Lycopene prevents experimental priapism against oxidative and nitrosative damage

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Abstract. - OBJECTIVE: Priapism is a persistent and often painful penile erection in the absence of sexual stimulation. It can cause progressive fibrosis, edema and drying of the erectile tissue and then it can lead to erectile dysfunction. Previous studies suggested that, neuronal nitric oxide levels increased during the priapism. High NO levels can result in the formation of reactive oxygen species (ROS) leading to oxidative stress in tissue and reproductive system. The aim of this study was to evaluate oxidative and nitrosative effects caused by pri in cavernosal tissue and serum, and deter a beneficial effects of lycopene on ischem apism.

MATERIALS AND METHODS: 32 rats v randomly divided into four gra d the f group being as the control. d grou sm was experimental ischemic pri med fo an hour and then 1hour s providusion ed. In the third grou lyc toneally given at the dose o ig/kg-.ne e were adh ered to rats fourth group, lyc with experimer ism. used a significant in-ituric acid reactive **RESULTS:** apis BARS (this crease in

d a significant de-) and NO leve substap the levels of GSH. T, GPx and SOD creas vernosal tissue of rats. However, m and g in sir cantly increased GSH, CAT, GPx lyco els but and S creased formation of RS p tion NO in rats with priapism. ur findings indicated that is-NCLU lead to significant oxidative ic pria cľ itrosative damage in cavernosal tissue and es of rats. However lycopene treatates these negative effects induced ient . priapism. For this reason, we suggested that ene may be used in the treatment of pria .m.

Key Words:

Priapism, Lycopene, Oxidative stress, Nitrosative stress, Cavernosal tissue.

troducti

Priapismus a p ent and often painful penile erection in the ce of sexual stimulaough it has h occurred in domestio nimals, it is a problem for human^{3,4}. The etiti y of priapismelated to use of some pharma-0 such as antipsychotics and ^{2,5}. Also it can be associated С tical agen onal dru rec cal disorders, metabolic disorwith lers, trauma, tumors, neurological disorders and the spider and scorpion^{2,6}. Firstly, priin cause progressive fibrosis, edema and drying of the erectile tissue and then it can lead to erectile dysfunction². Previous studies suggested that neuronal nitric oxide (NO) levels increased during the priapism4,8. Also, it was demonstrated that abnormally high NO levels can result in the formation of reactive oxygen species (ROS) leading to oxidative stress in tissue and reproductive system. This situation may cause severe infertility after priapism treatment. Therefore, it follows that antioxidant therapy may prevent side effects of priapism in the reproductive system.

Lycopene is a main carotenoid and is the most potent antioxidant among the various common carotenoids⁹. It is synthesized by microorganisms and plants red fruit and vegetables including tomatoes, watermelons, pink-grapefruits, apricots, papaya, guava and rosehip^{10,11}. Humans and animals do not synthesize lycopene and, thus, depend on dietary sources mainly tomatoes¹². Recently, many studies reported the beneficial effects of lycopene against cancer such as prostate and toxicities of antineoplastic agent (e.g. nephrotoxicity, hepatotoxicity) due to its highly efficient antioxidant and free radical scavenger properties¹³⁻¹⁵. The antioxidant activity of lycopene is assayed by inhibition

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of thiobarbituric acid reactive substances (TBARS) formation and induction of antioxidant defense system¹⁶. Atessahin et al¹⁷ determined that lycopene prevents oxidative damage in testis tissue of rat. Similarly, many authors^{12,13} claimed that prevention of oxidative stress by lycopene results in chemotherapeutic effect against many cancer types such as prostate. Since oxidative stress is one of the important results of priapism, it is, therefore, reasonable to consider that lycopene may play a role in the cellular defenses against oxidative stress induced by priapism.

Thus, in the current study, we aimed to determine of oxidative and nitrosative damage caused by ischemic priapism in serum and covernosal tissue and to find out whether or not lycopene has any beneficial effect against priapism in adult rats.

Materials and Methods

Chemicals

Lycopene 10% FS (Redivivo TM, Code 7803) was obtained from DSM Nutritional Products (Istanbul, Turkey). (Warwickshire, UK). All chemicals were purchased from Sigma Cleve Co. (St. Louis, MO, USA).

Animals

Exp

Thirty two healthy adult mal -Dawl rats (between 2-3 months) 00 g. and 2weight) obtained from the perime l Animal Institute, Malatya-Tu key his Animals were kept *erilize* propy rat cages, in 12-h dark cycle n ambient or them temperature Niet and w periments were perwere given ad libitu formed b d on Animal s Guidelines of Instituti Animals Ethics ittee.

er Protocol

performed under sterile All ions we ar als were anesthetized with lition ne (1 here p.) and ketamine injection (50 g, ip) duing the ishcemic priapism and m period. Priapism was accomplished dified method described by Uluocak et 1th un and Sanli et al¹⁹. Briefly, a 50-cc syringe was for the vacuum erection device and before application of vacuum to the penis, a constriction band, which was cut from 16 Fr Foley catheter in 2 mm slices, was loaded around the tip of the vacuum erection device. Then, the tip of the syringe was placed to the base of the penis and withdrawn

gently to induce erection in the rat penis. After induction of erection in sufficient grade, the constriction band was placed to the base of the penis by slipping off the syringe (Figure 1).

The animals were divided into four groups with each containing eight rats. The first was kept as the control group and give ny un group, istilled water as carrier. In the second chemic priapism was formed durin, ur. Then, the band was removed from the base penis and the penile tissue was all d to rep for one hour. In the third gro lycopene was kg) to r ed intraperitonally (10 s without apism. In the fourth gro ne was administered at 30th m of pria period then reperfusion y provided for in this ine animals group. End erfusion pe ernosal tissues were imwere sacrified an mediately removed a issected over ice-cold nomogeniza f cavernosal tissues gla carried out in a teflong ass homogenizer with mM KCl (pV7.4) to obtain 1: 10 (w/v) diluhomogenate. Blood samples ti of the who llected f h the left ventricle with an inwe sthesia. Sera were obtained by jector entrifugation (3000 g, 20 minutes, at 4°C) of blood. Tissue and serum samples were -50°C in a deepfreeze until analysis.

Biochemical Assay

The levels of homogenized tissue and serum TBARS, as an index of lipid peroxidation, were determined by thiobarbituric acid reaction using the method of Yagi²⁰. The product was evaluated spectrophotometrically at 532 nm and results are expressed as nmol/g tissue and nmol/ml. The reduced



Figure 1. Formed experimental priapism models in rat by syringe.

glutathione (GSH) levels content of the testis homogenate was measured at 412 nm using the method of Sedlak and Lindsay²¹. The GSH level was expressed as nmol/ml. SOD (superoxide dismutase) activity was measured by the inhibition of nitroblue tetrazolium (NBT) reduction due to O₂generated by the xanthine/xanthine oxidase system²². One unit of CuZn-SOD activity was defined as the amount of protein causing 50% inhibition of the NBT reduction rate. The product was evaluated spectrophotometrically at 560 nm. Results are expressed as IU/mg protein. Catalase (CAT) activity of tissues was determined according to the method of Aebi²³. The enzymatic decomposition of H_2O_2 was followed directly by the decrease in absorbance at 240 nm. The difference in absorbance per unit time was used as a measure of CAT activity. Tissue protein content was determined according to the method developed by Lowry et al using bovine serum albumin as standard²⁴.

Determination of NO

Serum nitric oxide (NO) level was determined by the evaluation of its stable oxidation products; nitrite (NO $_{2}$), and nitrate (NO $_{3}$). Serum levels were measured by the Griess rea Reduction of nitrate to nitrite was accomp he by catalytic reaction using cadmium. The produced was determined by diazotization of fanilamide and coupling to na ylene amine. Absorbance of this mple as mea 1 curve as estabsured at 545 nm. A star lished with a set of second a trite. Linear regres ne was oy peak area from the trite standa e resulting equation was inknown to calculate sample concentration ults were expressed as er serum²⁶. moles pe

Sta ical Aralysis

1 gr of significance was set at p < 0.001. vay analys of variance (ANOVA)

and post hoc Tukey-HSD test were used to determine the differences among the groups. All the analyses were carried out using the SPSS/PC (Version 11.5; SPSS, Chicago, IL, USA) software package program. Data are presented as mean±standard error of means (SEM).

Results

The cavernosal tissue S GSH, and TBARS levels are given Table I. SOL and GSH levels were nificar v decrea whereas TBARS level nificantly inpared. creased in rats w priapis control and other ps. There nc nificant ene group. changes bet control and vcopene+p.napism group On the other hand SOD, CAT, GSH IN were increased, and TB vels were de sed compared with tł of priapism group. When given together with brought SOD, CAT, GSH and pism, lycoper Т RS levels cl r to the control level. SOD, a ities and TBARS, GSH levels e given in Table II. In priapism in set roup, a significant decrease in SOD activities H level was observed in the serum samats compared with that of control and other groups. However, the TBARS and NO levels were significantly increased in the priapism group compared to other groups. In the lycopene+priapism group, TBARS, NO and GSH levels were near to control group value, but CAT and SOD activities remained unchanged com-

Discussion

pared with priapism group.

Oxidative and/or nitrosative stress has an important role in the pathogenesis of ischemic priapism. It is largely known that the generation and

1. The SOD and CAT activities and GSH and TBARS levels in cavernosal tissue of rats.

	Control	Lycopene	Priapism	Priapism+ Lycopene	<i>p</i> value
ARS µmol/ml GSH nmol/ml SOD U/mg protein CAT U/mg protein	$\begin{array}{c} 3.92 \pm 0.32^a \\ 109.5 \pm 3.21^a \\ 16.24 \pm 0.51^a \\ 0.053 \pm 0.0014^a \end{array}$	$\begin{array}{c} 2.91 \pm 0.32^a \\ 114.3 \pm 3.93^a \\ 16.55 \pm 0.91^a \\ 0.052 \pm 0.0026^a \end{array}$	$5.86 \pm 0.33^{b} \\ 92.2 \pm 2.11^{b} \\ 12.16 \pm 1.08^{b} \\ 0.028 \pm 0.0016^{b} \\ \end{cases}$	$\begin{array}{c} 2.92 \pm 0.13^a \\ 111.3 \pm 3.00^a \\ 13.91 \pm 0.26^c \\ 0.043 \pm 0.0027^c \end{array}$	0.001 0.001 0.005 0.001

Means bearing different superscripts within same row were significantly different.

	Control	Lycopene	Priapism	Priapism+ Lycopene	<i>p</i> value
TBARS μmol/ml GSH nmol/ml SOD U/mg protein	$\begin{array}{c} 3.33 \pm 0.28^{a} \\ 66.14 \pm 3.49^{a} \\ 10.73 \pm 0.61^{a,c} \end{array}$	3.06 ± 0.40^{a} $63.46 \pm 1.11^{a,c}$ 11.06 ± 1.51^{a}	$\begin{array}{c} 6.63 \pm 0.49^{\text{b}} \\ 40.63 \pm 1.13^{\text{b}} \\ 8.55 \pm 0.42^{\text{b,c}} \end{array}$	4.20 ± 0.29^{a} 57.01 ± 2.14 ^c 9.26 ±1.20 ^c	0.001
CAT U/mg protein NO µmol/l	$\begin{array}{c} 0.081 \pm 0.0014^{a} \\ 7.15 \pm 0.88^{a} \end{array}$	$\begin{array}{c} 0.084 \pm 0.0026^{a} \\ 6.92 \pm 0.51^{a} \end{array}$	0.078 ± 0.0016^{a} 11.13 $\pm 0.31^{b}$	0.083 ± 0.0027^{a} 8.84 ± 0.42^{c}	0.195 0.01

Table II. The SOD and CAT activities and GSH and TBARS levels in serum samples of rats.

Means bearing different superscripts within same row were significantly different.

activity of reactive oxygen (ROS) and nitrogen species (RNS) in the penis influence vascular homeostasis of this organ, with adverse effects exerted at cellular and molecular levels^{1,8}. In this context, we thought that ROS scavengers and antioxidant agents could be beneficial in the clinical management of ischemic priapism. In this work we showed that ischemic priapism lead to oxidative and nitrosative damage in both covernosal tissue and serum samples. On the other hand, lycopene treatment in ischemic priapism prevent oxidative and nitrosative stress and caused reoxygenation of the tissue.

TBARS, a degradation product from li droperoxidation, provides an index of per tion of lipids in biological tissues²⁷. It has a portant role in the etiology and pathogenesi several diseases and disorders ischen priapism². In our study the r s sho that or cant lip hour priapism lead to a si beroxidation by increasing TBARS and corporal tissue et ats. Si Un al¹⁸ determined ne hour p caused a significant ing level, a nalondiald marker of oxidative str rats. Besides, Kanika et al² als erimental priapism oserved that d to damage in d model al tissue proteins eased l'hid peroxidations. Previous studvia ies² rm results. Lycopene could decrease RS leve nduced by priapism and elevate pid roxidations. In agreement prev hany previous studies clearly our fin nstrated that lycopene is a potent antioxidant de lipid peroxidations^{12,13,16}. A, GPx (enzymatic) and GSH (nonen-SOF natic) are members of body antioxidant defense

ms protecting against oxidative damage caused by free radicals production in normal physiological conditions and against molecules such as TBARS²⁸. In the current study, it was shown that 1 hour priapism caused an attenuation in both enzymatic and nonenzymatic antioxidant defense sys-

). CAT Px and tems via a decrease of levels in rats. However, N atment significantly induced a *k*idant ity in s and cavernosal tir and could idative riapism. Tr sults were damage car confirmed y prev researches which showed ease of antioxidant cathat priorism lead to a ts¹⁸. Besides al reports^{29,30} clearly pag in ated that the lycopene deatment increased annany diseases including cancer ti dant status ir exication. results clearly showed that 1 ai apism c ed an imbalance in oxidant/anhot nd may lead to erectile dysfunctioxia ions due to cavernosal tissue damage in penis. er, lycopene can prevent oxidative damage losal tissue and may reverse the deleterious effects of priapism.

NO, known as endothelium-derived relaxing factor (EDRF), is an important cellular signaling molecule involved in many physiological and pathological processes³¹. The endothelium of blood vessels uses nitric oxide to signal the surrounding smooth muscle to relax, thus resulting in vasodilation and increased blood flow³²⁻³⁴. Also, NO has an important function in penile erection via coordination with corpus cavernosal smooth muscle relaxation³². In our study it was observed that priapism caused a significant increase in NO levels, but lycopene decreased elevated NO levels in serum. Elevation of NO may be due to ischemia conditions and elevated oxidative stress in penis. Similarly, Uluocak et al¹⁸ showed that priapism increase NO levels in serum and melatonin brought the levels of NO closer to normal levels. These findings agree with our findings.

Conclusions

Oxidative and nitrosative stress play important role in many ischemic disorders such as pri-

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apism. For that, if oxidative and nitrosative stress are prevented with any agent, the damage caused by priapism can be reversed. In this study, it was observed that an hour priapism caused ischemia and a significant oxidative and nitrosative damage via an increase of TBARS and NO levels and a decrease of SOD, CAT, GSH and Gpx levels in serum and cavernosal tissue in rats. Additionaly, it was determined that lycopene eliminates oxidative and nitrosative damage in cavernosal tissue and serum due to their strong antioxidative properties. In conclusion because cavernosal damage may occur in erectile dysfunction in male, and therefore, quality of life may be impaired. lycopene may be used for the clinical treatment of priapism.

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Conflict of Interest

The Authors declare that there are no conflicts of intere

References

- 1) ROCHAT MC. Priapism: a review Dyiogenol 2001; 56: 713-722.
- KANIKA ND, MELMAN A, Day KP. Experiental principality apism is associated to increase coxidative stress and activation of provide the ways in corporal to ze. Int success 22: 363-373.
- 3) KUTTIN ES, the second ska A. Uron marcinoma with metas sis as the ted with pria, or in a sea lion. Is 1 J Vet Med 150: 163-165.
- Muther N, Hosking Dh. corporeal phenylerene in the treatment of papism. J Urol 1996; 141-1
- 5) Service MES W, BADON MJ. Priapism associated with a call antiper notic medications: a review. Int Cline Shore macol 2008; 23: 9-17.
 - UNES KI, CONVALVES A, LANZA LF, CORTES SF, CORDEIRO MN, RICHARDSON M, et al. Tx2-6 toxin of Conduction igriventer spider potentiates rat unction. Toxicon 2008; 51: 1197-206.
 - EL-BAHNASAWY MS, DAWOOD A, FAROUK A. Low-flow priapism: risk factors for erectile dysfunction. BJU int 2002; 89: 285-290.
- 8) KANIKA ND, TAR M, TONG Y, KUPPAM DSR, MELMAN A, DAVIES KP. The mechanism of opiorphin-induced experimental priapism in rats involves activation of the polyamine synthetic pathway. Am J Physiol Cell Physiol 2009; 297: 916-927.

- HEBER D, LU OY. Overview of mechanisms of action of lycopene. Experimental Biol Med 2002; 227: 920-923.
- RIED K, FAKLER P. Protective effect of lycopene on serum cholesterol and blood pressure: metaanalyses of intervention trials. Maturitas 2011: 68: 299-310.
- RAO AV, AGARWAL S. Role of antioxide a gcopend in cancer and heart disease. J Am A Nutr 2000; 19: 563-569.
- SAHIN K, SAHIN N, KUCUK O. Ly the and Chemotherapy Toxicity. M Cancel 1: 62: 988-995.
- WERTZ K, SILER U, GOL ZYK R. L'INOPENE: In of action to promote unite burn. Archives of Biochemistry 201 Bio, 2004; 421: 127-134.
- 14) CONKLIN Key stary antioxics of ang cancer chemoty ap, spact on chemotyrapeutic effectiveness and solopment of side effects. Nutr Cancer 2000; 37: 10
- 15) BD, KELLY KM, Los EJ, SAGAR SM, VICKERS A, BLUMBERG JG. Should applemental antioxidant admininistration be avoided during chemotherapy and radiation prapy? J Natl Cancer Inst 2008; 10: 773-783
 - V Racting. Carotenoids and human health. es 2007; 55: 207-216.
 - ATESSAHIN A, ÇERIBA I AO, YILMAZ S. Lycopene, a notenoid, attenuates cyclosporine-induced dysfunction and oxidative stress in rats. Basic Clin Pharmacol Toxic 2007; 100: 372-376.
- 18) ULUOCAK N, ATILGAN D, ERDEMIR F, PARLAKTAS BS, YASAR A, ERKORKMAZ U, AKBAS A. An animal model of ischemic priapism and the effects of melatonin on antioxidant enzymes and oxidative injury parameters in rat penis. Int Urol Nephrol 2010; 42: 889-895.
- 19) SANLI O, ARMAGAN A, KANDIRALI E, OZERMAN B, AHME-DOV I, SOLAKOGLU S, NURTEN A, TUNÇ M, UYSAL V, KA-DIOGLU A. TGF-b1 neutralizing antibodies decrease the fibrotic effects of ischemic priapism. Int J Impot Res 2004; 16: 492-497.
- YAGI K. Simple assay for the level of total lipid peroxides in serum or plasma. Methods Mol Biol 1998; 108: 101-106.
- SEDLAK J, LINDSAY RH. Estimation of total, proteinbound, and nonprotein sulfhydryl groups in tissue with Ellman's reagent. Anal Biochem 1968; 25: 192-205.
- 22) SUN Y, OBERLEY LW, LI YA. Simple method for clinical assay of superoxide dismutase. Clin Chem 1988; 34: 497-500.
- AEBI H. Catalase. In: Bergmeyer HU (eds). Methods of enzymatic analysis. Academic Pres New York 1974; pp. 673-677.
- 24) LOWRY OH, ROSEBROUGH NJ, FARR AL, RANDALL RI. Protein measurement with folin phenol reagent. J Biol Chem 1951; 193: 265-275.

- 25) CORTAS NK, WAKID NW. Determination of inorganic nitrate in serum and urine by a kinetic cadmium-reduction method. Clin Chem 1990; 36: 1440-1443.
- 26) AKDAG MZ, BILGIN MH, DASDAG S, TUMER C. Alteration of nitric oxide production in rats exposed to a prolonged, extremely low-frequency magnetic field. Electromagnetic Biol Med 2007; 26: 99-106.
- 27) CIFTCI O, AYDIN M, OZDEMIR I, VARDI N. Quercetin prevents 2, 3, 7, 8-tetrachlorobenzo-p-dioxininduced testicular damage in rats. Andrologia 2012; 44: 164-173.
- 28) SCHILLER HJ, REILLY PM, BULKLEY GB. Tissue perfusion in critical illnesses. Antioxidant therapy. Crit Care Med 1993; 21: 92-102.
- 29) TODA N, AYAJIKI K, OKAMURA T. Nitric oxide and penile erectile function. Pharmacol Ther 2005; 106: 233-266.

- 30) Rostoka E, Isajevs S, Baumane L, Line Alja, Silina K, Dzintare M, Sharipova J, Svirina D, Kalvinsh I, Sjak-STE N. Effects of lycopene, indole-3-carbinol, and luteolin on nitric oxide production and inos expression are organ-specific in rats. Arc Hig Rada Toksikol 2010; 61: 275-285.
- 31) HOU YC, JANCZUK A, WANG PG. Current the development of nitric oxide de CD Pharm Des 1999; 5: 417-441.
- 32) BURNETT AL. Role of nitric oxide physiology of erection. Biol Reprod 1995; 52: 4

m

- 33) Ignarro LJ, Buga GM, Wo S. Endo -deed and relea rived relaxing factor pro kide. Proc Natl Ad artery and vein is nitr 1987; 84: 9265-996
- 34) FURCHGOTT RF. histo ey and pr spects eikatsu DRF. N of research Gakkai 435-440. Zasshi 198