

# CircRNA EPB41L2 inhibits tumorigenicity of lung adenocarcinoma through regulating CDH4 by miR-211-5p

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**Abstract.** – **OBJECTIVE:** Circular RNAs (circRNAs) can make contributions to cell proliferation, migration and invasion in lung adenocarcinoma (LUAD). Circ\_Band 4.1-like protein 2 (circ\_EP41L2) was found to have anti-tumor roles in lung cancer. Herein, we investigated the roles of circ\_EP41L2 in LUAD tumorigenicity.

**PATIENTS AND METHODS:** The expression of circ\_EP41L2, microRNA (miR)-211-5p, and Cadherin-4 (CDH4) was measured using quantitative real-time polymerase chain reaction. Western blot was used to detect the levels of CDH4, Ki67, and E-cadherin. Cell proliferation, migration and invasion were examined using 3-(4,5)-dimethylthiazol(-z-y1)-3,5-diphenyl tetrazolium bromide (MTT) assay and Transwell assay, respectively. The interaction between miR-211-5p and circ\_EP41L2 or CDH4 was confirmed by Dual-Luciferase reporter assay. *In vivo* experiments were conducted using the murine xenograft model.

**RESULTS:** Circ\_EP41L2 and CDH4 were down-regulated in LUAD tissues and cell lines. Overexpressed circ\_EP41L2 acted as a tumor suppressor in the LUAD cell proliferation, invasion and migration *in vitro*. MiR-211-5p directly bound to circRNA\_EP41L2 and CDH4 3'-UTR and circ\_EP41L2 indirectly promoted CDH4 expression by binding to miR-211-5p in LUAD cells. Furthermore, rescue assay showed circ\_EP41L2 played a protective role by repressing proliferation, migration and invasion through regulating CDH4 and miR-211-5p in LUAD cells. Besides, *in vivo* experiments indicated circ\_EP41L2 overexpression also inhibited tumor growth through regulating miR-211-5p and CDH4.

**CONCLUSIONS:** Circ\_EP41L2 functioned as a tumor suppressor to inhibit the proliferation, migration and invasion through regulating CDH4 by miR-211-5p in LUAD cells, suggesting a new understanding on the tumorigenesis of LUAD and providing potential molecular therapeutic targets.

**Keywords:** circ\_EP41L2, CDH4, MiR-211-5p, LUAD, Tumorigenicity

**Introduction**

Lung cancer has been regarded as the leading cause of cancer-associated mortality worldwide, with the highest incidence and mortality rates among all cancers<sup>1</sup>. Lung adenocarcinoma (LUAD) is the main histological type of non-small cell lung cancer (NSCLC), accounting for around up to 50% of lung cancer cases<sup>2</sup>. Currently, surgical resection is considered the most effective therapy for LUAD patients, while the survival rate and quality of life after pneumonectomy are unsatisfactory. Studies that, most LUAD patients treated with standard cytotoxic chemotherapy ultimately become resistant to chemotherapy<sup>4</sup>. Thus, further investigations on the pathological mechanisms of LUAD to develop novel effective strategies or targeted agents are necessary.

Circular RNAs (circRNAs) are novel, endogenous, non-coding RNAs that have the covalently closed loop structures with high stability and conservativeness<sup>5</sup>. CircRNAs involve in normal developmental and pathological processes by functioning as oncogenes or antioncogenes, including cancers<sup>6,7</sup>. For example, circ-Foxo3 suppressed cell growth, migration, and invasion by regulating PTEN through miR-23a in esophageal squamous cell cancer<sup>8</sup>. Circ\_0074027 promoted tumor malignancy by inducing cell proliferation and invasion in NSCLC through miR-185-3p/BRD4/MADD pathway<sup>9</sup>. In LUAD, circ-TSPAN4 interacted with miR-665 to facilitate cell metastasis via ZEB1<sup>10</sup>. Circ\_0001946 contributed to cell growth through regulating miR-135a-5p/SIRT1 axis-mediated activation of Wnt/ $\beta$ -catenin signaling pathway<sup>11</sup>. All these studies suggest the potential application of circRNAs in targeted therapy in LUAD. Recently, it was showed that circ\_Band 4.1-like protein 2 (circ\_EP41L2) was decreased in LUAD and had the diagnostic potential for LU-

AD patients. However, its molecular role in the tumorigenicity of LUAD has not been clarified.

Cadherin superfamily is a transmembrane adhesion, calcium-dependent cell adhesion molecule family, which mediates cell-cell adhesion through homo- or heterotypic interactions<sup>12</sup>. Cell-cell adhesion plays a critical role in determining cell polarity, as well as in establishing and maintaining tissue homeostasis. Besides, this organized adhesion can be disturbed via genetic and epigenetic changes, which contribute to the loss of contact inhibition, changes in signaling, cell migration and stromal interactions during oncogenesis<sup>13</sup>. Thus, cadherins are important for the tumorigenesis of cancer cells<sup>13,14</sup>. Cadherin-4 (CDH4) (also known as retinal cadherin (R-cadherin)) is a classical cadherin from the cadherin superfamily and belongs to type I cadherins in cancer<sup>14</sup>. Recently, it was also discovered that CDH4 mRNA expression was down-regulated in lung cancer tissues, and was related to the grade and histotype, indicating the potential anti-tumor roles of CDH4 in lung cancer. Herein, we investigated the role of CDH4 in LUAD.

This research aimed to assess the expression patterns and the roles of circ\_EPB41L2 and CDH4 in LUAD, as well as investigated the potential regulatory relationship between circ\_EPB41L2 and CDH4 in the tumorigenesis of LUAD.

## Patients and Materials

### Patients and Specimens

32 pairs of matched LUAD tissues and matched para-tumor normal tissues were collected from patients who received surgical resections in Huaihe Hospital of Chongqing University. All specimens were identified using histological and pathological diagnoses by two experienced pathologists. This investigation was permitted by the Ethics Committee of Huaihe Hospital of Chongqing University, and all patients signed written informed consent.

### Cell Culture and Transfection

Human bronchial epithelial cell line (HBE) and four LUAD cell lines (A549, HCC827, H460, H1975, Calu-3) were obtained from Shanghai Academy of Life Science (Shanghai, China), and were cultured in the Dulbecco's Modified

Eagle's Medium (DMEM; Invitrogen, Carlsbad, CA, USA) in accompany with 10% fetal bovine serum (FBS), as well as 100 U/ml penicillin-streptomycin solution (Gibco, Rockville, MD, USA) at 37°C with 5% CO<sub>2</sub>.

The microRNA (miR)-211-5p (miR-211-5p), miR-211-5p inhibitor (anti-miR-211-5p), negative control (miR-NC) and miR-NC inhibitor (anti-miR-NC) were obtained from RiboBio (Guangzhou, China). Small interfering RNA (siRNA) sequence targeting circ\_EPB41L2/CDH4 (si-circ\_EPB41L2/si-CDH4), siRNA negative control (si-NC), circ\_EPB41L2 overexpression vector (circ\_EPB41L2-ov), control vector (NC), si-NC empty vector (pcDNA) and si-NC empty vector (pcDNA) were designed and synthesized by Invitrogen (Carlsbad, CA, USA). Cells were transfected with siRNAs using Lipofectamine 3000 (Invitrogen, Carlsbad, CA, USA) according to the standard procedure.

### Nucleic Acid Preparation and Quantitative Real-Time Polymerase Chain Reaction (qRT-PCR)

Total RNA was extracted using TRIzol reagent (Invitrogen, Carlsbad, CA, USA). Genomic DNA was isolated with a QIAamp DNA Mini Kit (Qiagen, Valencia, CA, USA), and cytoplasmic and nuclear RNA isolations were carried out using NE-PER Nuclear and Cytoplasmic Extraction Reagents (Thermo Fisher Scientific, Waltham, MA, USA) following the recommendations of manufacturer. Prime-Script RT reagent Kit (TaKaRa, Dalian, China) was used to synthesize the complementary DNA (cDNA), and then quantitative PCR was conducted using SYBR Green methods on the ABI7500 system. The fold changes were normalized with U6 or GAPDH and qualified by  $2^{-\Delta\Delta Ct}$  method. The specific primer sequences were listed as follows: circ\_EPB41L2, F 5'-CCTCTGGATCGGAAGACTGA-3' and R 5'-TGC-CAAGGGACAAGTGTATT-3'; miR-211-5p, F 5'-CGCTTCCCTTTGTCATCCT-3', and R 5'-TATGGTTTTTGTACTGTGTGAT-3'; CDH4, F 5'-CGTCCATCATCAAAGTCAAGGT-3', and R 5'-GGTCGTAGTCCTGGTCCTCCT-3'; U6, F 5'-GCTTCGGCAGCACATATACTAAAT-3', and R 5'-CGCTTCACGAATTTGCGTGTAT-3'; GAPDH, F 5'-AAGAAGGTGGTGAAGCAGGC-3', and R 5'-GTCAAAGGTGAGGAGTGGG-3'.

### Western Blot Assay

Transfected cells were collected and then lysed using the radioimmunoprecipitation assay (RIPA) buffer (Beyotime, Shanghai, China). Then cell lysates were separated by 10% sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE), shifted onto polyvinylidene difluoride (PVDF) membranes, and blocked with 5% nonfat-milk. After that, the membranes were interacted with primary antibodies against CDH4 (1:20000, ab76011, Abcam, Cambridge, MA, USA), Ki67 (1:1000, ab16667, Abcam, Cambridge, MA, USA), E-cadherin (1:1000, ab15148, Abcam, Cambridge, MA, USA), as well as GAPDH (1:10000, ab8245, Abcam, Cambridge, MA, USA), and followed by interaction with secondary horseradish peroxidase (HRP)-conjugated antibody (1:1000, ab9482, Abcam, Cambridge, MA, USA). Finally, protein bands were detected using an enhanced chemiluminescence (ECL) detection system (Thermo Fisher Scientific, Waltham, MA, USA).

### RNase R Treatment

Total RNA (2 mg) was interacted with RNase R without 3 U/mg of RNase R (Qiagen) at 37°C for 20 min. The resulting RNA was purified using a RNeasy MinElute Cleanup Kit (Qiagen, Hilden, Germany).

### Cell Proliferation Assay

Transfected cells were plated in 96-well plate for overnight, 10<sup>4</sup> cells (4,500 cells/well) of HCC827 cells were cultured in DMEM (4,500 cells/well) with 10% FBS. MTT solution (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) solution (Sigma-Aldrich, St. Louis, MO, USA) was supplied into per well. Subsequently, the supernatant was resolved at 4 h post-reaction, and followed interaction with 200  $\mu$ L dimethyl sulfoxide (DMSO; Sigma-Aldrich, St. Louis, MO, USA) to dissolve the generated formazan. Finally, the absorbance was examined at 490 nm on a microplate reader.

### Transwell Assay

For migration assay, transfected cells were seeded in the transwell upper chamber with 200  $\mu$ L serum-free DMEM, and then 700  $\mu$ L DMEM fixed with 10% FBS was added into the lower well chamber. After incubation for 24 h, cells on the lower face of the membranes were fixed and stained. Lastly, migrated cells were counted in six randomly selected visual fields. For invasion assay, the upper transwell chambers

were pre-coated with Matrigel (BD Bioscience, Franklin Lakes, NJ, USA), and other procedures of measurement was similar to the above migration assay, except that.

### Dual-Luciferase Reporter Assay

Wild (wt) type and mutant (mutant) circ\_EPB41L2 and CDH4 3' UTR containing the putative binding sites of miR-211-5p were amplified and inserted into the pmir-REPORT Vector (Promega, Madison, WI, USA) 549 and HCC827 cells were co-transfected with the constructed vectors and miR-211-5p mimics or miR-NC using Lipofectamine 3000 (Invitrogen) following the protocol of manufacturer. Finally, a Dual-Luciferase reporter assay kit (Promega, Madison, WI, USA) was used to detect the relative Luciferase activity.

### Orthotopic Xenograft Experiments In Vivo

Athymic BALB/c nude mice (4-6-week-old, 18-20 g) were purchased from Vital River Laboratory Animal Technology (Beijing, China). The experiments were permitted by the Animal Research Committee of Huaihe Hospital of Henan University and performed in compliance with the guidelines of the National Animal Care and Use Committee institution. Six mice were divided into 2 groups for stable injection of HCC827 cells (NC or circ\_EPB41L2). The tumor size in mice was detected every 3 days. After injection for 28 days, the tumor masses were weighed and harvested for further analysis.

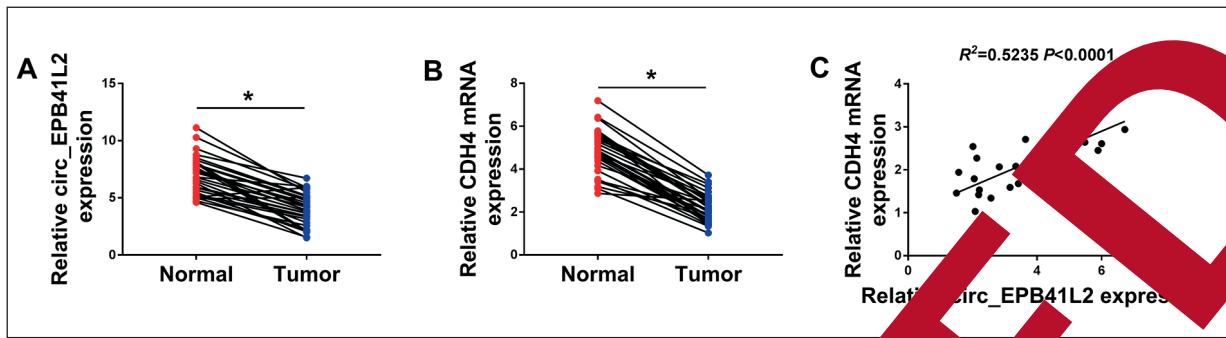
### Statistical Analysis

Data were expressed as the mean  $\pm$  standard deviation (SD). Statistical differences between groups were compared using one-way analysis of variance (ANOVA) followed by Tukey's post-hoc test or Student's *t*-test. The correlation analysis was analyzed using Spearman's correlation test. All statistical analysis was performed using GraphPad Prism 7 (GraphPad Inc., La Jolla, CA, USA). *p*-values less than 0.05 suggested statistically significant.

## Results

### The Expression of Circ\_EPB41L2 and CDH4 Is Down-Regulated In LUAD Tissues

To identify whether the circ\_EPB41L2 and CDH4 played crucial roles in LUAD tumori-

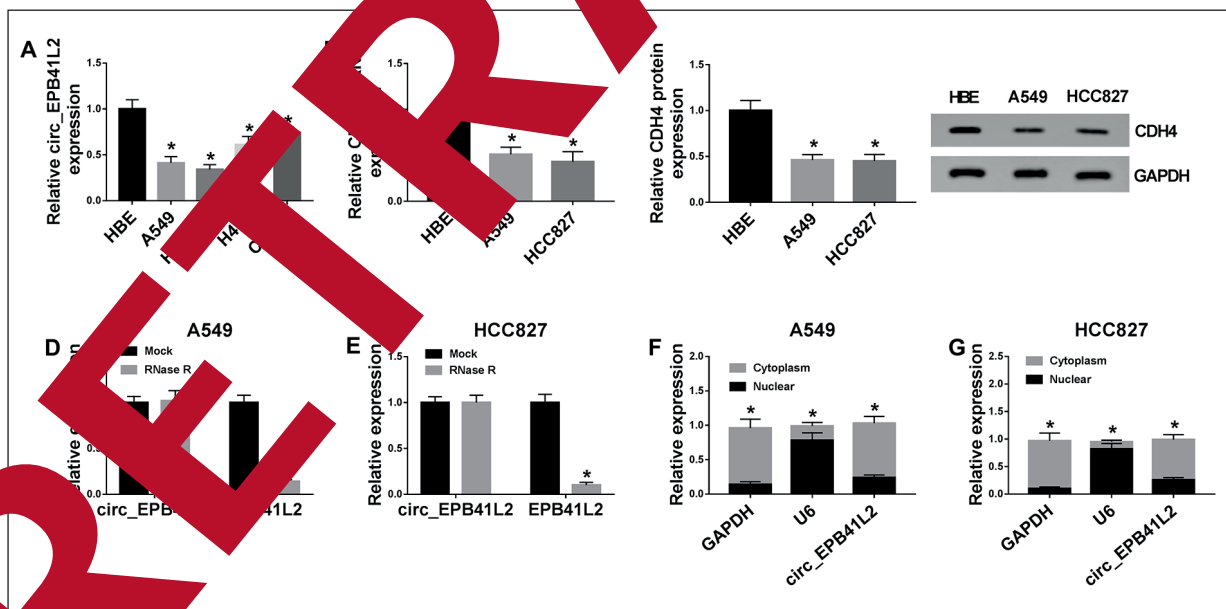


**Figure 1.** The expression of circ\_EPB41L2 and CDH4 is down-regulated in LUAD tissues. A, B, The expression of circ\_EPB41L2 and CDH4 was detected using qRT-PCR in LUAD tissues. C, The correlation between circ\_EPB41L2 and CDH4 was analyzed using Spearman's correlation test. \* $p < 0.05$ .

genesis, the expression of circ\_EPB41L2 and CDH4 was detected in 32 pairs of LUAD tissues and adjacent normal tissues. qRT-PCR analysis showed that circ\_EPB41L2 and CDH4 levels were markedly down-regulated in LUAD tissues (Figure 1A, B). Furthermore, a positive correlation between circ\_EPB41L2 and CDH4 expression in LUAD tissues was observed (Figure 1C). These results indicated that circ\_EPB41L2 and CDH4 possibly participated in tumorigenesis of LUAD.

### The Expression of circ\_EPB41L2 and CDH4 Is Down-regulated In LUAD Cell Lines

The expression of circ\_EPB41L2 and CDH4 was also detected in LUAD cell lines, and results showed that circ\_EPB41L2 and CDH4 levels were decreased in LUAD cell lines (Figure 2A-C). In addition, to confirm the existence of circ\_EPB41L2, we performed Northern blot analysis of total RNA from proliferating A549 and HCC827 cells. After treated with RNase R exonuclease, and qRT-PCR analysis exhibited that circ\_EP-



**Figure 2.** The expression of circ\_EPB41L2 and CDH4 is down-regulated in LUAD cell lines. A-C, The expression of circ\_EPB41L2 and CDH4 was detected in LUAD cell lines using qRT-PCR or Western blot, respectively. D, E, qRT-PCR analysis of circ\_EPB41L2 and EPB41L2 mRNA after treatment with RNase R in A549 and HCC827 cells was performed. F, G, Circ\_EPB41L2 in either the cytoplasm or the nucleus was analyzed using qRT-PCR analysis in A549 and HCC827 cells. \* $p < 0.05$ .

B41L2 existed in a circular form because of the resistance to digestion by RNase R (Figure 2D, E). Moreover, qRT-PCR analysis of nuclear and cytoplasmic RNAs in A549 and HCC827 cells indicated that circ\_EPB41L2 was preferentially localized in the cytoplasm (Figure 2F, G). Thus, circ\_EPB41L2 and CDH4 were down-regulated in LUAD cell lines.

### ***Circ\_EPB41L2 Acts As a Suppressor In the Proliferation, Invasion and Migration of LUAD Cells***

To explore the role of circ\_EPB41L2 in LUAD, we used two siRNA oligonucleotides to target the unique back splicing junction of circ\_EPB41L2, and circ\_EPB41L2 expression was expectedly decreased in A549 and HCC827 cells (Figure 3A, B). Besides, circ\_EPB41L2-overexpressing plasmid was synthesized, and circ-AKT3 expression was significantly increased in A549 and HCC827 cells (Figure 3A, B). After that, functional experiments were conducted. Results showed over-expressed circ\_EPB41L2 inhibited proliferation of A549 and HCC827 cells, reflected by the decreased proliferating cells (Figure 3C, D) and Ki67 protein level (Figure 3E). Afterwards, tube formation assay indicated that circ\_EPB41L2 restoration suppressed the migration and invasion of A549 and HCC827 cells (Figure 3F, G). Consistently, Western blot further demonstrated circ\_EPB41L2 restoration inhibited cell migration and invasion in LUAD by increasing the level of E-cadherin (Figure 3H). Conversely, knockdown of circ\_EPB41L2 promoted cell proliferation (Figure 3C, D), migration and invasion (Figure 3F, G) in LUAD. Taken together, circ\_EPB41L2 played a protective role by repressing proliferation, migration and invasion in LUAD cells.

### ***CDH4 Inhibits Cell Proliferation, Invasion and Migration In LUAD Cells***

To investigate the role of CDH4 in LUAD, the expression of CDH4 was increased or decreased by transfection with CDH4 or si-CDH4. As expected, CDH4 expression was up-regulated in A549 and HCC827 cells transfected with CDH4, but was down-regulated in A549 and HCC827 cells transfected with si-CDH4 (Figure 4A, B). Subsequently, MTT assay indicated CDH4 over-expression suppressed A549 and HCC827 cell proliferation (Figure 4C, D), and the inhibition of cell proliferation was also demonstrated by the reduction of Ki67 protein in A549 and HCC827 cells (Figure 4E). Meanwhile, CDH4

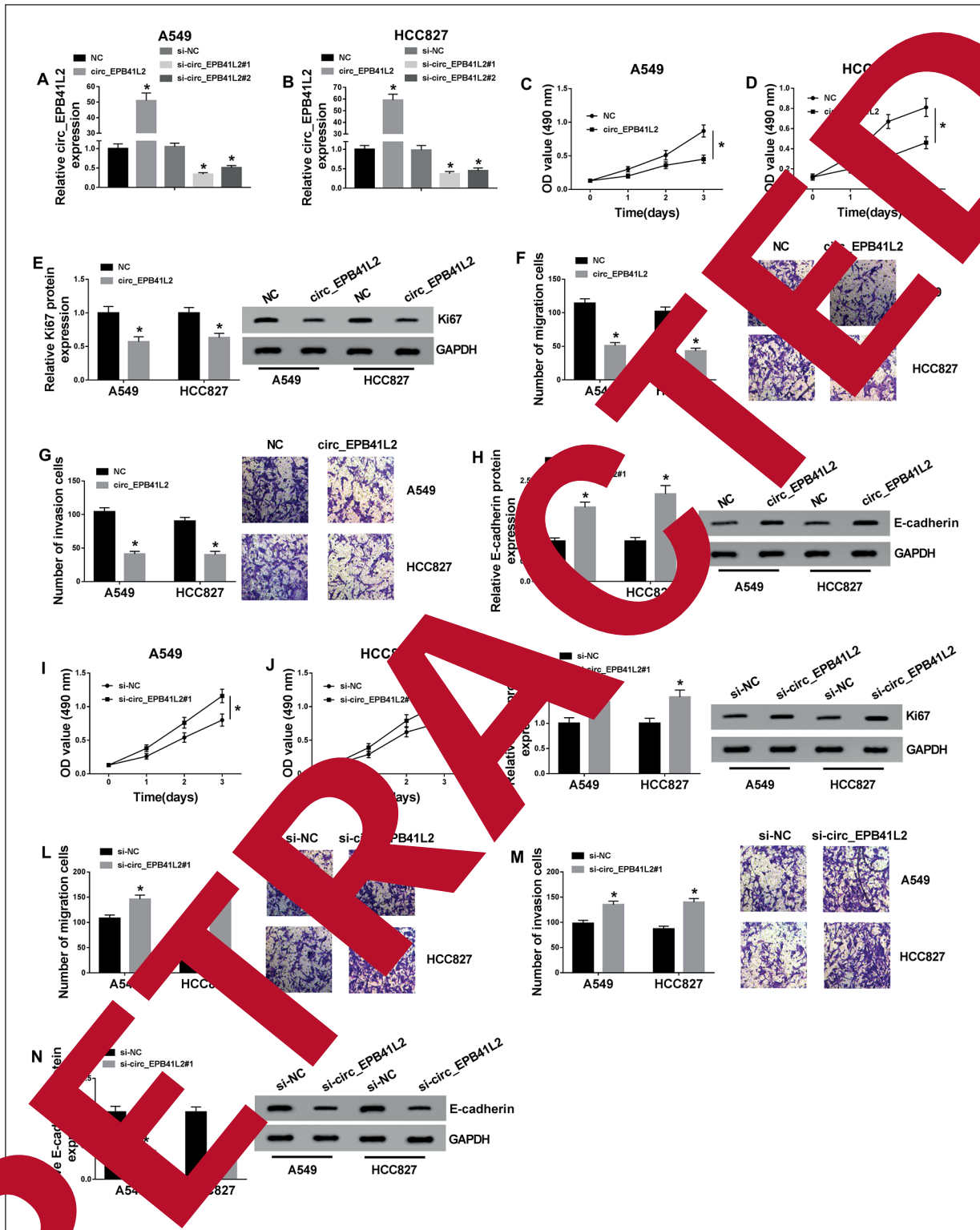
overexpression inhibited cell migration and invasion in LUAD, as illustrated by the decreased migrated and invaded A549 and HCC827 cells (Figure 4F, G), as well as the increased E-cadherin level in A549 and HCC827 cells (Figure 4H). To the contrary, CDH4 knockdown showed the carcinogenic effects to induce cell proliferation (Figure 4I-K), migration and invasion (Figure 4L-N) in LUAD. Altogether, CDH4 functioned as a tumor suppressor to inhibit cell tumorigenesis in LUAD.

### ***MiR-211-5p Directly Interacts With Circ\_EPB41L2 and CDH4***

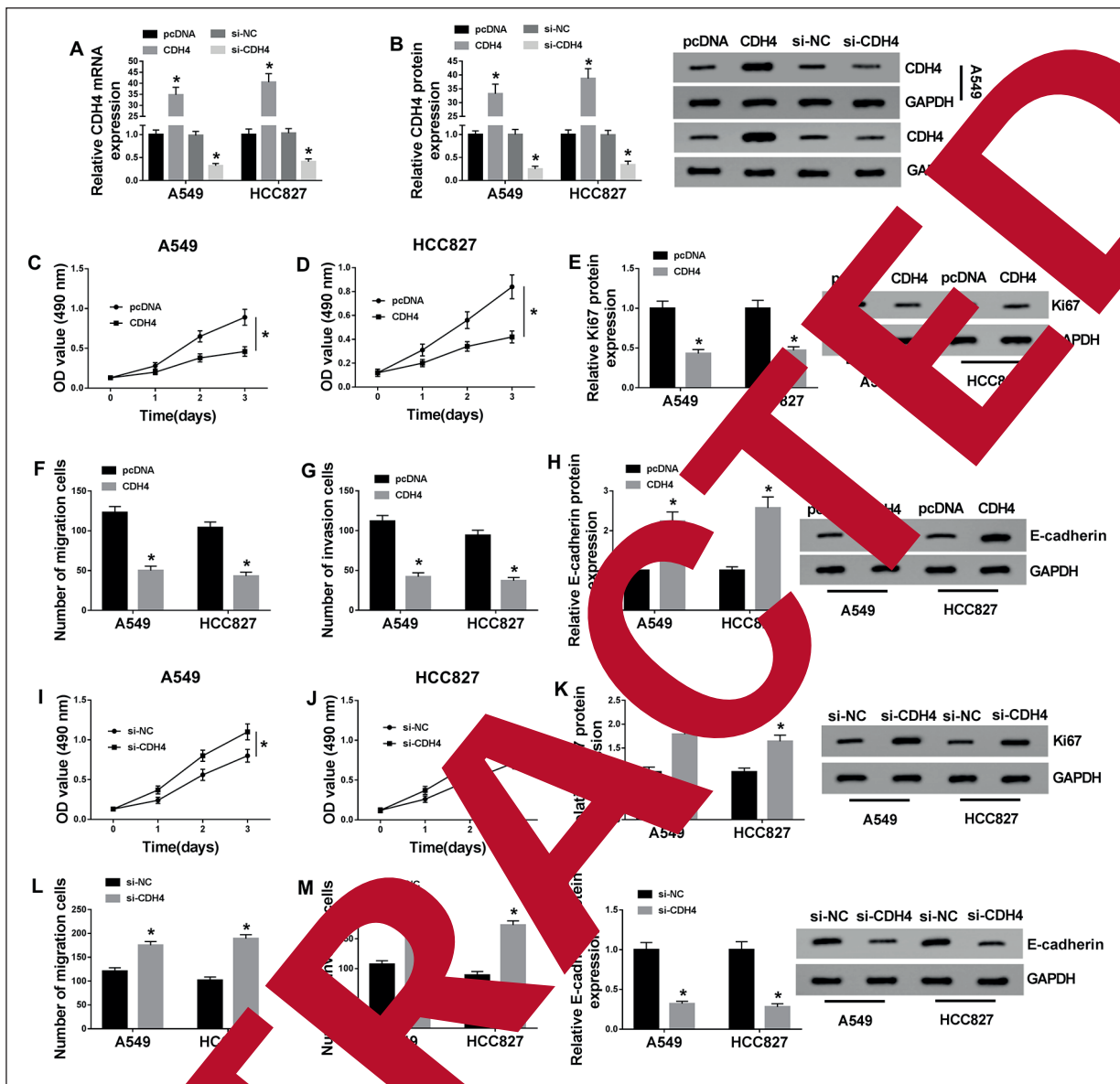
As circRNAs mostly serve as sponges of miRNAs to regulate downstream genes, we next investigated the ability of circ\_EPB41L2 to bind to miRNAs. According to the prediction of the online bioinformatics database (starBase3.0), we found miR-211-5p was a potential target of circ\_EPB41L2 (Figure 5A). Then, Dual-Luciferase reporter assay showed a great reduction of Luciferase activity in A549 and HCC827 cells co-transfected with circ\_EPB41L2-wt and miR-211-5p, reflecting the direct interaction between circ\_EPB41L2 and miR-211-5p (Figure 5B, C). In addition, we also observed that the expression of CDH4 mRNA was increased by circ\_EPB41L2 deletion, but was decreased by circ\_EPB41L2 overexpression in A549 and HCC827 cells (Figure 5D, E). Besides that, we also found miR-211-5p contained the binding sites of CDH4 (Figure 5F) through searching the Targetscan online bioinformatics database. Consistently, a significant reduction of Luciferase activity in A549 and HCC827 cells co-transfected with CDH4-wt and miR-211-5p confirmed the direct interaction between CDH4 and miR-211-5p (Figure 5G, H). Moreover, the level of CDH4 at mRNA and protein level was up-regulated by miR-211-5p inhibition but was down-regulated by miR-211-5p overexpression in A549 and HCC827 cells (Figure 5I-K). These results indicated miR-211-5p directly interacts with circ\_EPB41L2 and CDH4.

### ***Circ\_EPB41L2 Indirectly Regulates CDH4 Expression By Binding to MiR-211-5p In LUAD Cells***

Western blot assay was used to elaborate the regulatory relationship among miR-211-5p, circ\_EPB41L2 and CDH4 in LUAD cells. Results showed circ\_EPB41L2 overexpression increased the level of CDH4, while this increase was reversed by miR-211-5p up-regulation or CDH4



**Figure 5.** circ\_EPB41L2 acts as a suppressor in the proliferation, invasion and migration of LUAD cells. A549 and HCC827 cells were transfected with NC, circ\_EPB41L2, si-NC, or si-circ\_EPB41L2. **A, B,** The transfection efficiencies were detected by qRT-PCR. **C, D, I, J,** Cell proliferation was analyzed using MTT assay. **E, H, K, N,** Western blot was applied to examine the expression of Ki67 and E-cadherin. **F, G, L, M,** Cell migration and invasion were determined by transwell assay (magnification 100×), \* $p < 0.05$ .



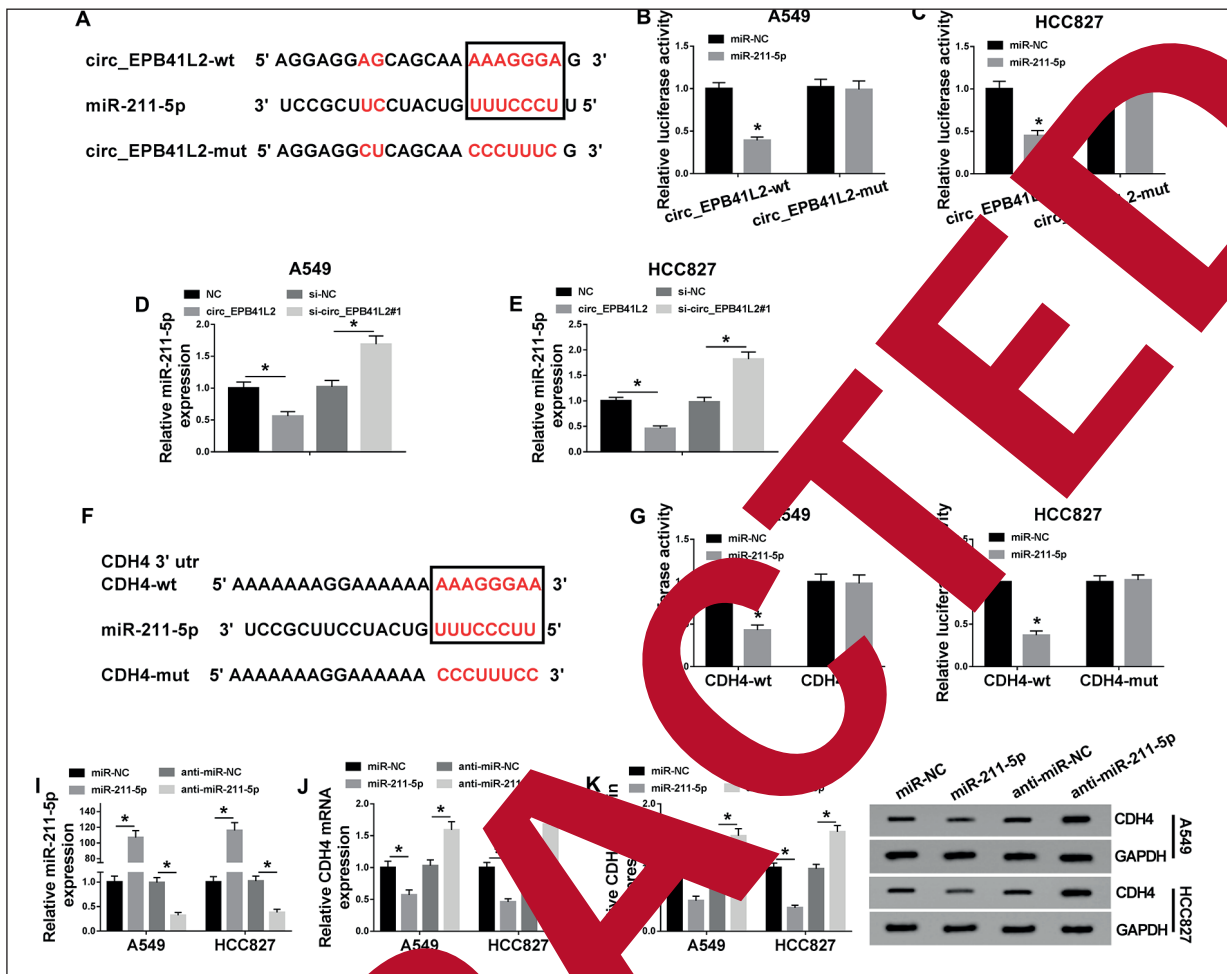
**Figure 4.** CDH4 inhibits cell proliferation, invasion and migration in LUAD. A549 and HCC827 cells were transfected with pcDNA, CDH4, si-NC or si-CDH4. **A, B,** The transfection efficiencies were detected by Western blot. **C, D, I, J,** Cell proliferation was analyzed using OD assay. **E, H, K, N,** The expression of Ki67 and E-cadherin was assessed using Western blot. **F, G, L, M,** Transwell assay was used to detect cell migration and invasion. \* $p < 0.05$ .

down-regulation in A549 and HCC827 cells (Figure 6C, D). Thus, a regulatory network of circ\_EPB41L2/miR-211-5p/CDH4 axis was identified in LUAD cells.

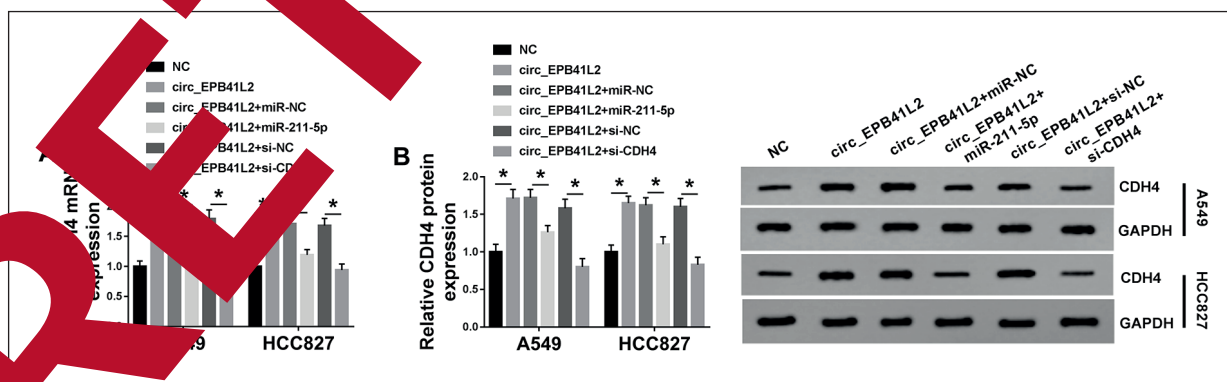
#### Circ\_EPB41L2 Performs Anti-Tumor Effects By Regulating CDH4 and MiR-211-5p in LUAD Cells

Based on the circ\_EPB41L2/miR-211-5p/CDH4 axis, we further explored the role of circ\_EPB41L2/miR-211-5p/CDH4 axis in LUAD tumorigenesis. Rescue assay suggested the inhi-

bition on proliferation (Figure 7A, B), migration and invasion (Figure 7D, E) induced by circ\_EPB41L2 overexpression was markedly reversed by miR-211-5p up-regulation or CDH4 down-regulation in A549 and HCC827 cells, indicating circ\_EPB41L2 suppressed cell tumorigenesis in LUAD by regulating CDH4 and miR-211-5p. Furthermore, Western blot analysis also exhibited that miR-211-5p up-regulation or CDH4 silence attenuated circ\_EPB41L2 overexpression mediated suppression on Ki67 expression (Figure 7C) and promotion on E-cadherin level (Figure 7F)

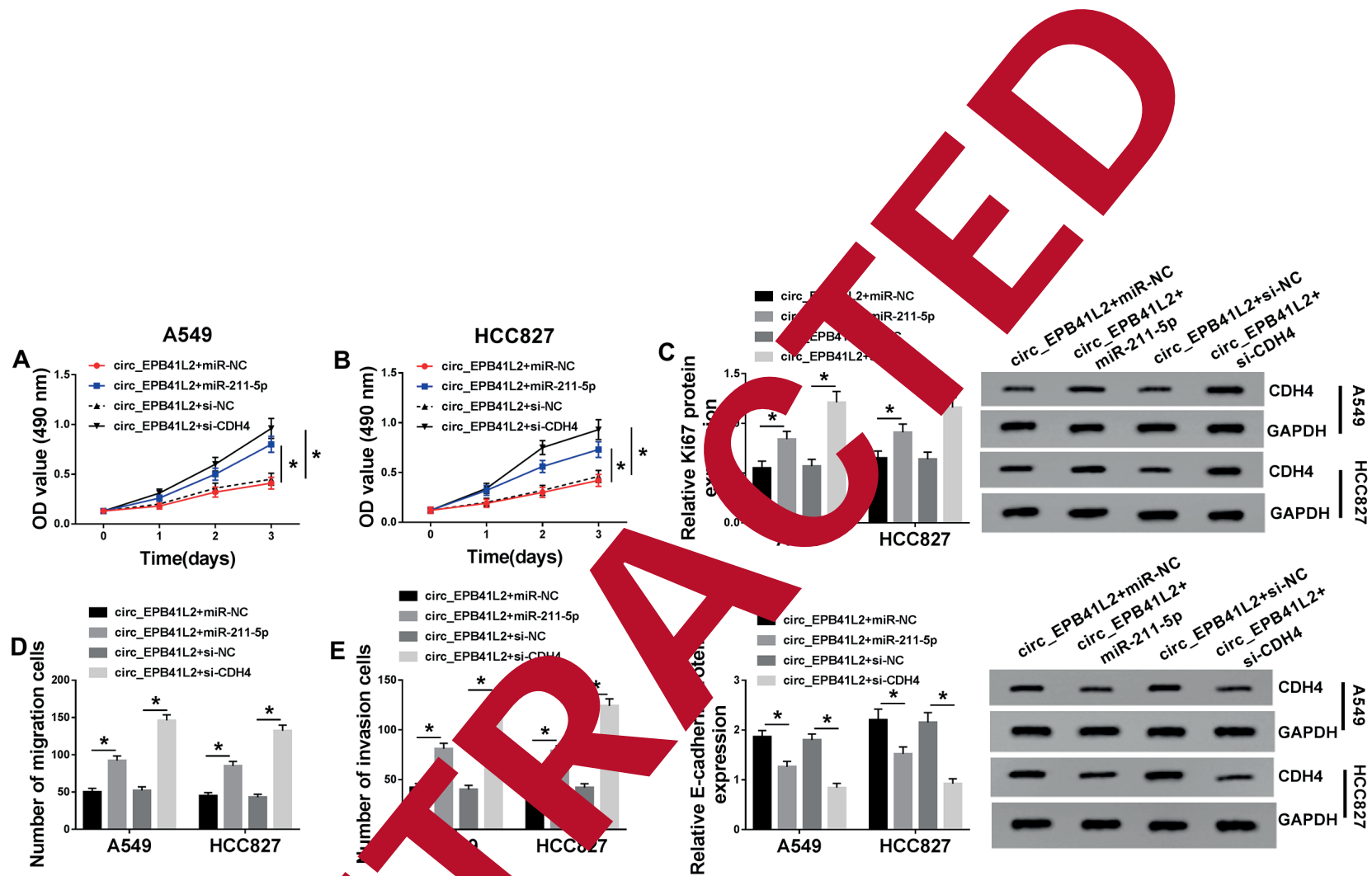


**Figure 5.** MiR-211-5p directly interacts with circ\_EPB41L2 and CDH4. **A**, The putative binding sites between circ\_EPB41L2 and miR-211-5p was presented. **B**, **C**, The interaction between circ\_EPB41L2 and miR-211-5p was confirmed by the dual-luciferase reporter assay in A549 and HCC827 cells, respectively. **D**, **E**, The expression of miR-211-5p was examined using qRT-PCR in cells transfected with NC, circ\_EPB41L2, or circ\_EPB41L2+miR-211-5p. **F**, The putative binding sites between CDH4 and miR-211-5p were listed. **G**, **H**, The interaction between CDH4 and miR-211-5p was confirmed using dual-luciferase reporter assay in A549 and HCC827 cells, respectively. **I**–**K**, The mRNA and protein expression of CDH4 was examined in A549 and HCC827 cells transfected with miR-NC, miR-211-5p, anti-miR-NC, or anti-miR-211-5p using qRT-PCR and Western blot, respectively. \* $p < 0.05$ .



**Figure 6.** Circ\_EPB41L2 indirectly regulates CDH4 expression by binding to miR-211-5p in LUAD cells. **A**, **B**, Western blot was used to detect the level of CDH4 in A549 and HCC827 cells transfected with NC, circ\_EPB41L2, circ\_EPB41L2 + miR-NC, circ\_EPB41L2 + miR-211-5p, circ\_EPB41L2 + si-NC, or circ\_EPB41L2 + si-CDH4. \* $p < 0.05$ .





**Figure 7.** Circ\_EPB41L2 performs anti-tumor effects by regulating CDH4 and miR-211-5p in LUAD cells. **A, B,** Cell proliferation was analyzed using MTT assay. **C, F,** The expression of Ki67 and E-cadherin was analyzed using Western blot. **D, E,** Transwell assay was used to detect cell migration and invasion. \* $p < 0.05$ .

in A549 and HCC827 cells. These data suggested that circ\_EPB41L2 suppressed cell malignancy in LUAD by regulating miR-211-5p/CDH4 axis.

### Circ\_EPB41L2 Inhibits LUAD Tumor Growth *In Vivo*

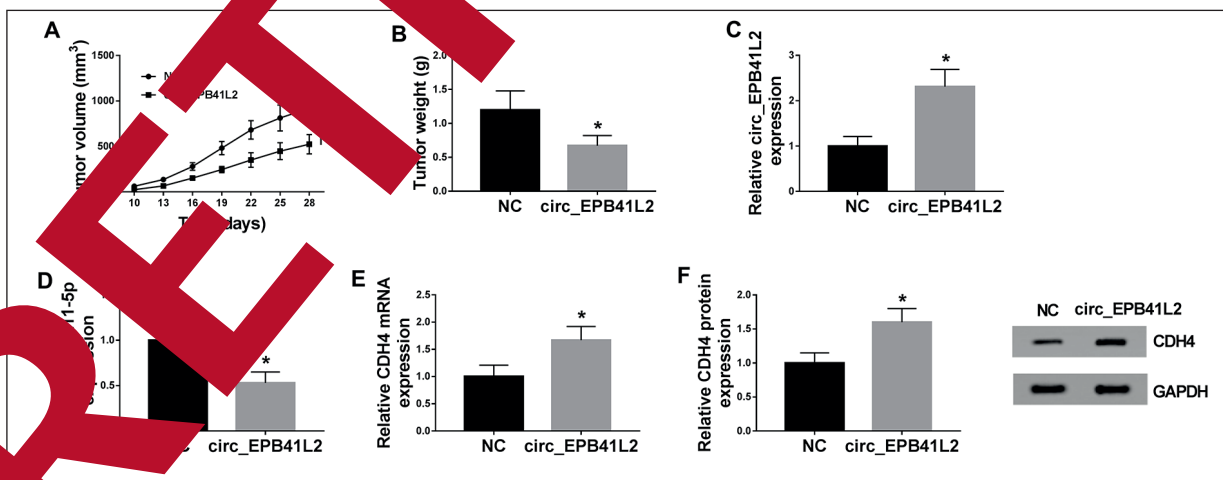
The carcinogenesis roles of circ\_EPB41L2 *in vivo* were further elucidated. We found circ\_EPB41L2 re-expression inhibited NSCLC tumor growth, demonstrated by the inhibition of tumor volume and weight in circ\_EPB41L2 overexpression group (Figure 8A, B). Additionally, molecular analysis showed circ\_EPB41L2 re-expression increased the levels of circ\_EPB41L2 (Figure 8C) and CDH4 (Figure 8E, F), but reduced the levels of miR-211-5p *in vivo* (Figure 8D). Collectively, these results revealed that circ\_EPB41L2 hindered LUAD tumor growth *in vivo* via regulating miR-211-5p and CDH4 expression.

## Discussion

The involvement of circRNA in cancers has been frequently reported, and its dysfunction has a significant impact on cancer cell proliferation, metastasis, and apoptosis<sup>15</sup>. CircRNAs can make contributions to cell proliferation, migration and invasion in LUAD cells. Overexpressed circPRKCI drives cell proliferation and tumorigenesis of LUAD by up-regulating E2F7 through sponging miR-45 and miR-589<sup>17</sup>. CircCRIM1 regulates leukemia inhibitory factor receptor signaling

and miR-182 to repress cell invasion and metastasis in LUAD<sup>18</sup>. CircPUM1 contributed to cell proliferation, migration and invasion, and induced apoptosis through binding miR-326 in LUAD<sup>19</sup>. Therefore, it is of great clinical significance to uncover the role of circ\_EPB41L2 in LUAD cell tumorigenesis. In the present study, we found decreased circ\_EPB41L2 in LUAD specimens and cell lines, circ\_EPB41L2 overexpression suppressed cell proliferation, invasion and migration of LUAD cells, while circ\_EPB41L2 down-regulation induced opposite effects. Besides, *in vivo* experiments also indicated circ\_EPB41L2 overexpression inhibited tumor growth through regulating miR-211-5p and CDH4. Therefore, circ\_EPB41L2 played a protective role in LUAD.

CircRNA can act as a molecular sponge of miRNAs (miRNAs) to involve in tumorigenesis and aggressiveness<sup>20</sup>. Herein, through Luciferase reporter assays, we confirmed circ\_EPB41L2 was capable of directly interacting with miR-211-5p in LUAD cells. miR-211-5p has been revealed to function as a tumor suppressor in renal cancer<sup>21</sup>, breast cancer<sup>22</sup>, and hepatocellular carcinoma<sup>23</sup>, and has carcinogenic effects on NSCLC<sup>24</sup>, suggesting the cell- or disease-context-dependent functions of miR-211-5p. In this study, we found that the inhibition on proliferation, migration and invasion mediated by circ\_EPB41L2 overexpression was markedly reversed by miR-211-5p up-regulation in LUAD cells. Thus, circ\_EPB41L2 suppressed cell malignancy in LUAD by regulating miR-211-5p.



**Figure 8.** Circ\_EPB41L2 inhibits LUAD tumor growth *in vivo*. **A**, Tumor volumes were calculated every 3 days. **B**, Tumor masses were excised and weighed on 28 days. **C**, **D**, The expression of circ\_EPB41L2 and miR-211-5p was determined using qRT-PCR. **E**, **F**, CDH4 expression was measured using qRT-PCR and Western blot. \* $p < 0.05$ .

Previous researches have reported that CDH4 participates in cell tumorigenesis, metastasis, differentiation and progression, and may perform anti-tumor functions in a variety of cancers, such as osteosarcoma, salivary adenoid cystic carcinoma, and gastric cancer<sup>25-27</sup>. This study investigated that CDH4 expression was down-regulated in LUAD, and overexpressed CDH4 performed anti-tumor effects in LUAD cells tumorigenesis, while decreased CDH4 promoted cell malignancy in LUAD. Besides that, we also demonstrated the upstream regulatory mechanism of CDH4 disorder in LUAD; circ\_EPB41L2 bound to miR-211-5p, ultimately regulating CDH4 expression and suppressing LUAD cell tumorigenesis. Additionally, CDH4 inhibition could reverse circ\_EPB41L2 re-expression mediated suppression on the carcinogenesis of LUAD. Thus, a circ\_EPB41L2/miR-211-5p/CDH4 regulatory network was identified in the tumorigenicity.

## Conclusions

We demonstrated that circ\_EPB41L2 up-regulated CDH4 expression by interacting with miR-211-5p to repress LUAD cell proliferation, migration, and invasion. The biological mechanisms of molecular therapies developing recently have provided promising strategies for the treatment of diverse cancers<sup>28,29</sup>. Emerging evidence confirmed that noncoding RNAs, including circRNAs and miRNAs, are vital biomarkers and therapeutic biological targets for diverse disease, including inflammatory diseases, metabolic diseases and cancers<sup>29,30</sup>. This study firstly identified the circ\_EPB41L2/miR-211-5p/CDH4 axis in LUAD, which provided a novel insight on the mechanisms underlying LUAD tumorigenesis, and might be targeted for molecular therapeutic benefits.

## Conflict of Interest

The Authors declare that they have no conflict of interests.

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