MicroRNA-191-5p promotes the development of osteosarcoma via targeting EGR1 and activating the PI3K/AKT signaling pathway

B. CHEN, Z.-Y. ZHENG, J.-Z. YANG, X.-G. LI

¹Department of Orthopedics, The First Affiliated Hospital, Zhejiang University, angzhou, C. ²State Key Laboratory for Diagnosis and Treatment of Infectious Diseases; For Affiliated Hosp College of Medicine, Zhejiang University, Hangzhou, China

Abstract. – OBJECTIVE: MicroRNA-191 (miR-191) has been reported to be abnormally expressed in human cancers and other diseases. The function of miR-191 was contradictory in different cancers. In the present study, we confirmed the specific function of miR-191-5p in osteosarcoma (OS).

PATIENTS AND METHODS: The effects of miR-191-5p on cellular behaviors of OS cells were investigated through MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide) nd transwell assay. The quantitative Real-tim d merase chain reaction (qRT-PCR) was ap examine the expressions of miR-191-5p an rly growth response gene 1 (EGR1). Western blo immunocytochemical assay were used to de the protein expression of EGR1. pinding re tionship between miR-191-5p was co firmed by Dual-Luciferase orter assay Xenograft tumor formation ay was c lucted to examine the in vivo effect **R-191** growth of OS. lated in OS **RESULTS:** MiR-1 p was tissues, which w lated to po gnosis of OS patients. M niR-191-5p ted cell , grath OS. Furthe d invasion V regulatproliferation, EGR1 was downing EGR1 OS tissues, regulate was associated prognosis of OS with p nts. MiR-191-5p nd to promote epithelial-mesenchymal was tra (E) and PI3K/AKT pathway, thus develop ent of OS. prom CON NS: M 91-5p promoted the demen argeting EGR1 and posiregula 13K/AKT signaling pathway. brds GR1, Osteosarcoma, PI3K/AKT.

Introduction

Osteosarcoma (OS) is a malignant bone tumor characterized by tumor cells directly invading

bone or bor OS mostly s the young and 20 years old, which is adolescen etwe rarely seen in people the age of 10 and after 30^{2} er, OS incide etween males and fes^o 15 5:2. Currently, cox prehensive treatments ed on high-dose chemotherapy and surgery are ely applied⁴. five-year survival rate of OS ts treated v standardized treatment is up and 6 of OS patients can retain their to all this, OS still has a high morlimbs lity rate among children and adolescents. Early and timely treatment have greatly ime survival rate of OS⁶. In particular, early diagnosis is also closely related to the prognosis of OS. Therefore, it is necessary to develop efficient biomarkers for the early diagnosis of OS. In recent years, MicroRNAs (miRNAs) have been identified as promising biomarkers involved in human cancers and other diseases7. Moreover, alternations of miRNA expressions have been reported to associate with pathogenesis and progression of OS⁸. For instance, miR-182 inhibits proliferation and promotes apoptosis in human OS cells by targeting HOXA99. Besides, upregulated miR-148a in osteosarcoma promotes cell growth by targeting PTEN (gene of phosphate and tension homology deleted on chromosome ten)¹⁰. In various miRNAs, alternation of miR-191 expression has been widely reported, which is contradictory in different human cancers. MiR-191 is upregulated and functioned as an oncogenic regulator in human hepatocellular carcinoma¹¹, breast cancer¹² and colorectal cancer¹³. Inversely, the downregulation of miR-191 has been examined in renal cell carcinoma14 and thyroid follicular tumors¹⁵. Those studies indicated that miR-191 might play an important role in the biology of human cancers. Wang et al¹⁶ demonstrated that increased expression of miR-191 could act as a potential serum biomarker for diagnosis and prognosis in human OS. Therefore, we further investigated the specific functions of miR-191-5p related with the progression of OS. Early growth response gene 1 (EGR1) belongs to the EGR family of transcription factors, which has a high homologous with EGR2, EGR3, and EGR4¹⁷. Abnormal expression of EGR1 has been found in the progression of human diseases and cancers¹⁸. However, the effect of EGR1 varies in different cancers. Zheng et al¹⁹ proposed that EGR1 is upregulated in gastric cancer which promotes tumor invasion and metastasis. EGR1 is capable of promoting growth and survival of prostate cancer cell²⁰. On the contrary, the downregulation of EGR1 has been identified in breast cancer cells²¹. Li et al²² indicated a crucial role of EGR1 in the transcriptional regulation of miR-20b in breast cancer. However, the interaction between EGR1 and miR-191-5p in OS is still unclear and need to be clarified. In the present work, miR-191-5p expression in OS tissues and cells was examined. The regulatory effects of miR-191-5p on cell proliferation, migration and invasion in OS, as well as epithelial-mesenchymal transition (EMT) and PI3K/ AKT signaling pathway were evaluated. Then, we verified the relationship between miR-191-5 EGR1 in OS. We hope these findings con tribute to better understanding the pathoge of OS and benefit the early diagnosis of OS.

Patients and M

Clinical Tissues

63 human OS tis es a liated Hoswere acquired from the Fir. pital of the Zhe University receiving patients signed inform None of the fore the operation. received any creatme frozen in **N** Tissues y nitrogen and then stored le -80°C refrigeral subsequent exrs. This study was approved by the Instiperi ommittee of The First Affiliated Tthic tut Hosph Zhejian Iniversity.

Cultu

DS, MGC Baos2 cell lines and human fetal reoblastic cell line hFOB1.19 were used for in the transformed series of the series of the series of the series of the series (Shanghai, a). Cells were seeded in Dulbecco's Modified E. Medium (DMEM; Gibco, Grand Island, NY, USA) with 10% fetal bovine serum (FBS; Gibco, Grand Island, NY, USA) and cultured at 37°C with 5% CO₂.

Quantitative Real Time-Polymerase Chain Reaction (qRT-PCR)

TRIzol reagent (Invitrogen, Carlsbad, CA, USA) was applied for extracting total RNA acco the standard method. Synthesis of com deoxyribose nucleic acid (cDNA) wa sed by the PrimeScript Reverse Transcriptase KaRa, Otsu, Shiga, Japan) based on the manufa instructions. Quantitative Real Time-Vymera in Reaction (RT-PCR) was carrie at through a RR fied Biosystems, Green PCR Master Mix û U6 ar City, CA, USA) on AB lyceraldeh le DH) we used 3-phosphate dehydroger GR1. T 191-5p as controls for m elative expressions w alculated us method. vere as fol-Primer sequ red in this s. AGACCGGMCCTTAC-3', lows: EGI F: 5 5'-GTCTAGAT R: TCCGTGGA-3'; miR-191 5'-CTGGT ACATCCTCCTG-3', R S-ACCATCGTGTCG CAAGG-3'; U6: F: GCTTCGGCACCACATATACTAAAAT-3', R: GCTTCAG TTGCGTGTCAT-3'; GAP-5'-CGCT CTGCTCCTCCTGTTC-3', R: Т GTTG 5'-1 ICCGACCTTCAC-3'.

Cell Transfection

miR-191-5p mimic or inhibitor, miR-191to point and negative control (NC) were obtained from Ribobio (Guangzhou, China). MG63 cells were transfected with Lipofectamine 2000 (Invitrogen, Carlsbad, CA, USA) based on the manufacturers' protocols.

MTT (3-(4,5-Dimethylthiazol-2-yl)-2,5-Diphenyl Tetrazolium Bromide) Assay

Transfected cells were cultured in 96-well plates with 2×10^3 /well and incubated for 24, 48, 72 h and 96 h. 20 µL MTT solution (Thermo Fisher Scientific, Waltham, MA, USA) was applied per well and incubated for 4 h at 37°C. Finally, the absorbance at 490 nm (OD=490 nm) was detected with a spectrophotometer.

Transwell Assays

Transwell chambers (8- μ m pore size membranes) were employed to measure the abilities of cell migration and invasion. The bottom chamber was added with 10% FBS and incubated at 37°C with 5% CO₂. The upper surface pre-coated with Matrigel (BD Biosciences, Franklin Lakes, NJ, USA) was used for cell invasion. Cell migration assay was conducted without Matrigel pre-coating. 2×10⁴ cells were cultured in the bottom chamber with serum-free medium. 24 h later, the migrated or invasive cells were fixed with methanol and stained with crystal violet. Finally, we counted the number of migrated or invasive cells using Image J software (NIH, Bethesda, MD, USA).

Western Blot Analysis

The protein samples were obtained using radioimmunoprecipitation assay (RIPA) lysis buffer (Beyotime, Shanghai, China). Proteins were separated through a 10% sodium dodecyl sulphate-polyacrylamide gel electrophoresis (SDS-PAGE) and transferred on polyvinylidene difluoride (PVDF) membranes (Millipore, Billerica, MA, USA). After blocking in 5% skim milk, the membranes were incubated with primary antibodies of E-cadherin, N-cadherin, vimentin and AKT, p-AKT, GAPDH primary antibodies (1:1000; Abcam, Cambridge, MA, USA) overnight at 4°C. After washing, they were incubated with the corresponding horseradish peroxidase-conjugated secondary antibodies (1:3000; Santa Cruz Biotechnology, Dallas, TX, USA). Then, the protein expression levels were measured by electrochemiluminescence (ECL, Pierce, Waltham USA).

Dual Luciferase Assay

The wild or mutant type of EGR1 3'-untr lated region (3'-UTR) were into pmirGLO Luciferase vector Iadisoi WI, USA) to perform Lug ase rep r exper-CR1 3'iments. Then, wild or m type UTR and miR-191-5p vim ual-Luciferinto MG63 cells. S *quently*, ase Assay System VI, USA) mega, Mad was applied to ciferase ac

Immune tochemica

The tion of OS tissue s dewaxed, hyand washed twice with Phosphate-Buffdrate <u>e (P</u> Gibco, Grand Island, NY, USA) ere ter block g with 5% goat serum for 5 S), se ons were incubated with luted antibody at 37°C for 1-2 h. GR1 three times, sections were in-PBS was. d with horseradish peroxidase (HRP)-conary antibody at 37°C for 1 h. After shing 5-times with PBS, diaminobenzidine B) mixture (Abcam, Cambridge, MA, USA) ed for color development of the section. The section was washed, counterstained, dehydrated, transparentized and mounted. Images were captured using a microscope.

Xenograft Tumor Formation Assay

is

Nude mice (4 weeks old) were provided by the Shanghai Lab Animal Research Center (Shanghai, China). All animal researches were a by the Animal Care and Use Ethics Co Zhejiang University Animal Center rst, 3×10 cells transfected with pre-miR-19 plasmid or negative control were injected into ight hind flank of nude mice. The tur r volu s observed every 3 days after /-day incu weeks later, mice were s nced by CO₂ asp tion and tumors were sted f arther stud.

Statistical And

al Prod-The data analyzed 19.0 (IBM, uct and S olutions (SI nd GraphPau Prism 6 (La 1, U. Armonk, Jolla, CA, USA). Da. e shown as mean \pm SD Student's The relationship beanal in miR-191-5p expression and clinic-pathoty cal features of OS patients was analyzed by χ^2 Kaplan-Me analysis was applied to draw rvival cur , and the log-rank test was tl he survival differences. A sigompar use nifican ce was defined at p < 0.05.

Results

MiR-191-5p Was Upregulated in OS Tissues

Primarily, miR-191-5p expression in OS tissues was detected by qRT-PCR. We found that miR-191-5p was markedly upregulated in OS tissues in comparison with normal tissues (Figure 1A). Moreover, we also analyzed the relationship between miR-191-5p expression and their clinic-pathological characteristics of OS patients. As shown in Table I, the expression of miR-191-5p was positively correlated to TNM stage (p=0.034). In addition, we also identified that miR-191-5p expression was negatively related to prognosis of OS patients (p=0.022, Figure 1B). Based on those results, we considered that miR-191-5p might be involved in the pathogenesis of OS and predict its prognosis.

MiR-191-5p Promoted Cell Proliferation, Migration and Invasion in OS

MiR-191-5p expression in U2OS, MG63, Saos2 and hFOB1.19 cell lines was detected. Similarly, miR-191-5p was also upregulated in U2OS, MG63, Saos2 cells contrast to that of hFOB1.19 cells (Fig-



expression was significantly enhanced by transct Target Was a L fection of miR-191-5p mimics, and was reduced in 191-5 h OS Cells of Fu

, EGR1 was predicted as a target ne of miR-191-5p in the database of TargetScan ww.targetscan.org) (Figure 3A). Then, acted Luciferase reporter gene assay to confirm that. Consistent with the prediction, we observed that Luciferase activity was reduced in MG63 cells co-transfected with miR-191-5p

Characteristics		miR-191-5p		<i>p</i> -value
		High	Low	
Age (yec				0.332
≥ 20	35	20	15	
	28	18	10	
<u>c</u>				0.651
M	33	23	10	
Fema	30	15	15	
vor size				0.218
cm	38	25	13	
cm	25	13	12	
7 Catago				0.034*
	23	11	12	
III-IV	40	27	13	
nph node metastasis				0.339
	30	16	14	
-8	33	22	11	

vn

onsisten

(Figur

Statistical analyses were performed by the χ^2 test.

MG63 cells transfected with miR-191-5p inh

MiR-191-5p overexpression promoted the

191-5p overexpression (Figure

miR-191-5p promoted cell iny

ation of MG63 cells, while miR-191-5p know

inhibited proliferative potential (Figure 2C) Similarly, cell migration was enhanced by r

*p < 0.05 was considered significant.



in the stand EGR1-Wt vector. However, the Luciferase activity of EGR1-Mut was not changed by transfection of miR-191-5p mimics (Figure 3B). Furthermore, the expression level of EGR1 was examined to be negatively correlated with miR-191-5p expression in OS tissues (p<0.05, R²=0.331; Figure 3C). Moreover, mRNA expression of EGR1 was declined by transfection of





91-5p and MG63 cells (Figure 3D) and creased corransfection of miR-191-5p inhilt c (Figure 3E). Therefore, miR-191-5p was afrectly target EGR1 and had a negve association with EGR1.

was Downregulated in OS Tissues

Subsequently, EGR1 expression was analyzed in OS tissues. IHC showed a positive detection of EGR1 protein expression in the cell nucleus of OS tissues (Figure 4A). Moreover, the protein expression intensity of EGR1 significantly decreased in OS tissues in comparison with the adjacent normal tissues (Figure 4B). Besides, high EGR1 expression was found to be correlated with good prognosis of OS patients (p=0.043, Figure 4C). EGR1 was suspected to participate in the tumorigenesis of OS and miR-191-5p might promote the development of OS *via* targeting EGR1.



Figure 4. and was downregulated OS tissues. **A-B**, The protein of ession of EGR1 in OS to be detected by immunohist the mistry. **C**, High EGR1 expression was related to long the rall surface of in OS patients. **p<0.01.

91-5, Dited EMT PI3K/Ak Ignaling Pathway

ar in

te, Western blot showed that the upgulation of miR-191-5p suppressed E-cadherin ession and promoted expressions of N-cadhe and Vimentin (Figure 5). On the contrary, the downregulation of miR-191-5p inhibited expressions of N-cadherin and Vimentin, but enhanced E-cadherin expression level (Figure 5).



EMPLOY EXAMPLE A Considered that miR-191-5p overtop promoted cell metastasis by regulating EMT. Moreover, we observed the protein expressions of AKT and p-AKT in MG63 cells transfected with miR-191-5p mimics or inhibitor to verify whether miR-191-5p regulated the PI3K/AKT signaling pathway in OS. Western blot showed that increased miR-191-5p expression apparently promoted expressions of AKT and p-AKT in MG63 cells (Figure 5). Inversely, decreased miR-191-5p expression inhibited expressions of AKT and p-AKT expression (Figure 5). MiR-191-5p was considered to promoted EMT and PI3K/AKT pathway in OS.

MiR-191-5p Promoted the Tumor Growth in vivo

Finally, we subcutaneously injected MG63 cells containing miR-191-5p stable transfection plasmid or miR-NC into nude mice. As shown in Figure 6A, the overexpression of miR-191-5p markedly enlarged the tumor volume compared with the control group. In Figure 6B, we also found the tumors with miR-191-5p stable transfection plasmid grew more quickly than that with miR-NC. These findings showed that miR-191-5p promoted the tumor growth of OS *in vivo*.



Discussion

In recent years, increasing miRNAs have been reported to associate with the pathogenesis of human cancers. MiRNAs could serve as therapeutic targets in human cancers²³. To date, more and more miRNAs have been proposed to regulate different biological processes of OS. For example, mi was found to inhibit proliferation, migration sion and EMT in OS by targeting ZEB1²⁴. demonstrated that miR-140 suppressed OS growth by enhancing anti-tumor immune resp and blocking mTOR signaling. er, miR was reported to predict poor OS p tients²⁶. However, further j stigation till need to be done to verify the f miR in OS Here, we found that my 191 tion of miR-OS tissues and cell s. The up 191-5p was identi iferation, o promote d d PI3K/ migration and well as EN AKT pathway in OS.

MiR-1 has been for o act as a tumor by regulating the 1 co-p53 pathway in atic cholangiocarcinoma²⁷. Same as the promot intra also identified the carcinogeneab ıdy, -5p in 📿 In addition, Shi et al²⁸ sis of uR-1 vas upregulated in human cated nich was similar to our recarc Moreover, the overexpression h this we su 2-191 has also been demonstrated to predict of s and promote proliferation and insion in esophageal squamous cell carcinoma²⁹. current research, we found that the upreguof miR-191-5p was related to poor prognosis of OS patients. Besides, Huang et al³⁰ proposed that miR-191 promoted OS cells proliferation by suppressing Chk2. Our work showed that miR-

191-5p promoted the colliferation of OS cells via course g EGR1. Like 191 demonstrated that no 191 targeted EGR1 and suppressed intimal to kening after cerotid injury. We also confirmed to EGR1 was correct target of miR-191-5p and provipated in the rogression of OS.

as sophisticated in the pathogenetion or of human cancers and diseases, which could as tumor suppressor-gene or oncogene in cancers. EGR1 has been reported to promote cell growth and survival in prostate cancer. On the contrary, the suppressive function of EGR1 has been identified in several cancers and diseases. For instance, miR-183 was found to function as an oncogene via targeting EGR1 to promote tumor cell migration³². In our research, we found that miR-191-5p promoted the migration of OS cells via targeting EGR1. In addition, miR-301b has been identified to promote the proliferation and EMT of bladder cancer cells by targeting EGR133, which was consistent with our findings. EGR1 served as a direct target that was downregulated in acute lymphoblastic leukemia³⁴, Alzheimer's disease³⁵ and B cell lymphomas³⁶. In OS, the downregulation of ERG1 was also examined and a negative correlation between EGR1 and miR-191-5p was detected in OS tissues. Therefore, we considered that miR-191-5p promoted the development of OS via targeting EGR1.

Conclusions

Increased expression of miR-191-5p was identified to be related to poor prognosis of OS patient. Moreover, miR-191-5p promoted cell proliferation, migration and invasion in OS by inhibiting EGR1 expression. MiR-191-5p also promoted EMT and PI3K/AKT signaling pathway in OS. Besides, miR-191-5p promoted OS tumor growth *in vivo*. These findings would help us develop novel diagnostic and therapeutic approaches of OS.

Conflict of Interests

The authors declare that they have no conflict of interest.

References

- 1) OTTAVIANI G, JAFFE N. The epidemiology of osteosarcoma. Cancer Treat Res 2009; 152: 3-13.
- MIRABELLO L, TROISI RJ, SAVAGE SA. Osteosarcoma incidence and survival rates from 1973 to 2004: data from the surveillance, epidemiology, and end results program. Cancer 2009; 115: 1531-1543.
- MIRABELLO L, TROISI RJ, SAVAGE SA. International osteosarcoma incidence patterns in children and adolescents, middle ages and elderly persons. Int J Cancer 2009; 125: 229-234.
- GELLER DS, GORLICK R. Osteosarcoma: a review of diagnosis, management, and treatment strategies. Clin Adv Hematol Oncol 2010; 8: 705-718.
- 5) GELDERBLOM H, JINKS RC, SYDES M, BRAMWELL VH, VAN GLABBEKE M, GRIMER RJ, HOGENDOORN PC, MC A, LEWIS IJ, NOOIJ MA, TAMINIAU AH, WANN Survival after recurrent osteosarcoma: day for 3 European Osteosarcoma Intergroup (EC domized controlled trials. Eur J Cancer 201 895-902.
- ZHOU W, HAO M, DU X, CHENNE Advances in targeted there is non-Discov Med 2014; 17: 30 - 07.
- 7) ZHAO Y, LIU X, LU YX, SoRNA-1 regulates the proliferation and app, so of recells by targeting to 1alph, so av most armacol Sci 2017 5580-555

G, Yand

sarcom

- 8) JONES KB, SAL Del Mare S, C M. Gaudio E, Nuovo G EBLANC K, PAL , Randall OCE CM, LIAN JB, AQEILAN RL, VOLINIA STEIN RI. mip VA signature ciate with pathogenesis progression o osarcoma. Cancer J12; 72: 1865-1877. R 9)
 - G ZF, WANG YJ, FAN SH, DU SX, Li XD, WU Lu J, Zhing YL. MicroRNA-182 downregulates the identification, inhibits proliferation, and the apoint is in human osteosarcoma cells is the etime in OXA9. Oncotarget 2017; 8: 11345-

ANG H, WILLI, XU T, LI C, WU J, HE Q, WANG G, NG C, LIU K, TANG H, JI F. Increased expression NA-148a in osteosarcoma promotes call growth by targeting PTEN. Oncol Lett 2016; 12: 3208-3214.

VAKIM E, SITBON E, FAERMAN A, TABAK S, MONTIA BELANIS L, DOV A, MARCUSSON EG, BENNETT CF, CHAJUT A, COHEN D, YERUSHALMI N. hsa-miR-191 is a candidate oncogene target for hepatocellular carcinoma therapy. Cancer Res 2010; 70: 8077-8087.

- 12) NAGPAL N, AHMAD HM, MOLPARIA B, KULSHRESHTHA R. MicroRNA-191, an estrogen-responsive microRNA, functions as an oncogenic regulator in human breast cancer. Carcinogenesis 2013; 34: 1889-1899.
- 13) ZHANG XF, LI KK, GAO L, LI SZ, CHEN K, ZHANG D, TU RF, ZHANG JX, TAO KX, WHANG D, TU RF, ZHANG JX, TAO KX, WHANG A CONTROL OF A CONTROL A CONTROL OF A CONTROL OF A CONTROL OF A CON
- 14) CHEN P, PAN X, ZHAO L, JIN L, LIN MAN J, HE T, ZHOU L, WU X, WANG Y, YU L, YANG Y, Y. MicroRNA-191-5p exerts a mor suppress of in renal cell carcinom 2xp Ther Med 20 1686-1693.
- 15) COLAMAIO M, BORBON ANCO M, FEDERICO Allante P A, CALIFANO D IIAPP NCONE G, BATTISTA sco A. I dow gulation s through thyroid foll plays a r CDK6 t Metab 2011; J Clin Endo -Eh 96: E
- 16) Wang Y, Ji F, Dannie Y, Yuan D. Increased expression of microPhysical as a potential serum set of diagnostical prognosis in human osteosarcoma. Cancer Branark 2015; 15: 543-550. Kim HJ, Hong JM, Yoon KA, Kim N, Cho DW, Choi JY, Lee IK, Kim Charly growth response 2 negatively modula posteoclast differentiation through regulation and helix-loop-helix proteins. Bone 51: 644-350.
- 18) BHA S, FANG F, TOURTELLOTTE W, VARGA J. Egr-1: new conductor for the tissue repair orchesdirects harmony (regeneration) or cacophony dis). J Pathol 2013; 229: 286-297.
- (9) Zhang L, Pu J, JIANG G, WENG M, HE J, MEI H, HOU X, TONG Q. Abnormal expression of early growth response 1 in gastric cancer: association with tumor invasion, metastasis and heparanase transcription. Pathol Int 2010; 60: 268-277.
- 20) VIROLLE T, KRONES-HERZIG A, BARON V, DE GREGORIO G, ADAMSON ED, MERCOLA D. Egr1 promotes growth and survival of prostate cancer cells. Identification of novel Egr1 target genes. J Biol Chem 2003; 278: 11802-11810.
- 21) Liu J, Liu YG, Huang R, Yao C, Li S, Yang W, Yang D, Huang RP. Concurrent down-regulation of Egr-1 and gelsolin in the majority of human breast cancer cells. Cancer Genomics Proteomics 2007; 4: 377-385.
- 22) LI D, ILNYTSKYY Y, KOVALCHUK A, KHACHIGIAN LM, BRONSON RT, WANG B, KOVALCHUK O. Crucial role for early growth response-1 in the transcriptional regulation of miR-20b in breast cancer. Oncotarget 2013; 4: 1373-1387.
- 23) GANDELLINI P, PROFUMO V, FOLINI M, ZAFFARONI N. MicroRNAs as new therapeutic targets and tools in cancer. Expert Opin Ther Targets 2011; 15: 265-279.
- 24) JIANG R, ZHANG C, LIU G, GU R, WU H. MicroR-NA-126 inhibits proliferation, migration, invasion, and EMT in osteosarcoma by targeting ZEB1. J Cell Biochem 2017; 118: 3765-3774.
- 25) JI X, WANG E, TIAN F. MicroRNA-140 suppresses osteosarcoma tumor growth by enhancing anti-tumor immune response and blocking mTOR signaling. Biochem Biophys Res Commun 2018; 495: 1342-1348.

- 26) REN X, SHEN Y, ZHENG S, LIU J, JIANG X. miR-21 predicts poor prognosis in patients with osteosarcoma. Br J Biomed Sci 2016; 73: 158-162.
- 27) Li H, ZHOU ZQ, YANG ZR, TONG DN, GUAN J, SHI BJ, NIE J, DING XT, LI B, ZHOU GW, ZHANG ZY. MicroR-NA-191 acts as a tumor promoter by modulating the TET1-p53 pathway in intrahepatic cholangiocarcinoma. Hepatology 2017; 66: 136-151.
- 28) SHI X, SU S, LONG J, MEI B, CHEN Y. MicroRNA-191 targets N-deacetylase/N-sulfotransferase 1 and promotes cell growth in human gastric carcinoma cell line MGC803. Acta Biochim Biophys Sin (Shanghai) 2011; 43: 849-856.
- 29) GAO X, XIE Z, WANG Z, CHENG K, LIANG K, SONG Z. Overexpression of miR-191 predicts poor prognosis and promotes proliferation and invasion in esophageal squamous cell carcinoma. Yonsei Med J 2017; 58: 1101-1110.
- 30) HUANG YZ, ZHANG J, SHAO HY, CHEN JP, ZHAO HY. MicroRNA-191 promotes osteosarcoma cells proliferation by targeting checkpoint kinase 2. Tumour Biol 2015; 36: 6095-6101.
- 31) LI Y, MCROBB LS, KHACHIGIAN LM. MicroRNA miR-191 targets the zinc finger transcription factor Egr-1 and suppresses intimal thickening after carotid injury. Int J Cardiol 2016; 212: 299-302.

- 32) SARVER AL, LI L, SUBRAMANIAN S. MicroRNA miR-183 functions as an oncogene by targeting the transcription factor EGR1 and promoting tumor cell migration. Cancer Res 2010; 70: 9570-9580.
- 33) YAN L, WANG Y, LIANG J, LIU Z, SUN X, LANG M, 301b promotes the proliferation poliity, and epithelial-to-mesenchymal transmit of bladder cancer cells by targeting EGN chem Cell Biol 2017; 95: 571-577.
- 34) VERDUCI L, AZZALIN G, GIGLE, S, CAR, LAU-DADIO I, FULCI V, MACINE, MICRORNA, LAUhances cell proliferation in acute lympholeukemia by target GR1. L Res 2015, J: 479-485.
- 35) ZHU QB, UNN ρΑ Û, Κ, Ηυ ERWER MicroR-R, BALESAR ao J, Bad WAAP NA-132 a arly growth re n nucleus basalis rt during the se of Alzhei-2016; 139: 08-921. mer's eas
- 36) CONTRERAS JR, PALENDAY JK, TRAN TM, FERNANDO TECHNERIGUEZ-MALAVA THE DOSWAMI N, ARBOLEDA VA, D, RAO DS. N. RNA-146a modulates B-cell oncogenesis by regulating Egr1. Oncotarget 2015; 6: 11023-11037.

