Clinical significance of PSMA, TERT and PDEF in malignant tumors of the prostate

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Abstract. – OBJECTIVE: To investigate the clinical significance of the expression of PS-MA (prostate specific membrane antigen), TERT (telomerase reverse transcriptase), and PDEF (prostate derived Ets factor) in malignant tumors of the prostate.

PATIENTS AND METHODS: The study was conducted with paraffin slices from 33 specimens of malignant tumors of the prostate and 17 of normal tissue. We found high levels of PSMA, TERT, and PDEF protein by Western blot and immunofluorescence in the malignant tumor prostate. We also detected upregulation MA, TERT, and PDEF mRNA in the malignant tumor of the prostate, suggesting complex nontion of these three genes in prostate cancer.

RESULTS: Variance analysis showed stat cally significant differences ca ng the e pression of PSMA, TERT, and e malig nant tumor of the prostate d nor tissues. A, TERT The high expression of **B** d PDEF in the malignant tumor rost the important roles of thes occurrence and dev alignant tument mors of the prost

CONCLUSION MA, TERT, and F may serve as a reference on linical diagnous and as potential targets for the lignant tumor of the prostate the rapeutics.

Key Wo PSN

tate

TERT, PDEF, Malignant tumor of the proscession designs.

Juction

I prise in modolity and mortality of malignary stores of the prostate in recent years make a ying effective markers that enable 'v and accurate diagnosis and intervention^{1,2}. The recent efforts, the pathogenesis of malight tumors of the prostate is still unclear, and effective treatments are no available³. Here, we deter the ssion levels for three relevant tumor marker. MA (Prostate Specific s Antigen), 🔪 Mem Telomerase Reverse ase) and PDEF rostate Derived Ets tor) – by different techniques in malignant nors of the pr te and normal prostate tissue. high expres of PSMA, TERT, and PDEF F malign tumor of the prostate at the ii A levels suggests their importpro ant roles development of malignant tumors the prostate. This information can be relevant cal diagnosis and treatment of prostate

Patients and Methods

Collection of Tumors

ΩŪ.

We collected 33 specimens of malignant tumors of the prostate and 17 of normal prostate tissue in our hospital. They were fixed in neutral formalin (10%) for over 48 h, embedded in paraffin, and cut into the 5 μ m slices. The Ethic Committee of the Third Affiliated Hospital approved this study.

Immunofluorescence

The tissue slices were de-waxed by dimethylbenzene several times and then dehydrated by ethanol gradient for antigen retrieval. After the slices were washed 3 times (5 min/time) with PBS (pH = 7.4), they were blocked in 10% bovine serum albumin (BSA) in a humidity chamber at 37° C for 30 min. Then, the slices were washed in phosphate-buffered saline (PBS) (pH = 7.4) once to remove excess of bovine serum albumin (BSA). Then, the primary antibody (1:70) was added into the slices and incubated in the humidity chamber at 4°C overnight. The next day, we washed the slices three times (5 min/time) with PBS and added the fluorescent secondary antibody in the dark and incubated in the humidity chamber at 37°C for 2 h. After the slices were washed with the secondary antibody, they were sealed in glycerol for observation under the fluorescence microscope. mAb anti- β -Actin, anti-PS-MA, anti-TERT, and anti-PDEF, as well as fluorescence secondary antibody, were obtained from Cell Signaling Technology (Danvers, MA, USA).

RT-PCR Detection

Two samples (50 mg) from the malignant tumor of the prostate and normal tissues were rapidly transferred to 1 mL of Trizol (Tiangen Biotech, Beijing, China) and homogenized. After 5 min at the room temperature, the homogenate was centrifuged at 12,000 \times g and 4°C for 15 min. The supernatant was mixed with chloroform. After 5 min at the room temperature, the mixture was centrifuged at $12,000 \times g$ at 4°C for 15 min. Then, the same volume of isopropanol was added to the mixture. After 10 min at the room temperature, the mixture was centrifuged at $12,000 \times g$ and $4^{\circ}C$ for 10 min and the was collected. We mixed the pellet with (75%) to wash the RNA. Then, it was d red in RNase-free water. The concentration of and the ratio of OD_{260}/OD_{280} were determined Finally, the mRNA for PSMA, TEPT, and PD were amplified using the prim in Tabl I. The products were used RT-P analysis (Tiangen Biotech, Beijing (ina).

Western Blot

Two tissue sample 50 mg) we have acted from the malignant turn of the prostal mormal tissues, washer a more hysiological some, and homogenized with lysis more. Then, the homogenates were sentrifuged as $90 \times \text{g}$ at 4°C for



10 min, and the supernatant was centrifuged again under the same conditions, and then we the supernatant. The protein concern assayed by the BCA protein assay Beyuntia Beijing, China) and the samples re preserved at -80°C. The extracts were min th 2X loading buffer (100 μ l + 4 μ l-mercap ol) at a volume ratio of 1:1, the d in ture wa boiled water for 5 min, then preserve refrigerator. The separation on gel and 5% stat gel were prepared ding the molecular he hom weight of the target nates e eleg horesis were loaded op ne gel, of 220 V was perform nder a const. until the br le bottom of nol blue reach in cut into pieces according the gel. 7 gel to the molecular we the target proteins and The polyvinylidene place to transfer by VDF) membrane vas cut into the shape f the gel and was placed into the transfer buffer er being soak n methanol for 10 s. Next, the was performed at a constant brane trans of 100 The PVDF membrane was In milk powder for 3 h at room bloc temperature on the shaker. The membranes were ubated with the primary antibodies at 4°C overr three washes in TTBS (10 min/time), orane was incubated with the secondary antibody on the shaker at a room temperature for 2 h and washed three times in TTBS (10 min/time). The mixed solution of reagents A and B in the electrochemiluminescence (ECL) assay kit were added onto the PVDF membranes for complete reaction. After 1 min of color development in the dark, the mixture was put into the gel imager and photographed using dynamic calculus pattern, and

Statistical Analysis

strips using Gel-Pro 4.0.

SPSS 18.0 (SPSS Inc., Chicago, IL, USA) was used for the statistical analysis of results. The experimental data were presented as mean \pm standard deviation (Mean \pm SD). *t*-test, *p*-test, and One-Way ANOVA variance analysis were performed in-group comparisons. *p* < 0.05 suggested that the differences were statistically significant.

the gray analysis was performed on the protein

Results

Pathological Slices

The glands were in back to back arrangement, and the original basal cells significantly disap-



Figure 1. Pathological results of the normal prostatic tissues and the malignant tumor of the patic tissues.

peared in the malignant tumor of the prostatic tissues (Figure 1). Compared with the slices of normal prostatic tissues, gradient vacuolization of nuclei and significantly infiltrated interstitial were identified in the malignant tumor of the prostatic tissues (Figure 1). Further we found that the nuclei were mutated glands were distributed irregularly, insult of the original arrangement.

Immunofluorescence of PSMA_TERT, and PDEF in Prostatic Tis

Fluorescent expression of SMA, PDEF was identified in f malignar RT, and umor of prostatic tissues, but not in the normal tises (Figure 2), and ditionally, PSMA, TERT, and R were high expressed in the malignant tumen 5 the proster e tissues. The result indicated the second second second prostate of the prostate.

evels of PSMA, TERT, and Prostatic Tissue

To examine the protein expression of PSMA, TERT, and PDEF by a more quantitative method, we conducted Western blot analysis. The protein levels of PSMA, TERT, and PDEF were highly elevated in the malignant tumor of



Figure 2. Expression of PSMA, TERT and PDEF in the normal tissues and the malignant tumor of the prostatic tissues (×400).

the prostatic tissues, but very lowly expressed in the normal tissues (Figure 3). Each protein was elevated more than ten times in prostate tumors compared to normal prostatic tissue, supporting the strong signal observed by immunofluorescence.

mRNA Levels of PSMA, TERT, and PDEF in Prostatic Tissue

After observing the marked upregulation of PSMA, TERT, and PDEF at the protein level, we decided to analyze the expression of PSMA, TERT, and PDEF mRNA to understand how these genes are regulated. PSMA, TERT, and PDEF mRNA were significantly increased in malignant tumor of the prostatic tissues compared to the normal prostate (Figure 4). PSMA showed the highest upregulation of the three genes analyzed (Figure 4). These observations are consistent with the elevated levels of PSMA, TERT, and PDEF at the protein level and suggest that both transcriptional and posttranscriptional mechanisms are involved in the upregulation of the prostate.

Discussion

Prostatic tumors are the most common lignant tumors in males and the second cause male cancer deaths⁴. Because the prostath tumors have become an interactant with issue as the male population in times to ge⁵. Un-

fortunately, the molecular mechanisms leading to the prostatic tumor and the develop malignant phenotype are mostly unl uncertain disease course, as well, ne hetero research to geneity of the disease, hinders better understand the occurr f prostatic tumor. Also, this situation comp the research with clinical spec hs or su the progression of the proic tumor ove years, the incident tain period7-9. In rec prostatic diseases crea continuously, and their progression alignant mors makes them a Older ious h once the largest of prosmales repres h are usual companied tatic tume naturia and dysuria¹¹. The by symp ıs h diagnosis of prosta. por mainly depends on prostatic bio imag nd the pathological on of specimen, obtained during sure y¹²⁻¹⁴. In recent years, most studies showed t PSMA, T and PDEF play important in the de pment of prostatic tumors. TERT, PDEF are expressed at low ssues, but are upregulated in leve tumors, ruging prostatic tumor^{15,16}. PSMA serve as a therapeutic target for the immutreatment of dendritic cells, and the of PSMA gene can also be used as the ðh. cytotoxic reagent mediated into the prostatic tumor cells¹⁷. Normal cells transfected with TERT

mor cells¹⁷. Normal cells transfected with TERT can be immortalized without any transformation in the phenotype of nuclei. TERT can maintain the length of telomeres and is believed to be



Figure 3. Expression of PSMA, TERT and PDEF protein in the malignant tumor of the prostatic tissues and normal tissues.

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8)



Figure 4. mRNA expression of PSMA, TERT and PDEF in the malignant tumor of the prostatic tissues and normal

associated with the aging process and tumors. The TERT expression can only be detected in cells with telomerase activity, but cannot be detected in most benign tissues¹⁸. TER endow immortality to cells, which is, step in the transformation of malignant rs. Thus, infiltrated tumor cells can be de with high sensitivity using TERT from be tissues. The role of PDEF is very import in hematopoiesis, angiogene genesi and neural connection¹⁹. N al di ntiation of epithelial cells in the state and e initial stages of the prostatic it re d the testosterone, but e fil 01 proic tumor becomes dependent ost horm and resistant to one therapies

Conclu

ady showed high expression of PSMA, Ou TE and PD in the prostatic malignant tuthe im rtant roles of PSMA, mor, EF in t development of the ma-TERT, h ostate. Therefore, PSMA, nt tun in serve as potential targets and PL for treatment a malignant tumor of the prosthe new reference for clinical diagnotat nt.

Co. of Interest

The Authors declare that they have no conflict of interests.

References

- KWAK JT, HONG CW, PINTO PA, WILLI KRUECKER J. Is visual registration advalent semiautomated registration in the tate biopsy? Biomed Res Int 2015; 2015: 32 2.
- FELDMAN BJ, FELDMAN D. The descent of androgen-independent promate can be at Rev Cancer 2001; 1: 34-45.
- WU W, LIU X, CHAFTAR 3) ruz Carreras Mi UPCHURC Assoc LEZ C, GONZALEZ C, J, Merriman h of body com-SM, DALAL S, YEU theraposition with outco axel che cer: a re py in metast pective pros review. PL ne 2015; 220
- HAYWARD CUNHA GR. The context development and provide logy. Radiol Clin. orth Am 2000; 38: 1

5) HAYWARD SW, BASING CHAUGHNEY PC, FOSTER SBA, AR, DAHIYA R, DEGS, CUNHA GR. Stroevelopment in the unitral prostate, anterior prostate and seminal vesicle of the rat. Acta Anat (Basel) 1996; 155: 94-103.

Wang Y, Hayy, S, Cao M, Thayer K, Cunha G. Cell Nifferentiation page in the prostate. Differentia-2001; 68 0e9.

AT, TEXENA MR. Detailed analysis of expression and promoter methylation status of apoptosis-regenes in prostate cancer. Apoptosis 2010; 6-965.

- ABATE-SHEN C, SHEN MM. Molecular genetics of prostate cancer. Genes Dev 2000; 14: 2410e34.
- TSUJIMURA A, KOIKAWA Y, SALM S, TAKAO T, COETZEE S, MOSCATELLI D, SHAPIRO E, LEPOR H, SUN TT, WILSON EL. Proximal location of mouse prostate epithelial stem cells: a model of prostatic homeostasis. J Cell Biol 2002; 157: 1257-1265.
- CHOL N, ZHANG B, ZHANG L, ITTMANN M, XIN L. Adult murine prostate basal and luminal cells are self-sustained lineages that can both serve as targets for prostate cancer initiation. Cancer Cell 2012; 21: 253-265.
- 11) TAYLOR BS, SCHULTZ N, HIERONYMUS H, GOPALAN A, XIAO Y, CARVER BS, ARORA VK, KAUSHIK P, CERAMI E, REVA B, ANTIPIN Y, MITSIADES N, LANDERS T, DOLGALEV I, MAJOR JE, WILSON M, SOCCI ND, LASH AE, HEGUY A, EASTHAM JA, SCHER HI, REUTER VE, SCARDINO PT, SANDER C, SAWYERS CL, GERALD WL. Integrative genomic profiling of human prostate cancer. Cancer Cell 2010; 18: 11-22.
- 12) Ma X, ZIEL-VAN DER MADE AC, AUTAR B, VAN DER KORPUT HA, VERMEIJ M, VAN DUIJN P, CLEUTJENS KB, DE KRIJGER R, KRIMPENFORT P, BERNS A, VAN DER KWAST TH, TRAP-MAN J. Targeted biallelic inactivation of Pten in the mouse prostate leads to prostate cancer accompanied by increased epithelial cell proliferation but not by reduced apoptosis. Cancer Res 2005; 65: 5730-5739.
- LEONG KG, GAO WO. The Notch pathway in prostate development and cancer. Differentiation 2008; 76: 699-716.

- 14) VAN LEENDERS GJ, SCHALKEN JA. Epithelial cell differentiation in the human prostate epithelium: implications for the pathogenesis and therapy of prostate cancer. Crit Rev Oncol Hematol 2003; 46 (Suppl): S3-10.
- 15) VERHAGEN PC, VAN DUJIN PW, HERMANS KG, LOOIJENGA LH, VAN GURP RJ, STOOP H, VAN DER KWAST TH, TRAP-MAN J. The PTEN gene in locally progressive prostate cancer is preferentially inactivated by bi-allelic gene deletion. J Pathol 2006; 208: 699-707.
- FERREIRA R, OHNEDA KAND, YAMAMOTO M. GATA1 function, aparadigm for transcripti on factors in hematopoiesis. Mol Cell Biol 2005; 25: 1215-1227.
- COKIC VP, SMITH RD, BELESLIN-COKIC BB. Hydroxyurea induces fetal hemoglobin by the nitricoxide-dependent activation of soluble guanylylcyclase. J Clin Invest 2003; 111: 231-239.

- 18) DAVIES JH, EVANS BA, JENNEY ME. Effects of chemotherapeutic agents on the function of primery human osteoblast-like cells derived from Clin Endocrinol Metab 2003; 88: 67 - 2097.
- RHIM JS. In vitro human cell culture nodels for the study of prostate cancer. Protatic Dis 2000; 3: 229-235.
- 20) KARAN D, SCHMIED BM, DIVE BJ, STATU VA, LIN MF, BATRA SK. Decrease androgen assive growth of human processe cancer is as with increased generaliterations. Clin Control Res 2001; 7: 3472 - 30.
- 21) ALBERTI C. Prostate oth micro IA aberrant expre nd raautopha n-me diation the resistan sh ross-reference t igenetic/gene ism implisci 2016; 20: cations Med Pharma 1001

