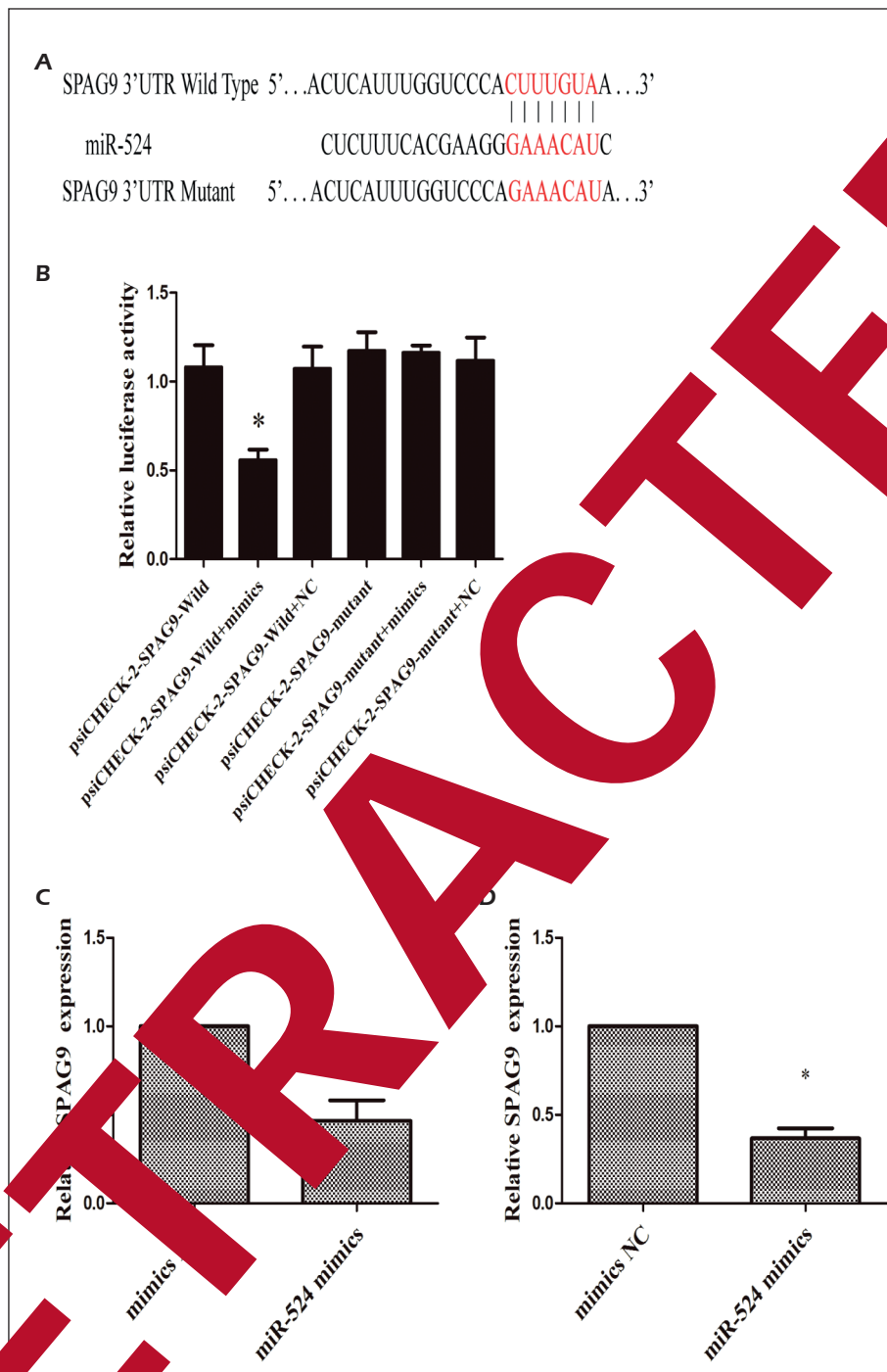


**Figure 1.** SPAG9 was a target gene of miR-524. **A**, The intersection of the three bioinformatics predictive software (miRDB, RNA22, and TargetScan) was assessed; **B**, 115 potential target genes of miR-524 were shown.

Previous studies have found that SPAG9 expression levels are overexpressed in prostate cancer<sup>21</sup>, lung cancer<sup>22</sup>, hepatocellular cancer<sup>23,24</sup>, leiomyosarcoma<sup>25,26</sup>, breast cancer<sup>27</sup>, and endometrial cancer<sup>28</sup>. Abnormal expression of SPAG9 contributes to the molecular biology processes.

In sum up, we first found the lower expression of miR-524 in thyroid cancer tissues, and the func-

tion of miR-524 tumor as suppressor gene through *in vitro* functional experiments. In addition, we further verified that SPAG9 is a downstream gene of miR-524. Further studies are needed to explore the role of SPAG9 downstream genes in thyroid cancer, so as to discover a complete regulatory pathway, and then improve the regulatory network in thyroid cancer.



**Figure 4.** miR-524 inhibited SPAG9 expression via targeting 3'UTR of SPAG9. **A**, The potential binding site was displayed (Wild type and mutant). **B**, The luciferase activity was assessed; qRT-PCR analysis showed the relative SPAG9 expression in WT (psiCHECK-2-SPAG9-Wild) and TPC1 (psiCHECK-2-SPAG9-mutant) cells with or without miR-524 mimics (\* $p < 0.05$ ).

## Conclusions

We first identified that miR-524 represses thyroid cancer cell proliferation and induces cell apoptosis *via* targeting SPAG9, which may pro-

vide a new diagnostic criterion and potential therapeutic target in treatment for PTC.

## Conflict of interest

The authors declared no conflict of interest.

## References

- 1) DAVIES L, WELCH HG. Increasing incidence of thyroid cancer in the United States, 1973-2002. *JAMA* 2006; 295: 2164-2167.
- 2) WANG Y, WANG W. Increasing incidence of thyroid cancer in Shanghai, China, 1983-2007. *Asia Pac J Public Health* 2015; 27: P223-P229.
- 3) HUNDAHL SA, FLEMING ID, FREMGEN AM, MENCK HR. A national cancer database report on 53,856 cases of thyroid carcinoma treated in the U.S., 1985-1995 [see comments]. *Cancer* 1998; 83: 2638-2648.
- 4) ALBORES-SAAVEDRA J, HENSON DE, GLAZER E, SCHWARTZ AM. Changing patterns in the incidence and survival of thyroid cancer with follicular phenotype--papillary, follicular, and anaplastic: a morphological and epidemiological study. *Endocr Pathol* 2007; 18: 1-7.
- 5) WYNFORD-THOMAS D. Origin and progression of thyroid epithelial tumours: Cellular and molecular mechanisms. *Horm Res* 1997; 47: 145-157.
- 6) ZOU YT, GAO JY, WANG HL, WANG Y, WANG H, LI PL. Downregulation of microRNA-630 inhibits cell proliferation and invasion and enhances chemosensitivity in human ovarian carcinoma. *Genet Mol Res* 2015; 14: 8766-8777.
- 7) SUN C, LI J. Expression of MiRNA-137 in oral squamous cell carcinoma and its clinical significance. *J BUON* 2018; 23: 167-172.
- 8) XIE L, ZHANG Z, TAN Z, HE R, ZENG X, XIE Y, ZHANG G, TANG H, HE X. MicroRNA-124 inhibits proliferation and induces apoptosis by directly repressing EZH2 in gastric cancer. *Mol Cell Biochem* 2015; 392: 153-159.
- 9) PAN Y, LIANG H, CHEN W, ZHANG Y, ZHANG N, WANG F, ZHANG S, LIU Y, ZHAO C, YANG J, ZHANG J, ZHANG CY, GU H, ZEN K, CHEN Y. MicroRNA-100b and microRNA-200c promote colorectal cancer cell proliferation via targeting and reversing the expression of cysteine-rich protein with two zinc fingers. *PLoS One* 2015; 12: 276-289.
- 10) MAO G, LIU Y, LIU X, LIU Y, FAN Y, LIU L, LIU X, WANG N. Tumor-derived microRNA-100b promotes angiogenesis in small cell lung cancer. *Angiogenesis* 2015; 18: 381-382.
- 11) LIU F, LI Y, LIU G. MicroRNA-200c exacerbates the ischemia/reperfusion injury of heart through targeting the glutaminase (GLS)-mediated glutamate metabolism. *Eur Rev Med Pharmacol Sci* 2017; 21: 3287-3289.
- 12) CHEN M, HART RD, DUNGLAS S, MAKKI FM, PINTO D, BUTLER L, SULLOCK M, HEBBY MH, TRITES JR, TAYLOR SM, SINGH M. miRNA profiling to distinguish papillary thyroid cancer from benign thyroid masses. *J Otolaryngol Head Neck Surg* 2015; 44: 33.
- 13) WANG G, ZHANG R, XU J, GUO Y. Association between microRNA polymorphisms and papillary thyroid cancer susceptibility. *Int J Clin Exp Pathol* 2015; 8: 13450-13457.
- 14) YANG J, XUE X, HONG H, QIN M, ZHOU J, SUN Q, ZHANG H, GAO L. Upregulation of microRNA-524-5p enhances the cisplatin sensitivity of gastric cancer cells by modulating proliferation and metastasis via targeting SOX9. *Oncotarget* 2017; 8: 574-582.
- 15) CARVALHEIRA G, NOZIMA BH, CERUTTI JM, MOURA-NA-106b-mediated down-regulation of miR-106b expression induces apoptosis and suppresses invasion of thyroid cancer. *Oncotarget* 2015; 6: 28357-28370.
- 16) LIU GH, LIU YH, YANG Z, ZHU AL, LIU GL. MicroRNA-524-5p suppresses the growth and invasive abilities of gastric cancer cells. *Oncotarget* 2016; 11: 1926-1932.
- 17) GARG M, KANOJIA D, GUPTA S, GUPTA S, GUPTA A. Sperm-associated antigen 9: a novel diagnostic marker for thyroid cancer. *J Clin Endocrinol Metab* 2009; 99: 4614-4618.
- 18) DAVIDSON B, WANG CG, REEDS J. The clinical and diagnostic role of microRNAs in ovarian carcinoma. *Gynecol Oncol* 2014; 133: 648-652.
- 19) FURUTA T, KOBAYASHI T, TANAKA S, ARII S, YAMOTO I, INAZAWA J. MiR-124 and miR-103 are epigenetically silenced tumor-suppressive microRNAs in hepatocellular carcinoma. *Carcinogenesis* 2010; 31: 766-776.
- 20) LIU Y, WANG Y, NIU H, WU L, ZHANG Y, ZHANG Y, BAI X, HE P. SPAG9 is overexpressed in human prostate cancer and promotes cancer cell proliferation. *Tumour Biol* 2014; 35: 6949-6954.
- 21) ZHANG L, ZHANG F, LU Z, WANG J, HAN X, BAI J, LIU Q, XI Y, WANG J. SPAG9 expression is increased in human prostate cancer and promotes cell motility, invasion and angiogenesis in vitro. *Oncol Rep* 2014; 32: 2533-2540.
- 22) ZHANG ZF, WANG ZN, ZHAO TT, XU YY, WU JH, LIU XY, ZHANG Y, YOU Y, XU HM. Overexpression of SPAG9 in human gastric cancer is correlated with poor prognosis. *Virchows Arch* 2015; 467: 525-533.
- 23) XIE C, FU L, LIU N, LI Q. Overexpression of SPAG9 correlates with poor prognosis and tumor progression in hepatocellular carcinoma. *Tumour Biol* 2014; 35: 7685-7691.
- 24) YAN Q, LOU G, QIAN Y, QIN B, XU X, WANG Y, LIU Y, DONG X. SPAG9 is involved in hepatocarcinoma cell migration and invasion via modulation of ELK1 expression. *Onco Targets Ther* 2016; 9: 1067-1075.
- 25) XIAO C, FU L, YAN C, SHOU F, LIU Q, LI L, CUI S, DUAN J, JIN G, CHEN J, BIAN Y, WANG X, WANG H. SPAG9 is overexpressed in osteosarcoma, and regulates cell proliferation and invasion through regulation of JunD. *Oncol Lett* 2016; 12: 2674-2679.
- 26) YANG X, ZHOU W, LIU S. SPAG9 controls the cell motility, invasion and angiogenesis of human osteosarcoma cells. *Exp Ther Med* 2016; 11: 637-644.
- 27) JAGADISH N, GUPTA N, AGARWAL S, PARASHAR D, SHARMA A, FATIMA R, TOPNO AP, KUMAR V, SURI A. Sperm-associated antigen 9 (SPAG9) promotes the survival and tumor growth of triple-negative breast cancer cells. *Tumour Biol* 2016; 37: 13101-13110.
- 28) ZHANG L, YAN L, CAO M, ZHANG H, LI C, BAI Y, YU P, LI M, ZHAO X. SPAG9 promotes endometrial carcinoma cell invasion through regulation of genes related to the epithelial-mesenchymal transition. *Eur J Gynaecol Oncol* 2016; 37: 312-319.