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cinoma growth<sup>9</sup>. Studies have applied bioinformatics methods to screen gastric cancer and its adjacent non-cancerous tissues and found that MCM3AP-AS1 was up-regulated in gastric cancer. All evidence suggested that MCM3AP-AS1 may participate in the pathogenesis in human GC. However, the effects of MCM3AP-AS1 on biological behaviors such as cell proliferation and apoptosis of GC have not been studied completely. The study aimed to discover the role of MC-M3AP-AS1 on cell proliferation and apoptosis in GC cell lines *in vitro*.

Over the past decades, microRNA (miRNA) has been proved to be involved in tumorigenesis<sup>10,11</sup>. The expression of miR-181a-5p effected on cell proliferation in GC<sup>12</sup>. Meanwhile, miR-708-5p regulated prostate cancer cell apoptosis<sup>13</sup>; overexpression of miR-708-5p had a certain inhibitory effect on cell proliferation<sup>14</sup>. MiR-708-5p referred to the process of GC. The study demonstrated miR-708-5p expression was downregulated in GC cell lines. However, it has not been reported whether MCM3AP-AS1 targeted and regulated the biological behavior of miR-708-5p regulating GC cells *in vitro*.

In this study, bioinformatics methods, the lecular biology techniques were used to a stigate whether MCM3AP-AS1 targeted miles 5p to regulate the proliferation and apoptos gastric cancer cells, in order to find new the peutic targets for clinical.

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# Materials a

Cell Culture and ansien fection Normal gastrig cosal cells nd gastric cancer cells 803 and SGC J1 were Academy of Sciencpurchased fre he 🖒 es Cell Bark (Shanghai, Of which GES-1, MGc-802 nd SGC-7901 cc re cultured in Dulbeg Modified Eagle's M. dium (DMEM; , South ogan, UT, USA) medium con-HyC ₩ (**y** tai me fraction) fetal bovine serum Fisher 2 (FBS, htific, Waltham, MA, USA) and le an otics (penicillin 100 U/  $\mu$ g/mg) (Gibco, Rockville, 5% CO<sub>2</sub> in cell incubator. repton M SA) at a 🕽 vectors of MCM3AP-AS1 overexpres-1-MCM3AP-AS1), overexpresntrol (pcDNA3.1), MCM3AP-AS1 kdown (si- MCM3AP-AS1), knockdown control (si-NC), miR-708-5p inhibitor R-708-5p) and inhibitor negative control (ant)

(anti-miR-NC) were purchased from Sangon Biotech (Shanghai, China). Subsequently h after cell passage, the cells were a logarimic growth; next, Lipofectamine<sup>V</sup> 200 (Thermo Fisher Scientific) was carried to transfect vectors into cells according to h manufacturer's manual.

# Quantitative Real-Tupe Polymerase Chain Reaction ( -PCR)

The logarithmic pþ and trav fected 48 h cells were ollec e total ] A was he extracted by zol (The *ientific).* **First-Strand** sis System SuperScript entific) was to d to reverse (Thermo, for qRT-PCR, reacting transcril into solutions were mixed ing iQ<sup>™</sup> SYBR<sup>®</sup> Green Sup (Bio-Rad, h s, CA, USA), U6 DH were performed as internal refer-e genes. The process was as follow: 95°C for ninutes, 40 ( s at 95°C for 15 s, 60°C for and 72°C 20 s, and then signals were ed by Qu Studio 3 QS3 (Thermo Fisher C pecial primers were as follows: Scie miR-708-5p. (rorward: 5'-GGC GCG CAA GGA TAC AAT C-3', Reverse: 5'-GTG CAG AGG TAT-3'); U6: (Forward: 5'-GAG G CGG AAC G-3', Reverse: 5'-CTA CCA CAT AGT CCA GG-3'); MCM3AP-AS1: (Forward: 5'-GCT GCT AAT GGC AAC ACT GA-3', Reverse: 5'-AGG TGC TGT CTG GTG GAG AT-3'); GAPDH: (Forward 5'- GGT GAA GGT CGG AGT CAA CG-3', Reverse: 5'-CAA AGT TGT CAT GGA TGA ACC-3')

#### Western Blot

The total protein was extracted from MGc-803 and SGC-7901 cells condition in the logarithmic growth phase after transfecting for 48 h. The total protein concentration was determined by BCA Protein Assay Kit (Pierce, Waltham, MA, USA). SDS-polyacrylamide gel was prepared, 50 µg denatured protein was added per well, and electrophoresis was run. The isolated proteins were transferred onto polyvinylidene difluoride (PVDF) membrane (Roche, Basel, Switzerland); the voltage was controlled at 100 mV for 70 minutes. Then skim milk was used (Sangon Biotech) to block PVDF for 60 min; the diluted primary antibody was incubated at 4°C overnight. Tris Buffered Saline Tween (TBST) (Sangon Biotech) was washed three times for 5 min each. The diluted secondary antibody was incubated at room temperature for 60 minutes in the shaker, TBST was washed three times for 5 min each. In the end point, protein signals were identified by using SuperSignal Chemiluminescent Substrates (Thermo Fisher Scientific) through Image Lab software (Bio-Rad). The special antibodies used in the study are listed as follows: the primary antibodies of Cyclin D1 (ab16663, 1:100), P21 (ab109520, 1:5000), Bax (ab32503, 1:5000), Bcl-2 (ab182858, 1:2000) and GAPDH (ab9485, 1:2500) were purchased from Abcam (Cambridge, MA, USA).

#### MTT Assay

2.5% trypsin (Gibco) was carried out to digest MGc-803 and SGC-7901 cells, the cell concentration was adjusted to 1 ' 10<sup>5</sup>/ml and 100 µl per well were added in 96-well plates. Whereafter, they were mixed with 20 µl MTT reagent (Beyotime, Shanghai, China) and incubated for 4 hours before detection. Finally, the absorbance was measured at 490 nm (OD) by SpectraMax<sup>®</sup>iD3 (Molecular Devices, San Jose, CA, USA).

#### Flow Cytometry

After transfection, MGc-803 and SGC-7901 cells were cultured for 48 h, then the m was discarded and washed twice with PB phate-buffered saline) (HyClone). Subse tly, 500 ul 1×Binding Buffer, 5 ul Annexin V (Biosea Biotechnology, Beijing, China) were ed in solutions and incubated for 15 minutes the dark place; 2.5 µl PI (Pror lide) wa flow cyadded and incubated for 5 ates, etect ce tometry was performed poptosis in vitro.

#### Dual-Luciferase

Constructed wildty T-MCors: -pGL3-M3AP-AS1 3' mutant (M pression vectors of MCM3AP-A UT MCM3AP-ASI, renilla rs (pGL3) were purchase om Promega 🖪 n, WI, USA). dy, they were transpected with one Subseq of th into MSc-803 and SGC-7901 cells by ne<sup>™</sup> 2000 and cultured for 48 fect usi Dual-Lu rase assay kit (Promehours. lyse cells and stimulate trate was a signals were captured by ase sig olecular Devices, San Jose, Max®iD Sp CA ЬA).

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#### succes nalysis

PSS 22.0 (SPSS Inc., IBM, Armonk, NY, as used for data analysis. The Student's *t*-test, as used for comparison between the two groups. One-way analysis of variance with posthoc test (ANOVA) was followed by T for multiple comparisons of group or graph Prism (San Diego, CA, USA) we performed to plot images. p<0.05 indicates to the difference was statistically significant.

## The Level of MCn. As Was Strikingly Increased Cell Li

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### hockdown of MCM3AP-AS1 rikingly Superssed Cell liferation in GC Cell Lines linterfordg RNA (si-RNA) of MC-

M3. (STAC) MCM3AP-AS1) and negative control (STAC) were constructed and transfectinto GC cell lines, respectively. The level AP-AS1 was strikingly decreased in MC 3AP-AS1 group (Figure 2A and 2D). What's more, MTT assay was performed to detect GC cell proliferation *in vitro*. Evidence revealed that knockdown of MCM3AP-AS1 curbed cell proliferation in GC cell lines (Figure 2B and 2E). Furthermore, the expression of



Figure 1. The level of MCM3AP-AS1 was strikingly increased in GC cell lines. The level of MCM3AP-AS1 in MGc-803 and SGC-7901 cells was measured by qRT-PCR.  $^{a}p$ <0.05.



**Figure 2.** Knockdown of MCM3AP-AS1 strikingly suppressed can align the provided the superstantial strikingly suppressed can align the provided the superstantial strikingly suppressed can align the superstantial strikingly superstantial strikingly suppressed can align the superstantial strikingly strikely strikingly strikingly strikingly striking

Cyclin D1 and P21, which was relative to cell cycles, was measured by Western blot a the effect of MCM3AP-AS1 decomplation of cell proliferation *in vitro* gure and 2F) was verified.

<1 Downregulation МСМЪ Significantly P oted Cell Apoptosis in Lines NC or si-MCM3AP-After trans ling AS1 into MGc-803 and S 1 cells, cell apoptosis was dyzed by flow cy ry. The results that knockdown of M. M3AP-AS1 ensugges ell aportosis in GC cell lines (Figure 3A, hanc 3B. Meanwhile, the expression of nd was mea Bax a d, which was referred Vest olot. It proved the result apopto AP-AS1 on cell apoptosis effect nd 3F). (Figure in

# eroon, Was a Target Gene of W3AP-AS1

formatics software was used to predict the relat. ship between MCM3AP-AS1 and miR-

/08-5p and proved they had binding sites (Figure 4A). After transfection with WT-MCM3AP-AS1

4A). After transfection with W1-MCM3AP-ASI or mut-MCM3AP-AS1, miR-708-5p was verified to be a target gene of MCM3AP-AS1 (Figure 4B). Moreover, the level of miR-708-5p was decreased by the upregulation of MCM3AP-AS1 while enhanced the expression through knockdown of MCM3AP-AS1 (Figure 4C).

#### Knockdown of miR-708-5p Rescued the Effect of MCM3AP-AS1 on MGc-803 and SGC-7901 Cell Proliferation

After transfection with si-NC, si-MCM3AP-AS1, si-MCM3AP-AS1 + anti-miR-NC or si-MCM3AP-AS1 + anti-miR-708-5p into GC cell lines, the level of miR-708-5p was promoted by si-MCM3AP-AS1 while rescued *via* knockdown of miR-708-5p *in vitro* (Figure 5A and 5D). In addition, the downregulation of miR-708-5p reversed the effect of si-MCM3AP-AS1 on cell proliferation in GC cell lines (Figure 5B and 5E). Cell cycles relative proteins of Cyclin D1 and P21 also verified the conclusion of GC cell proliferation *in vitro*.



Figure 3. Downregulation of MCM34 cells (A-F), GC cell apoptosis was ide (C and F).  $^{a}p < 0.05$ .



by f

cantly promoted cell apoptosis in GC cell lines. After transfecting with si-MCM3AP-AS1 into MGc-803 and SGC-7901 AS1 sigr cytometry (A, B, D and E). The expression of apoptosis-relative proteins of Bax and Bcl2 was examined by Western blot



**Figure 4.** miR-708-5p a target get the CM3AP-AS1. The binding sites between MCM3AP-AS1 and miR-708-5p were predicted by bioinform as software of the **(A)**. Dual-luciferase reporter assay was used to verify the relationship of MCM3AP-AS1 and miR-708-5p were the level of miR-708-5p was measured after transfecting with up or down regulation more to the AD of the control of the transfecting with up or the transfecting wi

#### Dowr ulation of miR-70,-5p d the Sfect of MCM3AP-AS1 Res sis in GC Cell Lines Apo on То investi the role of MC-M3AP-A 08-5p on GC progresmi o-tran was carried out with MCx-708-5p vectors, including AS1 and M. si-MCM3AP-AS1, si-MCM3AP-AS1 + si-N an si-MCM3AP-AS1 + anti-miR-MGc-803 and SGC-7901 cells. expression of miR-708-5p regained the si-MCM3AP-AS1 on cell apoptosis (Figure 6A and 6C). Meanwhile, the in vi

conclusion was also verified by measuring the expression of Bax and Bcl-2 in GC cell lines (Figure 6B and 6D).

# Discussion

Gastric cancer is a malignant tumor that occurs in the gastric mucosa. Its incidence rate ranks first among all malignant tumors in China. Due to the different living habits and dietary structure in the north and the south, the incidence of gastric cancer in coastal areas and northwestern China



Figure 5. Knockdown of miR-708-5p it MCM3AP-AS1 + anti-miR-NC or si-MCM3A was used to detect cell prolifera cell h **F**). <sup>*a*</sup>*p*<0.05, <sup>*b*</sup>*p*<0.01.

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effect o CM3AP-AS1 on MGc-803 and SGC-7901 cell proliferation. Transfected vectors of si-NC, si-MCM3AP-AS1, si-AR-708-5p into GC cell lines (A-F). The level of miR-708-5p was measured by qRT-PCR (A and D). Moreover, MTT (E). Finally, the expression of apoptosis-relative proteins of Cyclin D1 and P21 was examined by Western blot (C and



**Figure 6.** Downregulation of miR-708-5p restored lines transfected with si-NC, si-MCM3AP-AS1, si-MC into MGc-803 and SGC-7901 cells (**A-D**), respectively Meanwhile, Bax and Bcl-2 expression was also identified

is significantly higher that at in the ath. The recurrence and metasta umor main reason for the difficult It is reported that t r metas d invasion are affected by v is lncRNAs c he past decades. Theref ole of IncRN gastric nce for the targeted cancer is of at sh treatment of gastric cane

LncR) is a non-coding that occupies 98% aman gene transcrip, and is greater J nt in ngth<sup>15</sup>. In recent years, it has than bee ncRNA can regulate the activd th les at multiple levels, ity of protein affecting ologi behaviors such as cell ation and apoptosis of turation, ls<sup>16-18</sup>. St s have shown that lncRNA mo AP-AS1 expression in primary liver can-MC ce higher than adjacent tissues and tissues<sup>19</sup>. Further, MCM3AP-AS1 significantly up-regulated in glioblastoma, bition of MCM3AP-AS1 expression inhibu ell viability, migration, and angiogenesis

ffect of anti-mrR-NC or si-MCM3AP-AS1 + anti-miR-708-5p into ptosis was measured by flow cytometry *in vitro* (A and C). estern blot (B and D).  ${}^ap$ <0.05,  ${}^bp$ <0.01.

in glioblastoma<sup>20</sup>. The qRT-PCR assay showed that the level of MCM3AP-AS1 in two GC cells was higher than that in normal GC cells, which was consistent with the above results, suggesting that MCM3AP-AS1 was a key factor in the development and progression of gastric cancer. Meanwhile, Wang et al<sup>9</sup> reported that MCM3AP-AS1 was elevated in hepatocellular carcinoma, which promoted the growth of hepatocellular carcinoma by targeting inhibited expression of mir-194-5p. To further investigate the role of MCM3AP-AS1 in GC, this study repressed the expression of MCM3AP-AS1 in GC cell lines and detected cell proliferation and apoptosis in vitro. The results showed that after inhibiting the expression of MCM3AP-AS1, GC cell proliferation was significantly decreased, and the cell apoptosis rate was enhanced. The levels of Cyclin D1 and Bcl-2 were significantly decreased, and the levels of P21 and Bax were notably increased, suggesting that MCM3AP-AS1 may promote cell proliferation and inhibit apoptosis. In addition, the expression of miR-708-5p was significantly increased after inhibiting the expression of MCM3AP-AS1, suggesting that MCM3AP-AS1 may regulate the expression of miR-708-5p.

MiR-708-5p is a newly discovered microRNA that plays an important role in cell proliferation, migration, invasion, and apoptosis in tumor<sup>13,21</sup>. The miRcod bioinformatics software predicted that MCM3AP-AS1 and miR-708-5p had common fragment sequences, speculating that miR-708-5p may be the target gene of MCM3AP-AS1. Subsequently, Dual-Luciferase report assay and western blot were carried out to further verify MCM3AP- AS1 negatively regulated the expression of miR-708-5p by directly binding to the miR-708-5p 3'UTR region.

Studies have reported that miR-708-5p expression is decreased in human mesenchymal stem cells, and overexpression of miR-708-5p promoted cell proliferation and migration. The expression of miR-708-5p was down-regulated in renal cell carcinoma, and it could inhibit both cell apoptosis of renal cancer and the occurrence of renal cancer by inhibiting the oncogene zinc finger E-box binding to homologous boy tein 2<sup>22</sup>. Furthermore, miR-708-5p inhi migration of breast cancer by inhibiting endoplasmic reticulum membrane protein a ducing the activation of extracellular signal ulated kinases<sup>23</sup>. Afterward, the effect of M M3AP-AS1 targeting miR-70 GC cell was verified, anti-miR-708 sfected was into GC cells, which stab knockdo of MCd that M3AP-AS1. The results inhibitor reversed the effect tion of MCM3APon cell ration and apoptosis in vitr ression urthermore, nd Bcl-2 furth verified of Cyclin D1, P the above co sion.

In summary, MCMS. 1 targeted miR-708-5p to ulate cell prolite and apoptosis of GC s. This study clarifie, the mechanism 3AP-A21 targeting miR-708-5p and the of M hera or GC, providing potential thertar apeut Howey further investigations aman GC are needed. in clinica ent

# Conclusions

ed in GC cell lines. Further, the downregbe of MCM3AP-AS1 decreased cell proliferate and boosted apoptosis in MGc-803 and

SGC-7901 cells. miR-708-5p was a target gene of MCM3AP-AS1; its knockdown r effect of the downregulation of N GAP-A on cell proliferation and apopt in GC cell lines. **Conflict of Interest** The Authors declare that the ave no conflict of n *lefere* mates of the 1) Parkin P, FERLAY J. of eighteen major cancers in work le in 1985. Int J Can 3; 54: 594-606. KD, NEUGUT AI. niology of gastric can-2) orld J Gastroent 2006; 12: 354-362. Alberts SR, Cervantes A, Van de Velde CJ. Gastric cancer: epide ology, pathology and treatment. Ann Oncol 2 14: ii31-ii36. RENG WX, KC P, Mo YY. LncRNA-mediated ulation of signaling in cancer. Oncogene 5667.

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