

Effect and mechanism of propofol in hepatic ischemia/reperfusion injury of rat

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Abstract. – **OBJECTIVE:** Hepatic ischemia/reperfusion (I/R) injury remains to be one of the most common clinical diseases. This study aimed to explore the potential effect and mechanism of propofol in protecting rat liver from I/R injury.

MATERIALS AND METHODS: The hepatic I/R model was established in Sprague-Dawley (SD) rats by perfusing the liver with heparinized cold saline through the portal vein for 20 min. The rats were then received a 100 mg/kg/d propofol administration for the continuously 10 days. The hepatic function indexes of ALT, AST, and GGT were detected by ELISA. The apoptosis of hepatic cells was assessed by TUNEL assay, and Bax and Bcl-2 expression changes were detected by qRT-PCR and Western blotting. In addition, serum pro-inflammatory factors and signaling pathway-related protein expression were detected.

RESULTS: Propofol markedly attenuated the increases of ALT, AST, and GGT induced by I/R. Propofol reduced I/R-induced apoptosis and pro-inflammatory factors secretion. In addition, more, propofol could promote the expression of phosphorylated-AKT and inhibited the expression of p-mTOR.

CONCLUSION: Propofol protects hepatic I/R injury partly by reducing apoptosis and reducing the release of pro-inflammatory cytokines, which is possibly involved in the modulation of the PI3K/AKT/mTOR signaling pathway. All these data suggest that propofol may play certain positive roles in protecting the liver from I/R injury.

Key Words: Propofol, hepatic ischemia/reperfusion (I/R) injury, apoptosis, pro-inflammatory cytokines, PI3K/AKT/mTOR

Introduction

Hepatic ischemia-reperfusion (I/R) injury refers to liver tissue ischemia and hypoxia because of liver blood flow interruption or inadequate perfu-

sion in consequence of various reasons, which is recognized as a kind of uncontrolled inflammatory cascade¹⁻³. The reperfusion not only can't be able to restore functions of hepatic tissues, but aggravate the function of the liver cell metabolic disorders and structural damage⁴⁻⁶. Thus, hepatic I/R leads to organs dysfunction and failure, and other complications after liver resection and liver transplantation surgery, directly affecting the achievement rate of surgery and postoperative survival rate⁷⁻¹¹. With the deepening research of the hepatic I/R injury mechanism, the researchers found that hypoxic preconditioning (PPC) could effectively protect hepatic I/R injury¹²⁻¹⁴.

Regarding the present work, the hepatic I/R injury protective drugs are mainly oxygen-free radical scavenger, calcium antagonist, Kupffer cell activation inhibitors and drugs that could improve microcirculation or cell energy metabolism¹⁵⁻¹⁸. In the report of Li et al¹⁸, animal experiments were constructed and shown the role of the calcium channel blockade on hepatic I/R injury protection and found it assist the recovery of secretory function in hepatocytes. Moreover, the research of Chen et al¹⁹ illustrated that gadolinium chloride (GdCl₃) significantly weaken I/R-induced myocardial apoptosis in rats by inhibiting activation of both death receptor and mitochondria-mediated pathway. However, the toxic and side effects of these drugs affect its clinical application. Propofol is a new type of clinical commonly used and named as a kind of safety anesthetics, but the role of propofol in hepatic I/R injury research is still insufficient^{20,21}.

This research works on the new uses of anesthetic, aiming to explore the function and mechanism of propofol in hepatic I/R. The hepatic function was detected by ELISA, and propofol was found remarkably restores the liver function. In addition, propofol significantly ameliorated

apoptosis by increasing the Bcl-2/Bax ratio. Additionally, we found that propofol reduced the release of pro-inflammatory cytokines in hepatic I/R injury. qRT-PCR and Western blot results showed that propofol is possibly involved with the modulation of the PI3K/AKT/mTOR signaling pathway in hepatic I/R protection. In summary, propofol protects against hepatic I/R partly by reducing apoptosis and reducing the release of pro-inflammatory cytokines, which is possibly involved with the modulation of the PI3K/AKT/mTOR signaling pathway. All these findings suggest that propofol may be a new therapeutic target for hepatic I/R injury.

Material and Methods

Experimental Animals

Male Sprague-Dawley (SD) rats, aged 10 weeks (200 ± 10 g) were obtained from the Laboratory Animal Center of Tianjin Medical University (Tianjin, China). The rats were randomly divided into three groups: 1) Sham group ($n = 20$ rats), in which rats received a sham operation; 2) I/R group ($n = 30$ rats), in which I/R model was established; 3) I/R + propofol group ($n = 30$ rats), in which I/R model was first established and then the cells were treated with 10 mg/kg/d propofol (Shanghai Shengshun Biological Technology Co., LTD, Shanghai, China) for 10 days. Each group should ensure that at least 15 rats survived until the end of the research. Five rats in each group were killed at 1, 6, 12, 18, and 24 h after reperfusion. All the animal experiments were approved by the Ethics Committee of Hunan Provincial People's Hospital, The First Affiliated Hospital of Hunan Normal University.

Animal Model of Hepatic I/R

The rats were performed a midline laparotomy under chloral hydrate anesthesia. After clamping the hepatic artery, portal vein, suprahepatic vena cava (SHVC), and infrahepatic vena cava (IHVC), the portal vein and the IHVC were cannulated with a polyethylene tube and the liver was also perfused through the portal vein with heparinized cold saline (2.5 IU/mL, 10 mL/min) to wash out all the blood through SHVC. The transfixion pin was removed and the incision site was repaired after 20 minutes of cold perfusion. The anhepatic phase ended after all clips were unclamped²².

Determination of Plasma Liver Enzyme and Oxidation-related Parameters

The serum was obtained by centrifuging blood samples. The level of alanine aminotransferase (ALT), gamma-glutamyl transpeptidase (GGT) and aspartate aminotransferase (AST) were respectively detected by the spectrophotometer using an automated clinical chemistry analyzer 5400; Beckman Coulter, Brea, CA, USA²².

TUNEL Assay

The frozen liver sections were sectioned into slices of 5-mm thickness. *In situ* apoptosis was performed using TUNEL assay kit (Promega, Madison, WI, USA). We counted the cells displaying brown staining within the nucleus as apoptotic cells. The number of apoptotic cells were counted by a person blinded to the group assignment by 3 nonoverlapping microscopic eyes under high-power magnification ($\times 400$) and expressed as percentage²².

Determination of TNF- α and IL-6

Serum TNF- α and IL-6 levels were determined using an ELISA kit (Biosource International, CA, USA)²¹.

Western Blot Analysis

Liver tissues were rapidly homogenized in 200 mL of extraction protein buffer containing 50 mM Tris-HCl, pH 7.4, 2.0 mM EDTA, 2 mM Na_3VO_4 , 50 mM NaF, 1 mM 4-(2-aminoethyl) benzenesulfonyl fluoride hydrochloride (AEB-SF), 10 $\mu\text{g}/\text{mL}$ aprotinin, 10 $\mu\text{g}/\text{mL}$ leupeptin, and 10 $\mu\text{g}/\text{mL}$ pepstatin A²³. After incubation for 30 min on ice, the supernatant was centrifuged at 300 g for 10 min. Protein samples were separated on sodium dodecyl sulphate-polyacrylamide gel electrophoresis (SDS-PAGE) gels and transferred to polyvinylidene difluoride membranes (Millipore, Bedford, MA, USA)³, after boiling for 5 min at 95°C. Then, the membranes were incubated with the appropriate primary antibodies (Santa Cruz Biotechnology, Santa Cruz, CA, USA) overnight after blocking with 5% skim milk in TBS-T (powder-Tris-buffered saline with 0.1% Tween 20) for 1 h. Membranes were washed 3 times with TBS-T and incubated with horseradish peroxidase (HRP)-conjugated secondary antibodies (Santa Cruz Biotechnology, Santa Cruz, CA, USA) for 2 h. The final results were obtained by exposure to Kodak film (NY, USA).

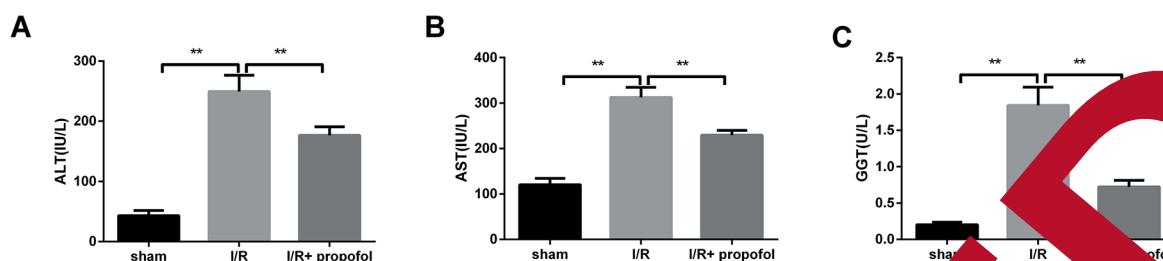


Figure 1. Effect of propofol treatment on hepatic ischemia-reperfusion (I/R) injury. Serum samples were collected from the rats in sham, I/R and I/R + propofol groups. **(A)** Alanine aminotransferase (ALT), **(B)** aspartate aminotransferase (AST), and **(C)** gamma-glutamyl transpeptidase (GGT) indexes were tested. Data presented as mean \pm SD (n = 6/group). ** $p < 0.01$ (Student *t*-test).

qRT-PCR

Total RNA was isolated from the liver tissue sample using the Trizol reagent (Invitrogen, Carlsbad, CA, USA). qPCR was performed using a Light-Cycler[®] 480Real-Time PCR System (Roche, Basel, Switzerland) and the SYBR Green qPCR Master Mix (2X) (Fermentas, Waltham, MA, USA).

Statistical Analysis

The data are presented as the mean \pm SD. Statistical analyses were performed using SPSS 20.0 statistical software (SPSS Inc., Chicago, IL, USA). Differences between two groups were analyzed by Student *t*-test. $p < 0.05$ was considered statistically significant.

Results

Propofol Restored the Liver Function in I/R

I/R mode was established in rats and the rats were received a 10 mg/kg/d propofol administration for the continuous 30 days. Then, the serum of the rats was collected and the hepatic index was detected using ELISA. As shown in Figure 1, ALT, AST, GGT indexes in I/R group were all significantly increased, while the liver function improved significantly after propofol administration. These data suggest a protective role of propofol in I/R injured liver.

Propofol Ameliorates Cell Apoptosis in I/R

TUNEL assay was performed to assess the ratio of apoptotic cells in the liver of the three groups. As results show in Figure 2A, the hepatic cell apoptosis induced by the I/R was decreased by propofol intervention. Next, we extracted the total mRNA and protein in the liver of the three groups and detected the expression levels

of Bax and Bcl-2. Bax is one of the pro-apoptosis promoting genes, while Bcl-2 is an anti-apoptotic gene. As shown in Figure 2 B and 2C, Bax was highly expressed in I/R group and the Bcl-2 was low expressed. More important, propofol intervention significantly alleviated I/R induced Bax up-regulation and Bcl-2 down-regulation.

Propofol Reduces Release of Pro-inflammatory Cytokines in I/R

HIR injury is accompanied by inflammation, thus we tested the influence of propofol on the release of pro-inflammatory cytokines in I/R. The results given in Figure 3 showed that propofol reduces pro-inflammatory factors release of IL-6, TNF- α , and MIP2 in I/R.

Propofol Promotes AKT Phosphorylation and Inhibits p-mTOR

Previous studies^{24,25} have reported the hepatic I/R injury may involve AKT/mTOR signaling pathway. So we performed qRT-PCR and Western blot experiments to analysis RNA and protein expression of AKT and mTOR in the liver of these three groups. As shown in Figure 4, the phosphorylated forms of AKT and mTOR were both up-regulated after I/R injury, while total AKT and mTOR was both down-regulated. Propofol administration could enhance I/R induced abnormal expression of p-AKT and AKT, and alleviate I/R induced abnormal expression of p-mTOR and mTOR.

Discussion

Hepatic I/R injury is confirmed an unavoidable consequence during hepatic resection, liver transplantation, and hypovolemic shock⁷. It has been implicated in the pathophysiology of many clinical entities following hepatic surgery and transplanta-

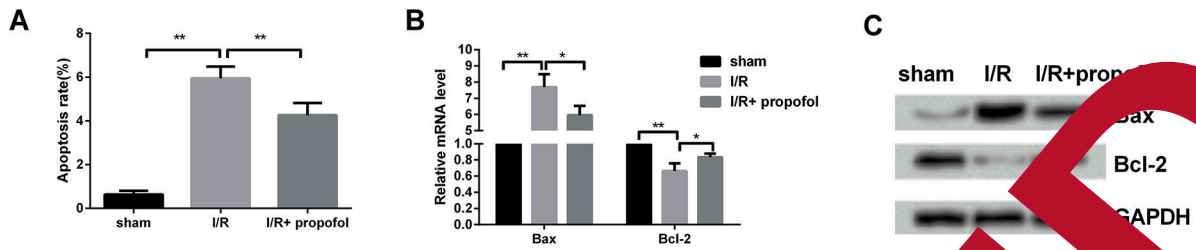


Figure 2. Effects of propofol on apoptosis in the liver after hepatic ischemia-reperfusion (I/R) injury. (A) TUNEL staining was performed to detect the apoptotic cells rate of sham, I/R, and I/R + propofol groups. (B) The mRNA and (C) protein levels of Bax and Bcl-2 were examined by qRT-PCR and Western blotting respectively. Data presented as mean \pm SD (n = 5/group). *, $p < 0.05$; **, $p < 0.01$ (Student *t*-test).

tion, as well as the dysfunction and injury of other organs¹². Although the research on hepatic I/R injury has made great progress, it is still a major cause of morbidity and mortality following liver surgery²⁶. In the present work, we found that propofol could partially recover I/R induced an increase in ALT, AST, and GGT levels. Propofol alleviated I/R induced apoptosis and the release of pro-inflammatory cytokines. Besides, propofol aggravated I/R induced p-AKT up-regulation and AKT down-regulation, as well as alleviated I/R induced p-mTOR up-regulation and mTOR down-regulation.

As a new type of anesthetics commonly used in clinical, the use of propofol has been reported to be safe for atrial fibrillation ablation, catheter ablation, internal cardioverter defibrillator implantation and many other ways. Furthermore, several investigations have revealed the protective role of propofol in I/R injury of liver. For instance, propofol can protect the liver from ischemia by reducing I/R induced increase in plasma ALT and AST^{29,30}. In accordance with the reported literatures, our study also confirmed the protective effects of propofol on hepatic I/R, that it could dramatically reduce ALT, AST and GGT levels in plasma.

Apoptosis is a major mechanism of cell death after hepatic I/R injury. Thus, we detected the apoptotic cells rate and the expression of apoptosis-related proteins, *i.e.*, Bcl-2 and Bax, to further understand the role of propofol in hepatic I/R injury. The results clearly showed that propofol significantly partially ameliorated apoptosis by increasing the Bcl-2/Bax ratio. Also, the inflammatory response plays an important role in liver dysfunction after hepatic I/R injury²¹. We tested the release of pro-inflammatory cytokines IL-6, TNF- α and MIP2 in hepatic I/R injury and finally found that propofol can decrease the release of pro-inflammatory cytokines in hepatic I/R injury. These findings were all in line with the previous research that, propofol can protect the liver from I/R injury by modulating the inflammatory responses and liver apoptosis³².

Evidence has strongly suggested that the PI3K/AKT/mTOR signaling pathway played an important role in hepatic I/R injury³³. To demonstrate the mechanism of propofol on hepatic I/R injury progress, we constructed qRT-PCR and Western blot experiments and tested AKT and mTOR expressions at both the mRNA and protein levels. Propofol was reported to activate

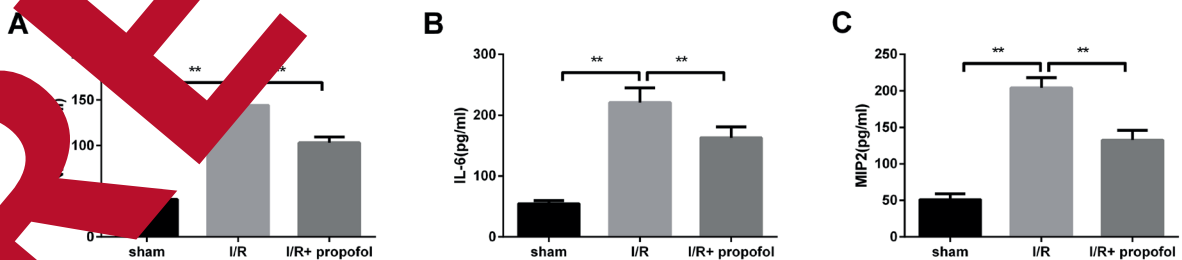


Figure 3. Effects of propofol on the pro-inflammatory factor expressions in the rats after hepatic ischemia-reperfusion (I/R). Serum in rats from the sham, I/R and I/R + propofol groups were collected. The release of (A) TNF- α , (B) IL-6 and (C) MIP2 were assessed by using ELISA kits. Data presented as mean \pm SD (n = 5/group). **, $p < 0.01$ (Student *t*-test).

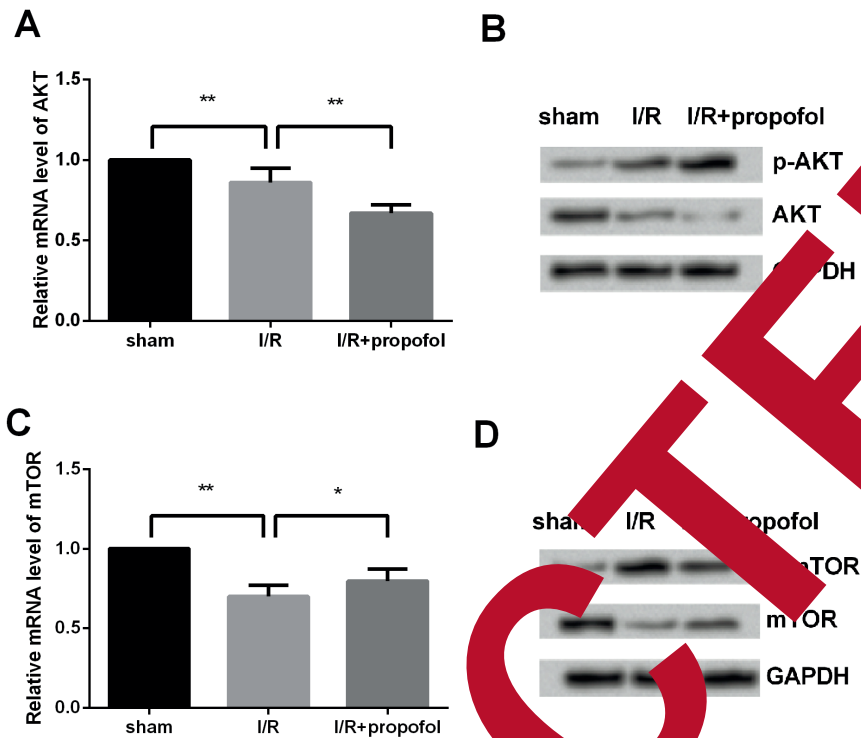


Figure 4. Effects of propofol on AKT and mTOR expression after hepatic ischemia-reperfusion (I/R). The (A) mRNA level of AKT, and the (B) protein levels of p-AKT and AKT were respectively detected by qRT-PCR and Western blotting. The (C) mRNA level of mTOR, and the (D) protein levels of p-mTOR and mTOR were respectively detected by qRT-PCR and Western blotting. Data presented as mean \pm SD (n = 5/group), $P < 0.05$, Student *t*-test).

AKT expression in hepatic I/R³²; this was also confirmed in this study that propofol significantly up-regulated p-AKT while it down-regulated AKT. However, this study provided the first evidence that propofol also could regulate mTOR expression in hepatic I/R. In the present paper, p-mTOR was down-regulated by propofol, while mTOR was up-regulated. Similarly, propofol was reported to decrease p-mTOR/mTOR level in cerebral I/R injury³⁴.

Conclusions

This study demonstrated that propofol preconditioning protects against hepatic I/R partly by reducing apoptosis and reducing the release of pro-inflammatory cytokines, which is possibly involved with the modulation of the PI3K/AKT/mTOR signaling pathway. Our study may provide a new method for clinical treatment of hepatic I/R injury and supply a molecular basis for the new use of anesthetic.

Acknowledgments

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Conflict of interest

The authors declare no conflicts of interest.

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