

Long noncoding RNA MIAT promotes the growth and metastasis of non-small cell lung cancer by upregulating TDP43

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Abstract. – OBJECTIVE: Recent researches have proved that long noncoding RNAs (lncRNAs) act and have an important role in many diseases. In this research, lncRNA MIAT was explored to identify how it functions in the development of non-small cell lung cancer (NSCLC).

PATIENTS AND METHODS: Real-time quantitative polymerase chain reaction (RT-qPCR) was utilized to detect MIAT expression in NSCLC patients. Next, we conducted cell counting kit-8 (CCK-8) assay, colony formation assay, ethynyl deoxyuridine (EdU) incorporation assay, wound healing assay and transwell assay to identify its biological function. Further experiments were performed to explore the potential mechanism.

RESULTS: By comparing with MIAT expression in adjacent tissues, MIAT expression level was significantly higher in NSCLC samples. Moreover, functional assays showed that cell growth ability of NSCLC cells was inhibited after MIAT was knocked down. In addition, the migrated and invaded ability of NSCLC cells was inhibited after MIAT was knocked down. Furthermore, the expression of TDP43 was downregulated by knockdown of MIAT. Meanwhile, it was found that TDP43 expression positively correlated to MIAT expression in NSCLC tissues.

CONCLUSIONS: Results also suggest that MIAT could enhance cell proliferation and metastasis of NSCLC by upregulating TDP43, which suggests that MIAT may be a potential therapeutic target in NSCLC.

Key Words:

Long noncoding RNA, MIAT, Non-small cell lung cancer, TDP43.

Introduction

Lung cancer is one of the most frequent malignancies in the world, which is also the lead-

ing cause of tumor-related deaths globally, accounting for almost 1.3 million deaths annually¹. Non-small cell lung cancer (NSCLC) accounts for 85% of lung cancer cases². Despite technology develops in the molecular biology, diagnosis and therapeutic treatment of NSCLC, most of NSCLC patients are unfortunately diagnosed at advanced stage without the opportunity to take curable surgery, which contributes to the poor survival rate³. Therefore, it is urgent to realize the underlying mechanism and find a new treatment strategy. Long non-coding RNAs (lncRNAs) are defined as non-protein-coding RNAs, which are longer than 200 nucleotides. Numerous studies have showed that lncRNAs are a new frontier in the research of malignant diseases. For instance, upregulation of PVT1 promotes proliferation, cell progression and metastasis of melanoma cells⁴. lncRNA-SNHG7 regulates proliferation, apoptosis and invasion of bladder cancer cells⁵. lncRNA MALAT1 accelerates cell migration and invasion via targeting miR-204 in hepatocellular carcinoma⁶. However, the function of lncRNA MIAT in NSCLC and the potential molecular mechanism haven't been studied so far. In this study, we found out that the expression of MIAT was remarkably higher in NSCLC tissues. Moreover, knockdown of MIAT inhibited the proliferation and metastasis of NSCLC cells. Moreover, our further experiment explored the underlying mechanism how MIAT functioned in NSCLC development.

Patients and Methods

Tissue Samples

A total of 56 NSCLC patients were obtained from NSCLC patients who underwent surgery at

Harbin Medical University Cancer Hospital. No radiotherapy or chemotherapy was performed before the surgery. All fresh tissues got from surgery were stored immediately at -80°C . This study was approved by the Ethics Committee of Harbin Medical University Cancer Hospital. Signed written informed consents were obtained from all participants before the study.

Cell Culture

Human NSCLC cell lines (A549, PC-9 and H358) and normal human bronchial epithelial cell line (16HBE) were obtained from American Type Culture Collection (ATCC) (Manassas, VA, USA). Culture medium consisted of 10% fetal bovine serum (FBS; Gibco, Rockville, MD, USA), Dulbecco's Modified Eagle's Medium (DMEM; Gibco, Rockville, MD, USA) as well as penicillin. Besides, cells were cultured in an incubator containing 5% CO_2 at 37°C .

Cell Transfection

The cDNA oligonucleotides specifically targeting MIAT (sh-MIAT) was synthesized by GenePharma (Shanghai, China) and inserted into the shRNA expression vector pCMV-neo. sh-MIAT was then used for transfection in NSCLC cells. 48 h later, Real-time quantitative polymerase chain reaction (RT-qPCR) was used to monitor the transfection efficiency.

RNA Extraction and RT-qPCR

TRIzol reagent (Invitrogen, Carlsbad, CA, USA) was utilized to separate total RNA. Through reverse Transcription Kit (Takara, Bio-technology Co., Ltd., Dalian, China), the total RNA was reverse-transcribed into complementary deoxyribose nucleic acids (cDNA). Following are the primers used for RT-qPCR: MIAT primers forward: 5'-GCGGTTTTCACAACCCTACTG-3', reverse: 5'-TCCACTTTGGCATTCTAGG-3'; GAPDH primers forward: 5'-CCAAAATCAGATGCCAACTGG-3' and reverse: 5'-TGATGGGCTGCTGCTGTCATTCA-3'. Thermal cycle was 30 s at 95°C , 5 s for 40 cycles at 95°C , 35 s at 60°C .

Cell Proliferation (CCK-8) Assay

Cell proliferation was measured by CCK8 assay (Dojindo Laboratories, Kumamoto, Japan), cell growth of transfected cells in 96-well plates was measured at 24, 48, and 72 hours. Spectrophotometer (Thermo Fisher Scientific,

Waltham, MA, USA) was used to measure the absorbance at 450 nm.

Colony Formation Assay

H358 cells were placed in 96-well plates for 10 days. Then, colonies were stained with 10% formaldehyde for 30 min and stained for 5 min with 0.5% crystal violet. The Image-Pro Plus 6.0 (Silver Spring, MD, USA) was used for data analysis.

Ethynyl Deoxyuridine Incorporation Assay

According to the manufacturer's manual, an EdU (EdU, Roche, Mannheim, Germany) was utilized to monitor the proliferation of transfected cells. Zeiss Axiophot Photomicroscope (Carl Zeiss, Oberkochen, Germany) was performed to capture the representative images.

Tube Formation Assay

Cells were cultured into 6-well plates were cultured in Dulbecco's Modified Eagle Medium (DMEM) medium overnight. After scratched with a plastic tip, cells were cultured in serum-free medium. Tube formation was viewed at 48 h. Each experiment was independently repeated in triplicate.

Transwell Assay

For detecting the migrating ability of these treated cells, 5×10^4 cells in 200 μL serum-free DMEM were transformed to the top chamber of an 8 μm pore size insert (Millipore, Billerica, MA, USA). For detecting the invading ability of these treated cells, 5×10^4 cells in 200 μL serum-free DMEM were transformed to top chamber of an 8 μm pore size insert (Millipore, Billerica, MA, USA) coated with 50 μg Matrigel (BD Biosciences, Franklin Lakes, NJ, USA). And the bottom chamber was added with DMEM and FBS. The top surface of chambers was wiped by cotton swab 48 h later and immersed for 10 min with precooling methanol. Then, they were stained in crystal violet for 30 min. Three fields were used to count the data for invasion membrane.

Statistical Analysis

Statistical analysis was conducted by Statistical Product and Service Solutions (SPSS) 17.0 (SPSS Inc., Chicago, IL, USA). Student *t*-test method was performed to the data. Data were presented as mean \pm SD (standard deviation). $p < 0.05$ was considered as statistically significant.

Results

MIAT Expression Level in NSCLC Tissues and Cells

Firstly, MIAT expression was detected *via* RT-qPCR in 56 NSCLC patients' tissues and 3 NSCLC cell lines. Results showed that MIAT was significantly upregulated in tumor tissue samples (Figure 1A). MIAT expression level of NSCLC cells was higher than that of 16HBE (Figure 1B).

Knockdown of MIAT Suppressed Cell Proliferation of NSCLC Cells

In this study, we chose A549 NSCLC cell line for the silence of MIAT. MIAT expression was detected by RT-qPCR (Figure 2A). Results of CCK8 assay showed that silence of MIAT inhibited cell growth ability of NSCLC cells (Figure 2B). The outcome of colony formation assay also revealed that the number of colonies was remarkably decreased after MIAT was silenced in NSCLC cells (Figure 2C). Moreover, results of EdU incorporation assay also revealed that the percentage of EdU positive cells reduced after silence of MIAT in A549 cells (Figure 2D).

Knockdown of MIAT Suppressed Cell Migration and Invasion of NSCLC Cells

To detect the function of MIAT in NSCLC metastasis, wound healing assay and transwell assay were performed. Results of wound healing assay showed that knockdown of MIAT inhibited cell migrated ability of NSCLC cells (Figure 3A). The outcome of transwell assay also revealed that the number of migrated and invaded cells was re-

markably decreased after MIAT was knocked-down in NSCLC cells (Figure 3B and 3C).

Interaction Between TDP43 and MIAT in NSCLC

Starbase v2.0 (<http://starbase.sysu.edu.cn/starbase2/rbpLncRNA.html>) was used to predict the target proteins of MIAT, among which TDP43 was selected for our following experiments. RT-qPCR results showed that the expression level of TDP43 in NSCLC tissues was remarkably lower in sh-MIAT group compared with that in control group (Figure 4A). Besides, we found that TDP43 expression in NSCLC cells was significantly higher when compared with 16HBE (Figure 4B). Furthermore, we found that TDP43 expression in NSCLC tissues was remarkably higher when compared with that of adjacent tissues (Figure 4C). Correlation analysis demonstrated that TDP43 expression level was positively correlated with MIAT expression in cancer tissues (Figure 4D).

Discussion

lncRNAs are important regulators in lung cancer initiation and progression. For example, miR-141 down-regulated FGF1 by overexpression of lncRNA H19 and lead to poor prognosis⁷. LncRNA PRNCR1 functions as an oncogene in NSCLC and promotes tumor progression through regulating miR-488/HEY2 signal network⁸. LncRNA MIAT, as a novel molecular, has been found to participate in numerous diseases, including the development of

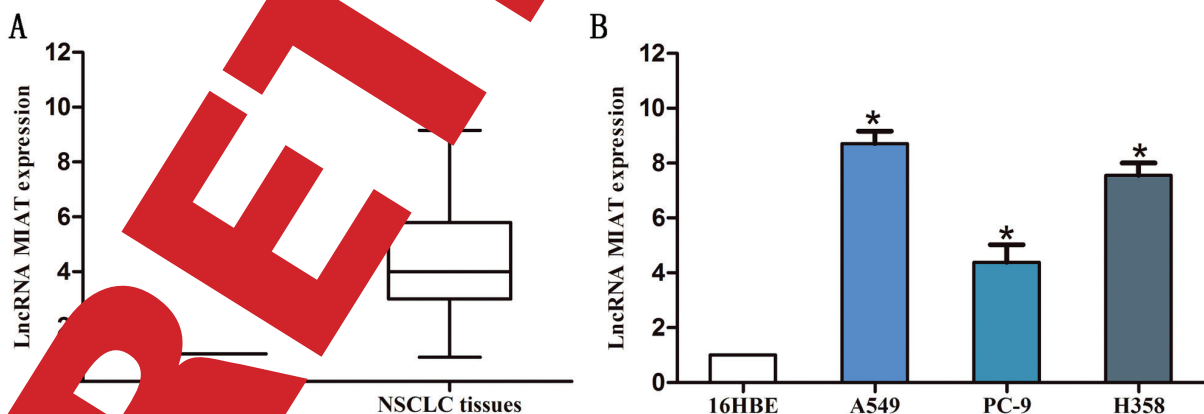


Figure 1 Expression levels of MIAT were increased in NSCLC tissues and cell lines. **A**, MIAT expression was significantly increased in NSCLC tissues compared with adjacent tissues. **B**, Expression levels of MIAT relative to GAPDH were determined in the human NSCLC cell lines and 16HBE (normal human bronchial epithelial cell) by RT-qPCR. Data are presented as the mean \pm standard error of the mean. * $p < 0.05$.

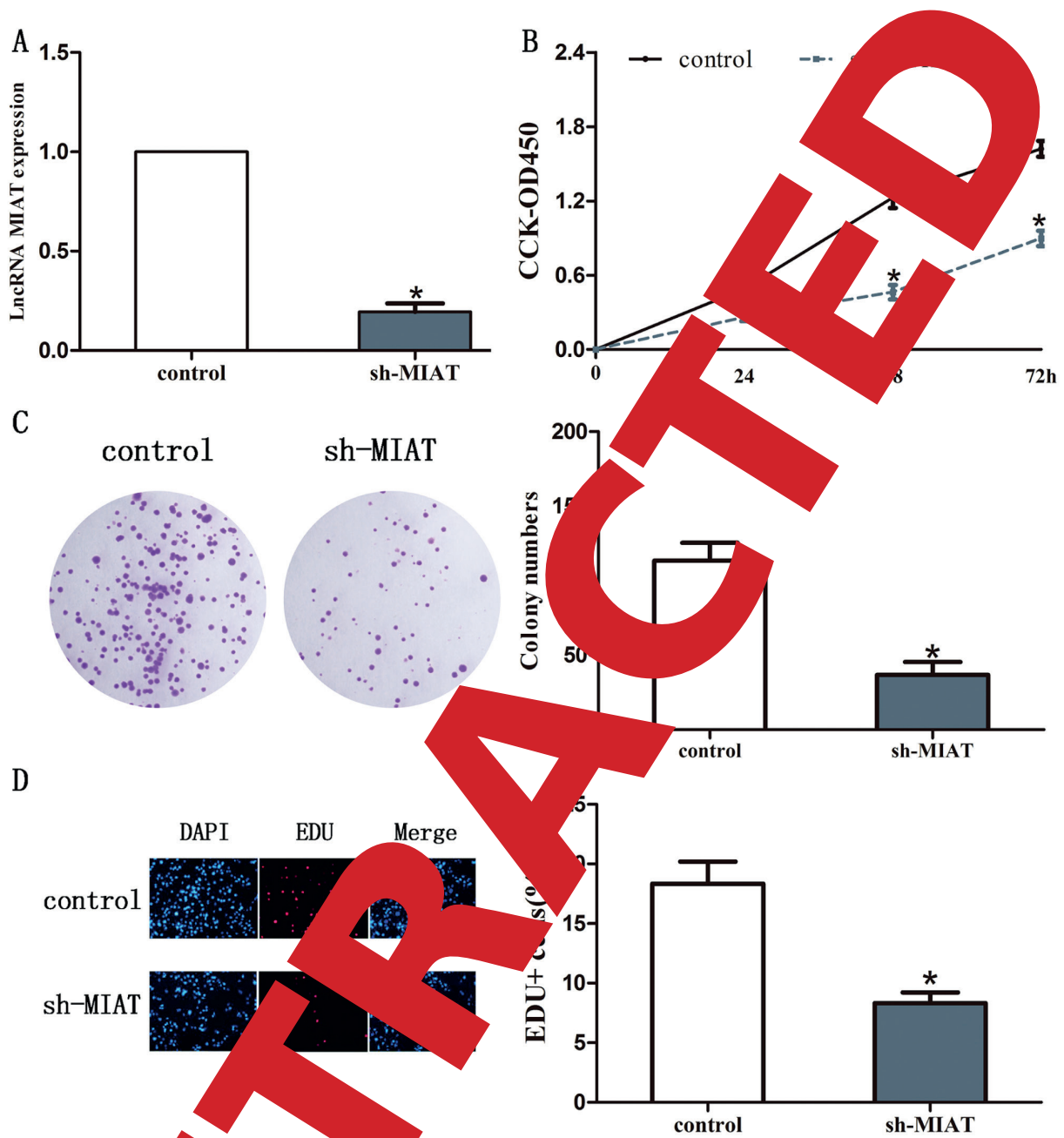


Figure 2. CCK8 assay showed knockdown of MIAT repressed NSCLC cell proliferation. **A**, MIAT expression in NSCLC cells transfected with sh-MIAT and control vector was detected by RT-qPCR. GAPDH was used as an internal control. **B**, CCK8 assay showed that knockdown of MIAT significantly repressed cell proliferation in NSCLC cells. **C**, Colony formation assay showed that number of colonies was significantly decreased *via* knockdown of MIAT in NSCLC cells (magnification: 40×). **D**, EdU incorporation assay showed that number of EdU positive cells was significantly decreased *via* knockdown of MIAT in NSCLC cells. Results represent the average of three independent experiments (mean ± standard error of the mean). * $p < 0.05$, as compared to control cells.

MIAT promotes the progression of breast cancer by serving as a ceRNA for miR-155-5p⁹. MIAT promotes colorectal cancer growth and tumor metastasis by targeting Derlin-1¹⁰. In this study, we found that MIAT was upregulated both

in NSCLC samples. Besides, silence of MIAT repressed cell proliferation of NSCLC cells. Above results indicated that MIAT promotes tumorigenesis of NSCLC and might act as an oncogene. To further identify the underlying mechanism of how

MIAT affects NSCLC, we used Starbase v2.0 to predict the target proteins of MIAT and picked TDP43 as the potential targets of MIAT. TDP43 is aberrantly expressed in breast cancer and is associated with patients' prognosis¹¹. TDP43 acts as an oncogene in melanoma and participates in

regulation of melanoma pathogenesis. Knockdown of TDP43 suppresses tumor progression via SRSF3 in triple-negative breast cancer¹³. TDP43 enhances cell proliferation and invasion via targeting HDAC6 in glioblastoma. A recent study¹⁵ reveals that TDP43/MALAT1 axis enhances

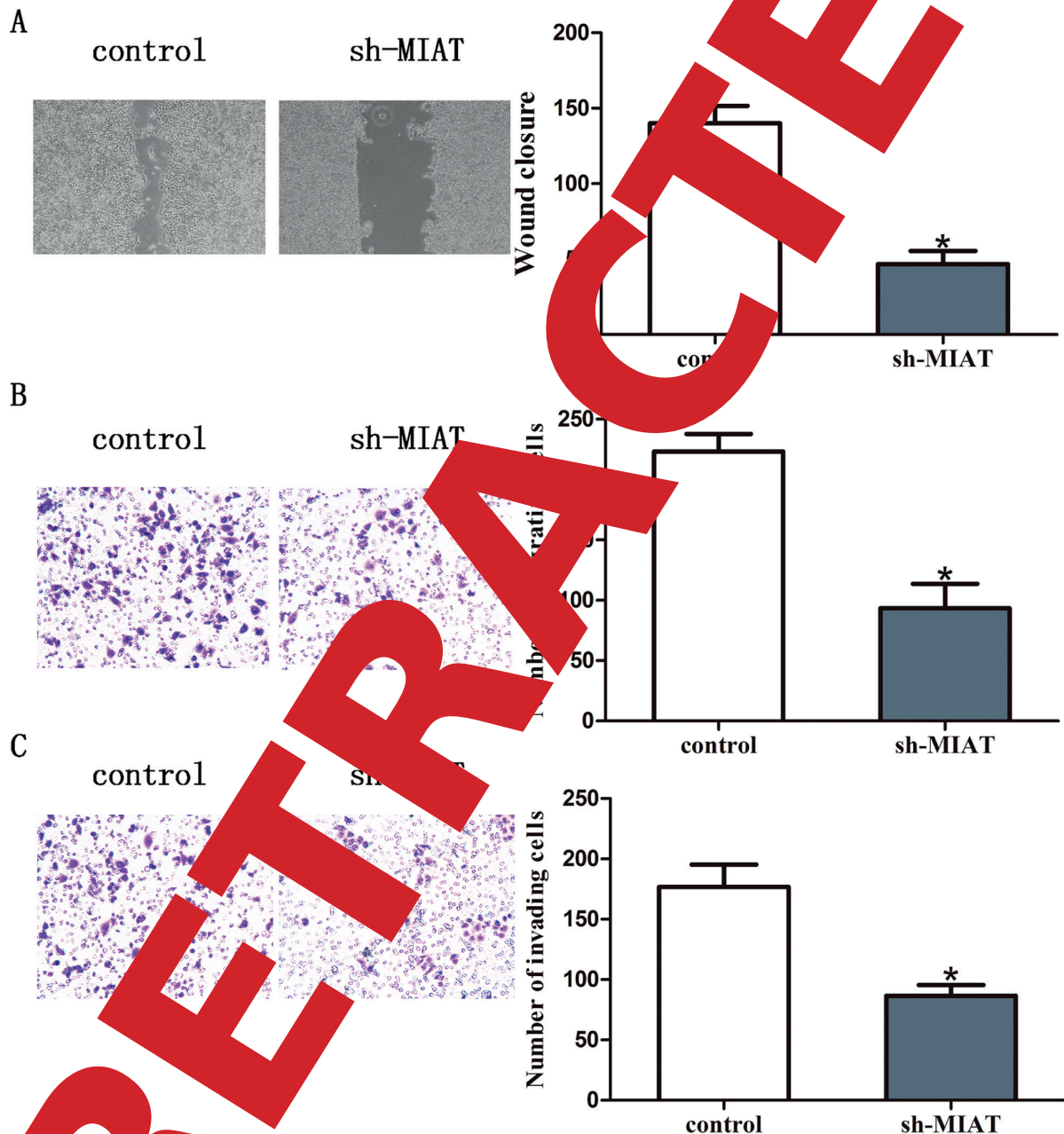


Figure 3. Wound healing assay and transwell assay showed that knockdown of MIAT inhibited NSCLC cell migration and invasion. **A**, Wound-healing assay showed that knockdown of MIAT significantly repressed cell migration in NSCLC cells (magnification: 40×). **B**, Transwell assay showed that number of migrated cells was significantly decreased via knockdown of MIAT in NSCLC cells (magnification: 40×). **C**, Transwell assay showed that number of invaded cells was significantly decreased via knockdown of MIAT in NSCLC cells (magnification: 40×). The results represent the average of three independent experiments (mean ± standard error of the mean). * $p < 0.05$, as compared with the control cells.

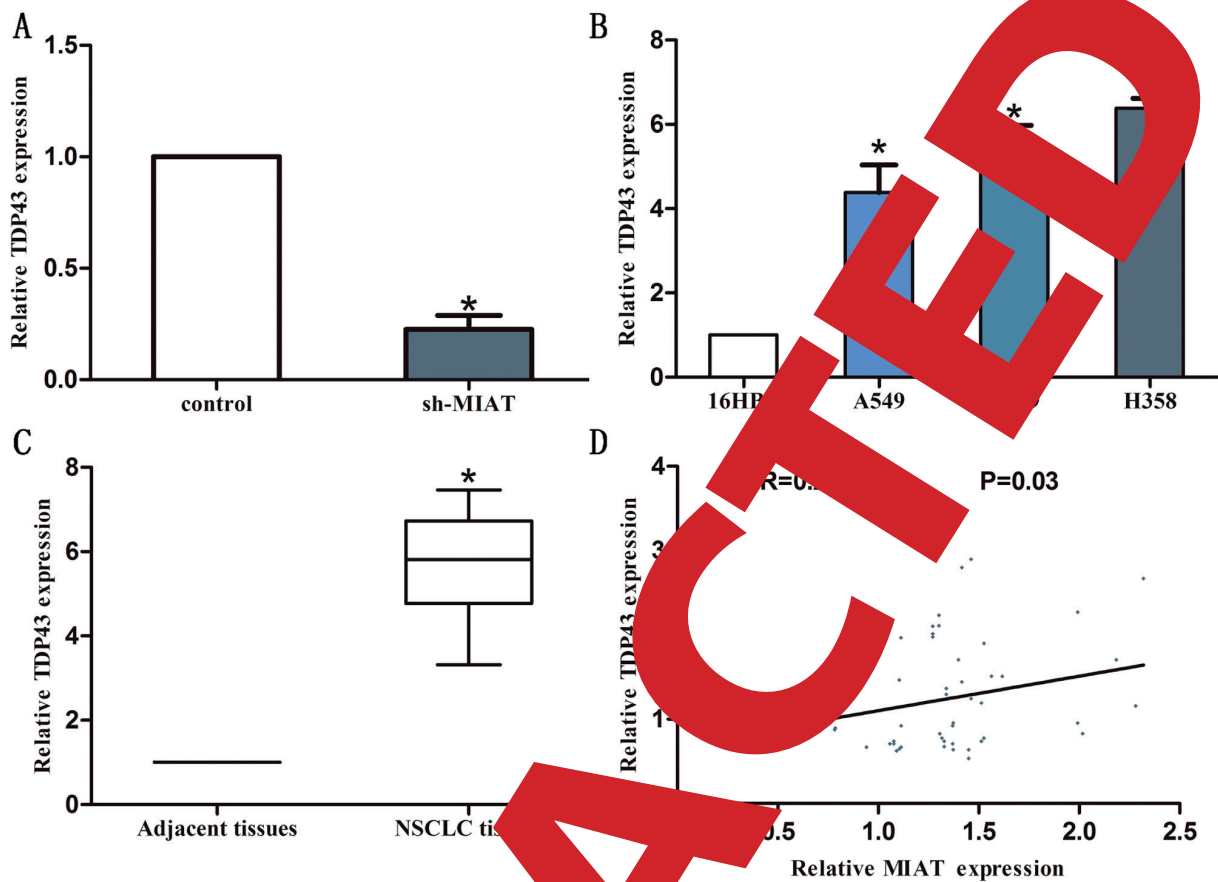


Figure 4. Interaction between MIAT and TDP43. **A**, The results showed that TDP43 expression was decreased in sh-MIAT compared with the control group. **B**, Expression level of TDP43 relative to GAPDH were determined in the human NSCLC cell lines and 16HBE by RT-qPCR. TDP43 was significantly upregulated in NSCLC tissues compared with adjacent tissues. **C**, The linear correlation between expression level of TDP43 and MIAT in NSCLC tissues. The results represent the average of three independent experiments. Data presented as the mean \pm standard error of the mean. * $p < 0.05$.

development and metastasis of NSCLC. In the present study, we first revealed the interaction between TDP43 and MIAT. TDP43 could be downregulated after knockdown of MIAT. Besides, TDP43 expression was higher in NSCLC cell lines. Furthermore, TDP43 expression in NSCLC tissues was positively related to MIAT expression. The results also indicated that MIAT might promote tumorigenesis of NSCLC through upregulating TDP43.

Conclusions

Our study demonstrated that MIAT could facilitate cell proliferation in NSCLC through upregulating TDP43, providing a candidate target for NSCLC treatment.

Conflict of Interest

The Authors declare that they have no conflict of interest.

References

- 1) TORRE LA, BRAY F, SIEGEL RL, FERLAY J, LORTET-TIEULENT J, JEMAL A. Global cancer statistics, 2012. *CA Cancer J Clin* 2015; 65: 87-108.
- 2) LIU Z, JIANG L, ZHANG G, LI S, JIANG X. MiR-24 promotes migration and invasion of non-small cell lung cancer by targeting ZNF367. *J BUON* 2018; 23: 1413-1419.
- 3) DUAN J, YANG Z, LIU D, SHI Y. Clinical efficacy of bevacizumab combined with gemcitabine and cisplatin combination chemotherapy in the treatment of advanced non-small cell lung cancer. *J BUON* 2018; 23: 1402-1406.

- 4) ZOU MF, LING J, WU OY, ZHANG CX. Long non-coding RNA PVT1 functions as an oncogene in ovarian cancer via upregulating SOX2. *Eur Rev Med Pharmacol Sci* 2018; 22: 7183-7188.
- 5) ZHONG X, LONG Z, WU S, XIAO M, HU W. LncRNA-SNHG7 regulates proliferation, apoptosis and invasion of bladder cancer cells assurance guidelines. *J BUON* 2018; 23: 776-781.
- 6) HOU Z, XU X, ZHOU L, FU X, TAO S, ZHOU J, TAN D, LIU S. The long non-coding RNA MALAT1 promotes the migration and invasion of hepatocellular carcinoma by sponging miR-204 and releasing SIRT1. *Tumour Biol* 2017; 39: 1393371529.
- 7) WU D, YANG B, CHEN J, XIONG H, LI Y, PAN Z, CAO Y, CHEN J, LI T, ZHOU S, LING X, WEI Y, LI G, ZHOU Y, QIU F, YANG L, LU J. Upregulation of long non-coding RNA RAB1A-2 induces FGF1 expression worsening lung cancer prognosis. *Cancer Lett* 2018; 438: 116-125.
- 8) CHENG D, BAO C, ZHANG X, LIN X, HUANG H, ZHAO L. LncRNA PRNCR1 interacts with HEY2 to abolish miR-448-mediated growth inhibition in non-small cell lung cancer. *Biomed Pharmacother* 2018; 107: 1540-1547.
- 9) LUAN T, ZHANG X, WANG S, SONG Y, ZHOU S, LIN J, AN W, YUAN W, YANG Y, CAI H, ZHANG Q, WANG L. Long non-coding RNA MIAT promotes breast cancer progression and functions as ceRNA to repress DUSP7 expression by sponging miR-155-5p. *Cell Biosci* 2017; 8: 76153-76164.
- 10) LIU Z, WANG H, CAI H, HONG Y, LI Y, SU D, FANG Y. Long non-coding RNA MIAT promotes growth and metastasis of colorectal cancer cells through regulation of miR-132/Derb1. *Int J Cancer* 2018; 123: 123-128.
- 11) KIM PY, TAN O, LIU B, TRAVIS L, LIU T, HABER M, MORRIS MD, MARSHALL GM, HARRIS BB. High TDP43 expression is required to maintain and induce inhibition of cancer cell growth in neuroblastoma with good prognosis. *Cancer Lett* 2016; 374: 311-323.
- 12) ZENG Q, CAO J, LI R, HUANG J, XIA K, TANG J, CHEN X, ZHOU M, XIE Y. Identification of TDP-43 as an oncogene in melanoma and its function during melanoma pathogenesis. *Cell Biosci* 2017; 8: 1-10.
- 13) KE H, ZHANG H, FENG X, XU H, HAO J, WANG S, LIU X, SU X, WANG L, WU C, WANG Y, NIU J, JIAO B. TDP43 inhibits progression of triple-negative breast cancer in coordination with SRSF3. *Proc Natl Acad Sci U S A* 2018; 115: E3426-E3435.
- 14) LIN TW, CHEN Y, LIN LT, HUANG PI, LO WL, YANG YP, LU KH, CHEN W, CHIOU SH, WU CW. TDP-43/MDAC6 axis promoted tumor progression and metastasis of neuroblastoma. *Oncotarget* 2017; 8: 56612-56625.
- 15) GUO F, JIAO F, SONG Z, LI S, LIU B, YANG H, ZHOU Q. TDP43 regulates the migration and invasion of non-small cell lung cancer cells *in vitro*. *Biochem Biophys Res Commun* 2015; 465: 293-298.