2017; 21: 3193-3199

Inhibition of miR-221 influences bladder cancer cell proliferation and apoptosis

H. LIU¹, J.-K. CHANG¹, J.-Q. HOU¹, Z.-H. ZHAO¹, L.-D. ZHANG²

¹Department of Urology, Huaihe Hospital, Henan University, Kaifeng, China ²Department of Urology, Shanxian Central Hospital, Shandong Province, Herochina

Ke

Hui Liu and Jun-Kai Chang are equal contributors

Abstract. - OBJECTIVE: Janus kinase (JAK) - signal transducer and activator of transcription (STAT) signaling pathway participate in cell proliferation and apoptosis. Suppressors of cytokine signaling 3 (SOCS3) are negative regulators of JAK-STAT3. SOCS3 was found significantly declined, while microRNA-221 (miR-221) obviously up-regulated in bladder cancer tissue. Bioinformatics analysis revealed the complementary binding site between miR-221 and 3'-UTR of SOCS3. This study investigated therable of miR-221 in regulating SOCS3/JAK-STAT naling pathway and bladder cancer cell pathenel ation and apoptosis.

PATIENTS AND METHODS: Bladder cand mor tissue and para-carcinoma tissue were lected from patients to test miR-201 and SOO expressions. Dual luciferase used test the targeting regulatory ect o l-221 or SOCS3. MiR-221, SOCS3 JAK1, p K2, and and survivin expressions we pare HBEC cells. T24 cells vere oR-SOCS3, miR-221 inhibitor, p R-bla and miR-221 inhi r + pSicoR groups. Flow cytometry plied to det apoptosis. EdU sta dopted to ev uate cell 1g proliferation. **RESUL** MiR-221 s. antly increased,

3 obviously real in bladder canwhile S e compared with para-carcinoma tiscer ti eted inhibited SOCS3 expres-R-221 † sue sion phosphorylated JAK1 (p-JAK1), phosp d JAK? JAK2), phosphorylat-TAT3 nd survivin levels mark-STAT p-reg nereas SOCS3 expression ntly de in T24 cells compared with ap tha HBEC cens. MiR-221 inhibitor and/or pSielevated SOCS3 expression, de-CC K1, p-JAK2, p-STAT3, and survivin els, enhanced cell apoptosis, and attenuated roliferation.

CLUSIONS: MiR-221 elevated, while SOC 3 reduced in bladder cancer tissue. Inhibition of miR-221 suppressed T24 cell proliferation and induced apoptosis by up-regulating SOCS3 exercises of lowering JA, AT3 signaling pather, act, and attenuating survivin expression.

uR-221, SOCS3, JAK-STAI, Bladder cancer, Proliftion, Apoptosis.

troduction

Bladder cancer (BC) is a common malignancy with the 9th morbidity among all types

⁴. It accounts for the leading morbidity and mortality in urinary system malignancy in our country². BC, especially muscle-invasive BC, is easy to occur pelvic lymph node metastasis or distant metastasis. Its 5-year survival may decrease lower than 30% once metastasis³. Following the development of detection and diagnostic technique, most BC patients were found in early stage. However, there are still about 30% patients appeared tissue infiltration and metastasis when diagnosed, leading to poor therapeutic effect and prognosis⁴. Janus kinase (JAK) - signal transducer and activator of transcription (STAT) signaling pathway widely exists in various tissues and cells. It participates in the regulation of cell proliferation⁵, apoptosis⁶, migration, and invasion⁷. Suppressors of cytokine signaling 3 (SOCS3) are members of SOCS family with the strongest activity. It suppresses JAK-STAT signaling pathway activation through direct inhibiting JAK⁸. SOCS3 was found down-regulated in breast cancer⁹ and prostate cancer¹⁰, and was closely associated with tumor progress and pathological classification. MicroRNA (miRNA) is a type of endogenous single stranded non-coding RNA at the length of 22-25 nt. It plays a degrading or inhibiting role on mRNA by binding with the 3'-UTR.

miRNA expression and function in tumorigenesis receive more and more attention¹¹. MiR-221 was showed significantly up-regulated in BC tissue, suggesting its potential oncogene role in BC occurrence¹². Bioinformatics analysis revealed the complementary binding site between miR-221 and 3'-UTR of SOCS3. This study investigated the role of miR-221 in regulating SOCS3/JAK-STAT3 signaling pathway and bladder cancer cell proliferation and apoptosis.

Patients and Methods

Patients

A total of 42 BC patients were enrolled, including 32 males and 10 females with mean age at 62.9 (51-78) years old. Patients received treatment in Huaihe Hospital, Henan University between Apr and Nov 2016. Tumor tissue and para-carcinoma tissue were collected and stored at -80°C for RNA, protein, and immunofluorescence detection.

This study was approved by Ethics Committee in Huaihe Hospital, Henan University and a enrolled objects had signed informed comments

Main Reagents and Materials

Human BC cell T24 was purchased from (Shanghai, China). Human norr ladder e thelial cell HBEC was boug ibo Bio logical Technology Co., L Shang China). OMEM), Dulbecco's Modified Ea Mediun fetal bovine serum (FRS), BRL. Co. tomycin were purch d from Ltd. (Grand Islan VY, USA). ire RNA Kit was obtain TransGen L h (Beieagent Liporectamine jing, China). .nsfee urchased from itrogen Life Tech-2000 was rimeScript[™] RT nologies arlsbad, CA, US. reager it and SYBR Greek were purchased KaRa lian, China). miR-NC, miR-221 fro inhibit and EdU cell prolifemim n kit w purchased from Ribobio ration a Jouse anti-human p-JAK1, ngzho na 3 primary antibodies were 2, andsed from Locam Biotechnology (Cambripur **ISA**). Rabbit anti-human p-STAT3 and ary antibodies were obtained from eTex Inc. (Irvine, CA, USA). Horseradish Pe-(HRP) conjugated secondary antibody rived from Bio-Rad Laboratories (Hercuwa. les, CA, USA). Fluorescence secondary antibody was purchased from Molecular Probes (Eugene,

OR, USA). pGRE-luc reporter gene plasmid, dual luciferase detection kit, apoptosis detection kit, and radio-immunoprecipitation assay (RIPA) were bought from Beyotime (Shanghen pSicoR-GFP was obtained from Advane (Lon. don, UK).

Cell Culture

T24 and HBEC cells we caltured the becco's Modified Eagle Maraam (DMEM) ning 10% FBS and 1° penicillic-streptom, Cells were passaged

Dual-Lucifer Report 1e The polyn e chain react) products containing ngth of SOC zene 3'-UTR re cloned to pGRE-luc. or mutan, segme Next, it was transform DH5 α competent cells ced to select and asmid with correct ence, namely pGRE-SOCS3-wt and pGRE-CS3-mut, respectively. Then, pGRE-SOCS3-3-mut) was co-transfected to or pGRE-S 93T cells using Lipofectamine 2000, to-F h miP 1 mimic (or miR-221 inhibitor, get ne luciferase activity was detected or mikter cultured for 48 h.

Over-Expression Plasmid

CDS segment of SOCS3 gene was amplified and connected to pSicoR-GFP plasmid after double digestion. After transformation, the positive bacterial strain was selected and amplified to extract the recombinant plasmid. The plasmid with correct sequence was named pSicoR-GFP-SOCS3. pSicoR-GFP-blank was treated as control.

Cell Transfection and Grouping

T24 cells were divided into miR-NC, miR-221 inhibitor, pSicoR-blank, pSicoR-SOCS3, and miR-221 inhibitor + pSicoR-SOCS3 groups. Cells were detected on 72 h after transfection.

qRT-PCR

Total RNA was extracted by using EasyPure RNA Kit and reverse transcribed to cDNA by using PrimeScript[™] RT reagent Kit. The reverse transcription condition was 37°C for 15 min and 98°C for 5 min. The PCR reaction was composed of 95°C pre-denaturation for 5 min, followed by 40 cycles of 95°C denaturation for 15 s, 60°C annealing for 30 s, and 74°C elongation for 30 s. Real-time PCR was performed on CFX96 Touch[™] to test the relative expression. The primers used were as follows. miR-221P_r: 5'-GTTGGTGGGAGCTACATTGTCTGC-3', miR-221P_R: 5'-GTGTCGTGGACTCGGCAATTC-3'; U6P_F: 5'-ATTGGAACGATACAGAGAAGATT-3', U6P_R: 5'-GGAACGCTTCACGAATTTG-3'; SOC-S3P_F: 5'-CCTGCGCCTCAAGACCTTC-3', SOC-S3P_R: 5'-GTCACTGCGCTCCAGTAGAA-3'; SurvivinP_F: 5'-AGGACCACCGCATCTCTACAT-3', SurvivinP_R: 5'-AAGTCTGGCTCGTTCTCA-GTG-3'; β-actinP_F: 5'-GAACCCTAAGGCCA-AC-3', β-actinP_R: 5'-TGTCACGCACGATTTCC-3'.

Western Blot

Total protein was extracted by RIPA from tissue and cells. A total of 50 µg protein was separated by 10% sodium-lauryl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) and transferred to membrane. Next, the membrane was blocked and incubated with primary antibody at 4°C overnight (SOCS3, p-JAK1, p-JAK2, p-STAT3, survivin, and β -actin at 1:300, 1:200, 1:200, 1:200, 1:200, and 1:800, respectively). Then the membrane was incubated with secondary antibody (1:10000) for 60 min after washed by phosphate buffer saline-tween 20 (PBST) for three times. At last, the protein expression was detect enhanced chemiluminiscence (ECL).

Flow Cytometry

The cells were re-suspended in 500 μ l bind buffer and incubated with 5 μ l Accessin V-FL avoid of light for 15 min. Neuroper the adde to 5 μ l PI and tested on V CS XL CL flow cytometry to evaluate cells optosis.

EdU Staining

Cells were ad to EdU sol n logarincubated for . Then, thmic phase ap 5% trypsin and were cells were dig red b ter, they we collected; ked in paraformaldehvde, neutralized in ine. Next, cells were bated with 0.1% Trito, X-100 and re-suin PB^c At last, the cells were stained by spe 500 room temperature for 10 min and CS XL tested a L flow cytometry (BD CA, USA). ience Jo

Im nofluore cence Detection

So the section was fixed in precooled acetemperature for 10 min. After the tion was washed by phosphate buffer sali-S) for three times, it was blocked in PBS containg 2% bovine serum albumin (BSA) and 0.1% Triton X-100 at room temperature for 60 min. Next, the section was incubated with mouse anti-human SOCS3 primary antibody at 4°C overnight. Then, the section was incubated with Alexa Fluor 594 labeled secondary antibody at room temperature for 60 min. At last, the was stained by 0.1% 4',6-diamiding phenylla. dole (DAPI) for 1 min and observed under the microscope.

Statistical Analysis

All data analyses were formed by St software (SPSS Inc., C ago, IL/USA). The asurement data were ed a $ean \pm standard$ deviation and cor st or Ma Whiarec atistical d a was co significance

In Control of Control

iously reduced in tumor tissue compared and acent normal control (Figure 1B).



Figure 1. MiR-221 upregulated, while SOCS3 reduced in BC tissue. (A) qRT-PCR detection of miR-221 expression; (B) immunofluorescence detection of SOCS3 expression. *p<0.05, compared with para-carcinoma tissue.



Figure 7 R-221 targeted regulated OCS3 expression in T24 cells. (*A*) qRT-PCR detection of miR-221 and SOCS3 expressions; (*C*) western blot detection of particle expressions; (*C*) the binding site between miR-221 the 3'-UTR of SOCS3 mRNA; (*D*) duciferased say; (*E*) qRT-PCR detection of miR-221 and SOCS3 expressions. p < 0.05, compared with HBEC; p < 0.05, compared with

21 Ta. 21 regulated SOCS3 ssion in 24 Cells

level was markedly lower in T24 ls compared with HBEC cells, suggesting the vial regulatory relationship between miR-221 d SOCS3 (Figure 2A and B). Western blot demonstrated that p-JAK1, p-JAK2, p-STAT3, and survivin levels markedly up-regulated in T24 cells compared with that in HBEC cells (Figure 2B). microRNA.org online prediction showed the complementary binding site between miR-221 and the 3'-UTR of SOCS3 mRNA (Figure 2C). Dual luciferase assay revealed that miR-221 mimic and miR-221 inhibitor significantly declined and enhanced relative luciferase activity in HEK293 cells (Figure 2D), indicating the regulatory relationship between miR-221 and SOCS3

Ex

mRNA. MiR-221 mimic or inhibitor transfection apparently reduced or elevated SOCS3 mRNA expression in T24 cells, confirming that miR-221 targeted regulated SOCS3 expression (Figure 2E).

Inhibition of miR-221 Upregulated SOCS3 expression, Suppressed T24 cell Proliferation, and Promoted Apoptosis

MiR-221 inhibitor and/or pSicoR-SOCS3 elevated SOCS3 expression, decreased p-JAK1, p-JAK2, p-STAT3, and survivin levels (Figure 3A and B), enhanced cell apoptosis (Figure 3C), and attenuated cell proliferation (Figure 3D).

Relative gene expression of miR-221 and SOCS3

103

10²

100

0 10 B1 1.3%

B3 96.1%

10

2

Discussion



3. Inhibition of miR-221 up-regulated SOCS3 expression, suppressed T24 cell proliferation, and promoted apoptosis. PCR detection of miR-221 and SOCS3 expressions; (B) Western blot detection of protein expressions; (C) flow cytom-(A) q. etry detection of cell apoptosis; (D) EdU staining detection of cell proliferation. $^{a}p<0.05$, compared with miR-NC; $^{b}p<0.05$, compared with pSicoR-blank; $^{\circ}p < 0.05$, compared with miR-221 inhibitor; $^{\circ}p < 0.05$, compared with pSicoR-SOCS3.

D

factor family contains seven members, including STAT1, STAT2, STAT3, STAT4, STAT5a, STA-T5b, and STAT6. STAT3 is most widely investigated and exhibits the closest relationship with tumor occurrence and development. Inhibitor of apoptosis protein (IAPs) is a protein family with homological structure and anti-apoptosis function¹⁴. Survivin is the most important member of IAPs family with the strongest function in apoptosis inhibition. Survivin blocks cell apoptosis by inhibiting caspase-3 and caspase-715. Moreover, survivin also plavs a crucial role in regulating cell cycle and facilitating cell proliferation¹⁶. It was showed that JAK-STAT3 signaling pathway activation mediated survivin enhancement plays an important role in promoting malignancy^{17,18}. SOCS3 is the strongest member to SOCS family⁸. It restrains JAK1 and JAK2 phosphorylation and kinase activity by binding to receptor, thus blocking STAT3 phosphorylation and playing a negative regulatory role in JAK-STAT3 signaling pathway¹⁹. SOCS3 down-regulation is closely associated with multiple tumor occurrence and development, including colorectal cancer²⁰. lung cancer²¹, and breast cancer²². Recently it was found that SOCS3 expression reduced in b cancer tissue, suggesting its potential an effect²³. MiR-221 level was reported to be e ted in bladder cancer tissue, indicating its pron effect on bladder cancer¹². Bioinformatics ana revealed the complementary bind te betwe miR-221 and 3'-UTR of SOC dy inve stigated the role of miR-221 regulat SOCS3 er cancer JAK-STAT3 signaling pa and bl cell proliferation and apopto revealed that miR-22 xpress s significantly higher in cance arcinoma sue than in nce detection tissue. Immuno ved that luced in tumor tissue SOCS3 level vious ith adjacent control. It suggecompared sted that R-221 up-regula may play a role sing SOCS3 expression and promoting in de norigenesis. Gottardo et al¹² debla cancer miR-221 evel significantly increamon cancer t e compared with normal sed in b. ². In this study, miR-221 tis ler m ously higher in tumor tissue sion wa red with para-carcinoma tissue, which was con Gottardo et al¹² findings. Gaballah SU d that SOCS3 expression markedly lined in bladder cancer tissue compared with control. Its expression was related to pacal grading, as it declined in patients with tho. higher pathological grade. This study observed that SOCS3 expression abnormally reduced in

tumor tissue, which was in accordance with Gaballah et al²³. MiR-221 expression was obviously higher, while SOCS3 level was markedly lower in T24 cells compared with HBEC cells. re, p-JAK1, p-JAK2, p-STAT3, and s vin leve hpared with markedly up-regulated in T24 cells that in HBEC cells. It indicated iR-221 elevation, SOCS3 down-regulation, and STAT3 lated signaling pathway enhance nt may to the malignancy of bl er cancer. Li gulation plays a key showed that miR-221 u in T24 cell epithelia eser mal transition cell inva (EMT) induced b and TG migration enha g th niR-221 ment, re abnormal ele n was a regu malignanal luciferase cy enhance adder cancel. assay reversed that -221 mimic and miR-221 inhibitor significantly ned and enhanced rela-293 cells, and appativ ase activity in y reduced or elevated SOCS3 mRNA expresn in T24 cells, and firming that miR-221 targeted lated SOCS. pression. Moreover, miR-221 or and/or coR-SOCS3 elevated SOCS3 in dee sed p-JAK1, p-JAK2, p-STAT3, exp vels, enhanced cell apoptosis, and and sur tenuated cell proliferation. Liu et al²⁴ reported ression of miR-221 markedly declined expression, enhanced E-cadherin level, and attenuated EMT, migration, and invasion in T24 cells induced by TGF- β . Fu et al²⁵ revealed that miR-221 inhibitor transfection obviously increased Bax expression, declined Bcl-2 level, and accelerated cell apoptosis in bladder cancer cell 5637, J82, and T24. Fu et al²⁵ also reported that inhibition of miR-221 apparently decreased MMP-2, MMP-9, and VEGF-C expression, and weakened cell migration and invasion in bladder cancer cells²⁵. In this study, down-regulation of miR-221 weakened bladder cancer malignancy, which was in accordance with Liu et al²⁴ and Fu et al²⁵. This study demonstrated that miR-221 elevation plays a role in reducing SOCS3 expression, enhancing JAK-STAT3 signaling pathway activity, and promoting bladder cancer tumorigenesis.

Conclusions

MiR-221 elevated, while SOCS3 reduced in bladder cancer tissue. Inhibition of miR-221 suppressed T24 cell proliferation and induced apoptosis by up-regulating SOCS3 expression, lowering JAK-STAT3 signaling pathway activity, and attenuating survivin expression.

Conflict of Interest

The Authors declare that they have no conflict of interests.

References

- 1) MENG FM, MENG FM, SONG XL. MiR-576-3p is a novel marker correlated with poor clinical outcome in bladder cancer. Eur Rev Med Pharmacol Sci 2017; 21: 973-977.
- 2) CHEN W, ZHENG R, BAADE PD, ZHANG S, ZENG H, BRAY F, JEMAL A, YU XO, HE J. Cancer statistics in China, 2015. CA Cancer J Clin 2016; 66: 115-132.
- 3) Rose TL, Deal AM, Nielsen ME, Smith AB, Milowsky MI. Sex disparities in use of chemotherapy and survival in patients with advanced bladder cancer. Cancer 2016; 122: 2012-2020.
- MASSON-LECOMTE A, RAVA M, REAL FX, HARTMANN A, 4) ALLORY Y, MALATS N. Inflammatory biomarkers and bladder cancer prognosis: a systematic review. Eur Urol 2014; 66: 1078-1091.
- 5) YU RX, HU XM, XU SQ, JIANG ZJ, YANG W. Effects of fucoxanthin on proliferation and apoptosis in human gastric adenocarcinoma MGC-803 cells via JAK/STAT signal pathway. Eur J Pharmacol 2011; 657: 10-19.
- LI HX, ZHAO W, SHI Y, LI YN, ZHANG LS, ZHA 6) WANG D. Retinoic acid amide inhibits . pathway in lung cancer which leads to a Tumour Biol 2015; 36: 8671-8678.
- 7) Kowshik J, Baba AB, Giri H, Deepak Reddy G, M, NAGINI S. Astaxanthin inhibits JAK/STATgnaling to abrogate cell prolife vasion a angiogenesis in a hamster cance PLoS One 2014; 9: e1091
- H Yoshi-8) TAMIYA T, KASHIWAGI I, TA R, Yası MURA A. Suppressors of C of proteins and JAK AT pa regulance T-cell inflamma CS3. Arteby SOCS1 c Biol 2011; 985. rioscler Throp
- YA S, STEELE R, Y RB. An-MUHAMMAD positive breast cancer ti-miR-203 uppres. growt nd stemness i eting SOCS3. Oncotarg J16; 7: 58595-586
- 10) NTI F, MARTINI M, PINTO F, CENCI T, CAPODIMON-CALARC Bassi PF, Larocca LM. Epigenetic OCS3 identifies a subset of prostang vith an a ssive behavior. Prostate te 2011 8-325
 - C, W 🗸, Liu Z, Zhang J, Wang Z, Li R, NG S, WANG Y, XUE Y, YANG J, TAN ang Z, Li Song X. mx-942 promotes cancer stem cellin esophageal squamous cell carcinoma tivation of Wnt/beta-catenin signalling pathway. Oncotarget 2015; 6: 10964-10977.
 - ottardo F, Liu CG, Ferracin M, Calin GA, Fassan Bassi P, Sevignani C, Byrne D, Negrini M, Pagano GOMELLA LG, CROCE CM, BAFFA R. Micro-RNA profiling in kidney and bladder cancers. Urol Oncol 2007; 25: 387-392.

- 13) CHEN W, HONG YO, MENG ZL. Bioinformatics analysis of molecular mechanisms of chronic obstructive pulmonary disease. Eur Rev Med Pharmacol Sci 2014; 18: 3557-3563.
- 14) Rodriguez-Berriguete G, Torrealba N Martinez-Onsurbe P, Olmedilla G, gua R. Guil CID M, FRAILE B, ROYUELA M. Pr stic value of inhibitors of apoptosis proteil s) and caspases in prostate cancer: caspa rms and XIAP predict biochemica ogressi radincer 2015; cal prostatectomy. BM
- 15) WANG C, ZHENG X, С, SHI X MicroRN ation ar igration by suppresses cell pr man triph-negageting BIRC5 and o Clin C tive breast ca r Res r ce 2012; 31: 58
- 16) Zнао X, J, YUAN X, Zhang Z, Fen Li J. Ef RNA-silenceo and survivin on luc cance. BUON 2014; 19: roliferation and apoptosis. J
- S, GAO Q, YANG Z. MI-17) QIU W, ZHAN A-204 targets JAK breast cancer and induces cell apoptosis through the STAT3/BCI-2/survivin pathway. Int J Exp Pathol 2015; 8: 5017-5025.

Glienke W, MA L, WICHT J, BERGMANN L. CURCUMIN ibits const ve STAT3 phosphorylation in hucancer cell lines and downregulaancre n/BIRC5 gene expression. Cancer

Invest 2010; 28: 166-171.

the

sis.

- INS AS, MCCOY CE, LLOYD AT, O'FARRELLY C, SON NJ. miR-19a: an effective regulator of S3 and enhancer of JAK-STAT signalling. PLoS One 2013; 8: e69090.
- 20) IGCI M, CAKMAK EA, OZTUZCU S, BAYRAM A, ARSLAN A, GO-GEBAKAN B, IGCI YZ, CENGIZ B, OZKARA E, CAMCI C, DEMIRYU-REK AT. Mutational screening of the SOCS3 gene promoter in metastatic colorectal cancer patients. Genet Test Mol Biomarkers 2012; 16: 1395-1400.
- 21) ZHANG S, WANG W, WANG E, QIU X. SOCS3 expression is inversely correlated with Pyk2 in non-small cell lung cancer and exogenous SOCS3 inhibits proliferation and invasion of A549 cells. Pathology 2012; 44: 434-440.
- 22) BARCLAY JL, ANDERSON ST, WATERS MJ, CURLEWIS JD. SOCS3 as a tumor suppressor in breast cancer cells, and its regulation by PRL. Int J Cancer 2009; 124: 1756-1766.
- 23) GABALLAH HH, SHAFIK NM, WASFY RE, ABOU FARHA MO. Significance of suppressor of cytokine signaling-3 expression in bladder urothelial carcinoma in relation to proinflammatory cytokines and tumor histopathological grading. Asian Pac J Cancer Prev 2015; 16: 307-314.
- LIU J, CAO J, ZHAO X. miR-221 facilitates the TGF-24) beta1-induced epithelial-mesenchymal transition in human bladder cancer cells by targeting STMN1. BMC Urol 2015; 15: 36.
- 25) FU B, WANG Y, ZHANG X, LANG B, ZHOU X, XU X, ZENG T, LIU W, GUO J, WANG G. MIR-221-induced PUMA silencing mediates immune evasion of bladder cancer cells. Int J Oncol 2015; 46: 1169-1180.