

Long noncoding RNA DARS-AS1 acts as an oncogene by targeting miR-532-3p in ovarian cancer

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Abstract. – OBJECTIVE: Ovarian cancer is one of the most ordinary malignant tumors. Recently, the role of long noncoding RNAs (lncRNAs) in tumor progression has caught attention of numerous researchers. In this research, lncRNA DARS-AS1 was studied to identify how it functions in the development of ovarian cancer.

PATIENTS AND METHODS: DARS-AS1 expression was detected by Real-time quantitative polymerase chain reaction (RT-qPCR) in ovarian cancer tissue samples. Moreover, functional experiments were conducted to detect the effect of DARS-AS1 on the proliferation and metastasis of ovarian cancer. In addition, the underlying mechanism was explored through luciferase assay and RNA immunoprecipitation (RIP) assay.

RESULTS: In this study, DARS-AS1 expression was remarkably higher in ovarian cancer tissues compared with that in adjacent ones. Cell proliferation was inhibited after DARS-AS1 silencing in ovarian cancer cells. Moreover, cell migration and invasion were also inhibited after DARS-AS1 silenced in ovarian cancer cells. Furthermore, results of luciferase assay and RIP assay showed that microRNA-532-3p (miR-532-3p) was a direct target of DARS-AS1 in ovarian cancer. The expression of miR-532-3p was up-regulated after DARS-AS1 was knocked down.

CONCLUSIONS: Our study suggests that DARS-AS1 enhances cell proliferation and metastasis via sponging miR-532-3p in ovarian cancer.

Key Words:

Long noncoding RNA, DARS-AS1, Ovarian cancer, miR-532-3p

Introduction

Ovarian cancer is one of the most fatal gynecologic malignancies and is also the fifth leading cause of death among malignant cancers¹. The

high mortality of ovarian cancer is associated with the high occurrence of chemoresistance and metastasis^{2,3}. Therefore, it is urgent to find out new strategies in early detection of these patients and establish new therapeutic regimens for successful intervention.

Recent studies have revealed that long non-coding RNAs (lncRNAs) play a vital role in a variety of cellular activities in malignant tumors. For example, lncRNA SNHG7 enhances tumor growth ability in osteosarcoma by sponging miR-34a⁴. LncRNA SLC16 inhibits cell apoptosis in papillary thyroid cancer⁵. LncRNA FENDRR is a tumor suppressor⁶ in non-small cell lung cancer by targeting miR-761. Silence of lncRNA MEG3 promotes glioma proliferation and metastasis *via* regulating phosphatidylinositol 3-kinase (PI3K)/protein kinase B (AKT)/mammalian target of rapamycin (mTOR) pathway⁷.

In this study, we found out that the expression of DARS-AS1 was remarkably higher in ovarian cancer tissues. Moreover, DARS-AS1 promoted the proliferation, migration and invasion of ovarian cancer *in vitro*. In addition, we further found that the function of DARS-AS1 in ovarian cancer was also associated with microRNA-532-3p (miR-532-3p).

Patients and Methods

Tissue Specimens

52 cases of ovarian cancer tissues and their adjacent tissues were collected from patients who received surgery at Chinese PLA General Hospital. No chemotherapy or radiotherapy was received before surgical resection. All cases were diagnosed with ovarian cancer by two independent pathologists without any controversial. Writ-

ten consent form was obtained from each patient. This study was approved by the Ethics Committee of Chinese PLA General Hospital.

Cell Culture

Three cancer cell lines (A2780, SKOV3 and OVCAR-3), and normal ovarian cell (ISOE80) were purchased from the American Type Culture Collection (ATCC) (Manassas, VA, USA). Cells were maintained in 10% fetal bovine serum (FBS; Invitrogen, Carlsbad, CA, USA), Dulbecco's Modified Eagle Medium (DMEM; Invitrogen, Carlsbad, CA, USA) as well as 100 U/mL penicillin 100 µg/mL streptomycin (Sigma-Aldrich, St. Louis, MO, USA). Besides, cells were cultured in an incubator containing 5% CO₂ at 37°C. Cell culture medium was refreshed every two days.

Cell Transfection

Specific shRNA against DARS-AS1 was designed and synthesized by Invitrogen (Carlsbad, CA, USA). Negative control shRNA was also synthesized. To establish cell lines with stable knockdown of DARS-AS1, SKOV3 cells were transfected with DARS-AS1 or negative control shRNA through Lipofectamine 3000 reagent (Invitrogen, Carlsbad, CA, USA) according to the manufacturer's protocol.

RNA Extraction and Real-Time Quantitative Polymerase Chain Reaction (RT-qPCR)

Total RNA was extracted from cultured cancer cells or patients' tumor tissues using TRIzol reagent (TaKaRa Bio. Inc., Otsu, Shiga, Japan) and then reverse-transcribed to cDNA through reverse Transcriptase kit (TaKaRa Bio. Inc., Otsu, Shiga, Japan). RT-qPCR was performed three times in the following sequence: denaturation at 95°C for 1 min, followed by 40 cycles at 95°C for 15 s, 60°C for 30 s, and 72°C for 30 s. 2^{-ΔΔCt} method was utilized for calculating relative expression. The primer sequences used for RT-qPCR were as follows: DARS-AS1 forward: 5'-AGCCA-AGGACTGG-3' and reverse: 5'-CTGTACTGGTGG-3'; glyceraldehyde 3-phosphate dehydrogenase (GAPDH), forward: 5'-GACCCCAAGGCTGAGAAC-3' and reverse: 5'-TGGTCCAGTGG-3'.

Cell Proliferation Assay

Cell proliferation was monitored with Cell Counting Kit-8 (CCK-8) assays (Dojindo, Kumamoto, Japan). Briefly, 2 × 10³ SKOV3 cells were seeded

and incubated in 96-well plates. These cells were incubated with DARS-AS1 shRNA for 24 h and then incubated with 10 µM CCK-8 for 3 h. The absorbance was measured at 450 nm with an ELISA reader (Molecular Ascent, LabSystems, Helsinki, Finland).

Colony Formation Assay

To detect the long-term effect of DARS-AS1 on ovarian cancer cell proliferation, colony formation assays were conducted. 2 × 10³ SKOV3 cells/well were placed in 6-well plates and culture medium was replaced every 7 days. 7 days later, colonies were fixed with 75% ethanol for 30 min and stained with crystal violet. Colonies were photographed and counted.

EdU Labeling Assay (EdU Incorporation Assay)

EdU Kit (RiboBio, Guangzhou, China) was used for detecting cell proliferation of transfected cells. 2 × 10³ transfected cells were cultured in 96-well plates. Next, each well was added with 50 µM EdU labeling medium incubated for 2 h at 37°C. Finally, cells were stained with EdU working solution. Hoechst 33342 was used to label cell nuclei. Representative photograph was taken by a fluorescent microscope (Olympus, Tokyo, Japan).

Transwell Assay

Transwell chambers with 8 µm pores were provided by Corning (Corning, NY, USA). Cells were then seeded into the upper chambers of a 24-well plate. 20 % FBS-DMEM was added to the lower chamber of the culture inserts. After cultured for 24 h, these inserts were fixed with methanol for 30 min and stained by hematoxylin for 20 min. The number of migrated cells was counted by a light microscope (Olympus, Tokyo, Japan). For the transwell invasion assays, the membrane was precoated with 50 µL Matrigel (BD Biosciences, Franklin Lakes, NJ, USA). Next, the experiments were the same as described above.

Scratch Wound Assay

1.0 × 10⁴ cells were seed into a 6-well plate in each well. Three parallel lines were made on the back of each well. After growing to about confluent of 90%, cells were scratched with a pipette tip and cultured in a medium. Cells were photographed under a light microscope after 0 and 48 h. Each assay was independently repeated in triplicate.

Luciferase Reporter Gene Assay

DIANA LncBASE Predicted v.2 was used to predict the potential target gene and fragment sequences containing DARS-AS1 reaction sites. The DARS-AS1 3'-UTR wild-type (WT) sequence named DARS-AS1-WT was 5'-UCUCACUCAAAGUGGGAGU-3' and the mutant sequence of DARS-AS1 3'-UTR missing the binding site with miR-532-3p named DARS-AS1-MUT was 5'-UCUCACUCAAACACCCUCU-3'. Luciferase reporter gene assay kits (Promega, Madison, WI, USA.) were used to detect the luciferase activity of ovarian cancer cells. The luciferase reporter gene vector was constructed, and SKOV3 cells were transfected.

RNA Immunoprecipitation (RIP) Assay

To confirm the endogenous relationship between DARS-AS1 and miR-532-3p, RIP assay was carried out using the EZMagna RIP RNA-binding protein immunoprecipitation kit (Millipore, Billerica, MA, USA). Treated SKOV3 cells were collected and lysed using RIP lysis buffer containing protease inhibitor and RNase inhibitor. Cells were incubated with the RIP buffer containing magnetic beads coated with Ago2 and IgG (Millipore, Billerica, MA, USA). IgG acted as a negative control (input group). After incubation for 2 h at 4°C, co-precipitated RNAs were isolated and measured by RT-qPCR analysis.

Statistical Analysis

All statistical analyses were performed using Statistical Product and Service Solutions (SPSS,

21.0 (IBM Corp., Armonk, NY, USA). Independent-sample *t*-test was used to compare the difference between the two groups. Moreover, $p < 0.05$ was considered to indicate a statistically significant difference.

Results

DARS-AS1 Expression Level in Ovarian Cancer Tissues and Cell Lines

First, RT-qPCR was used for detecting DARS-AS1 expression in patients' tissues and 3 ovarian cancer cell lines. As a result, DARS-AS1 was significantly upregulated in tumor tissue samples (Figure 1A). DARS-AS1 level of ovarian cancer cells was higher than that of ISOE80 (Figure 1B).

Knockdown of DARS-AS1 Inhibited Cell Proliferation in SKOV3 Ovarian Cancer Cells

We chose SKOV3 cell line for the knockdown of DARS-AS1. Next, RT-qPCR was utilized for detecting the DARS-AS1 expression (Figure 2A). Moreover, CCK-8 assay showed that cell growth ability of SKOV3 cells was significantly repressed after DARS-AS1 was knocked down (Figure 2B). Colony formation assay showed that the colonies were significantly reduced after DARS-AS1 was knocked down (Figure 2C). Furthermore, EdU incorporation assay also showed that EdU positive cells were reduced after knockdown of DARS-AS1 in SKOV3 cells (Figure 2D).

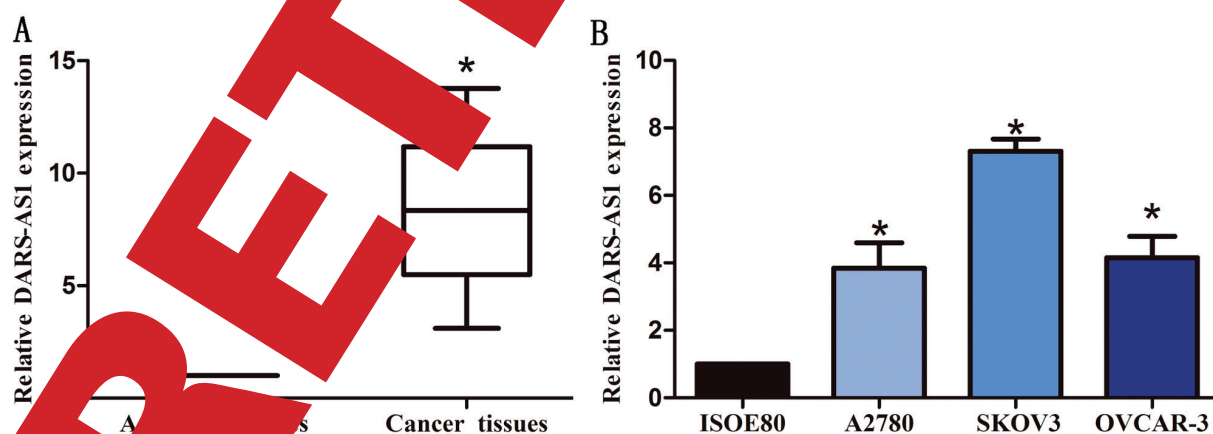


Figure 1 Expression levels of DARS-AS1 were increased in ovarian cancer tissues and cell lines. **A**, DARS-AS1 expression was significantly increased in the ovarian cancer tissues compared with adjacent tissues. **B**, Expression levels of DARS-AS1 relative to GAPDH were determined in the human ovarian cancer cell lines and ISOE80 by RT-qPCR. Data are presented as the mean \pm standard error of the mean. * $p < 0.05$.

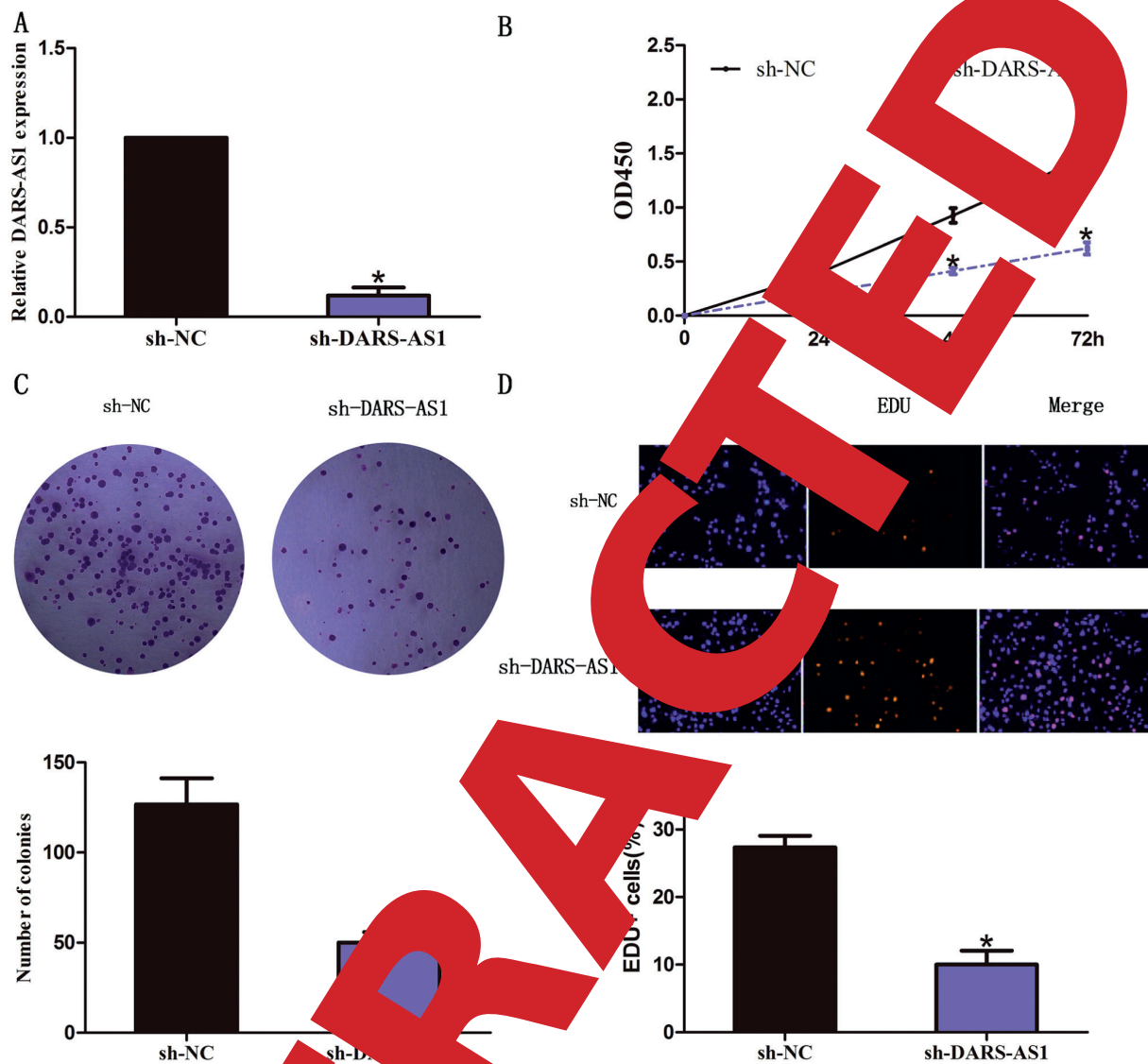


Figure 2. Knockdown of DARS-AS1 inhibited SKOV3 ovarian cancer cell proliferation. **A**, DARS-AS1 expression in SKOV3 ovarian cancer cells transfected with DARS-AS1 shRNA (sh-DARS-AS1) and negative control shRNA (sh-NC) was detected by RT-qPCR. GAPDH was used as internal control. **B**, CCK-8 assay showed that knockdown of DARS-AS1 significantly inhibited cell growth in SKOV3 ovarian cancer cells. **C**, Colony formation assay showed that the number of colonies was reduced after knockdown of DARS-AS1 in SKOV3 cells (magnification: 10×). **D**, EdU incorporation assay showed that EdU positive cells were reduced after knockdown of DARS-AS1 in SKOV3 cells. The results represent the average of three independent experiments (mean ± standard error of the mean). * $p < 0.05$, as compared with the control cells.

Knockdown of DARS-AS1 Inhibited Cell Migration and Invasion in SKOV3 Ovarian Cancer Cells

Transwell assay showed that the migrated number of SKOV3 cells was significantly decreased after DARS-AS1 was knocked down (Figure 3A). Boyden chamber assay showed that the number of migrated cells was significantly reduced after DARS-AS1 was knocked down (Figure 3B).

Transwell invasion assay showed that the number of invaded cells was significantly reduced after DARS-AS1 was knocked down (Figure 3C).

The Interaction Between miR-532-3p and DARS-AS1 in Ovarian Cancer

Starbase v2.0 (<http://starbase.sysu.edu.cn/mir-LncRNA.php>) was used to find the miRNAs that contained complementary base with DARS-AS1.

MiR-532-3p was selected from these miRNAs, which were interacted with DARS-AS1 (Figure 4A). RT-qPCR assay showed that the expression of miR-532-3p was higher in DARS-AS1/shRNA cells than that in control cells (Figure 4B). Furthermore, the luciferase assay revealed that co-transfection of DARS-AS1-WT and miR-532-

3p largely decreased the luciferase activity, while co-transfection of DARS-AS1-WT and miR-532-3p had no effect on luciferase activity either (Figure 4C). Meanwhile, RIP assay also identified that miR-532-3p could be remarkably enriched in the DARS-AS1 group compared with control group (Figure 4D).

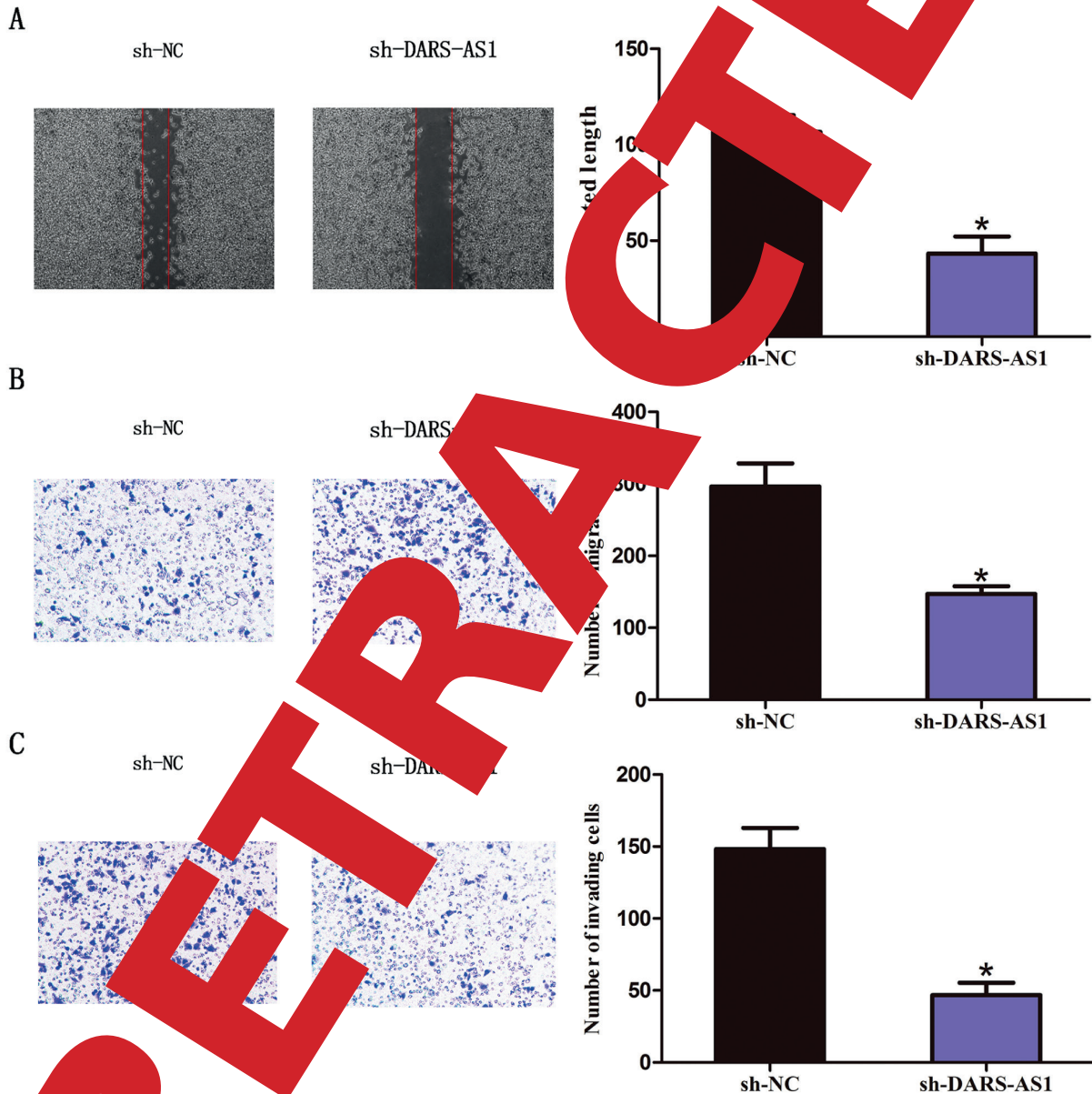


Figure 3. Knockdown of DARS-AS1 inhibited SKOV3 ovarian cancer cell migration and invasion. **A**, Scratch wound assay showed that migrated length of SKOV3 cells was significantly decreased after DARS-AS1 was knocked down (magnification: 40×). **B**, Transwell assay showed that number of migrated cells was reduced after knockdown of DARS-AS1 in SKOV3 cells (magnification: 40×). **C**, Transwell invasion assay showed that number of invaded cells was reduced after knockdown of DARS-AS1 in SKOV3 cells (magnification: 40×). The results represent the average of three independent experiments (mean \pm standard error of the mean). * $p < 0.05$, as compared with the control cells.

Discussion

Ovarian cancer is one of the most fatal gynecologic malignancies and the fifth leading cause of death in cancer. Most ovarian cancer cases are diagnosed at a late stage due to atypical or absent symptoms at early stage. Plenty of lncRNAs have been revealed to play important roles in oncogenesis and progression of ovarian cancer. For example, silencing of lncRNA MNX1-AS1 suppresses cell proliferation and migration of ovarian cancer, which may be a potential target for ovarian cancer⁸. lncRNA TUG1 promotes epithelial ovarian cancer cell proliferation and invasion via the WNT/beta-catenin pathway⁹. lncRNA BA-CE1-AS inhibits the proliferation and invasion of ovarian cancer stem cell and functions as a novel target for treating ovarian cancer¹⁰. Besides, lncRNA ElncRNA1 functions as an oncogene in the proliferation of epithelial ovarian cancer cells which is upregulated by estrogen¹¹.

lncRNA DARS antisense RNA (DARS-AS1), located in 2q21.3, is a novel lncRNA discovered recently. Its role in cancer development has not been studied. Our study first uncovered the association between DARS-AS1 and ovarian cancer progression. The results demonstrate that DARS-AS1 was upregulated in ovarian cancer samples and cells. Besides, after DARS-AS1 was knocked down, ovarian cancer cell proliferation was inhibited. Meanwhile, after DARS-AS1 was knocked down, ovarian cancer cell migration and invasion were found to be inhibited. The data indicated that DARS-AS1 promoted the origin of ovarian cancer and might act as an oncogene.

Bioinformatics software predicted miR-532-3p as a possible target of DARS-AS1. MiR-532-3p functions as a tumor suppressor in a diverse of cancers by regulating various biological processes^{12,13}. For example, miR-532-3p is downregulated in bladder cancer and inhibits tumor development through targeting Wnt signaling pathway¹⁴. MiR-

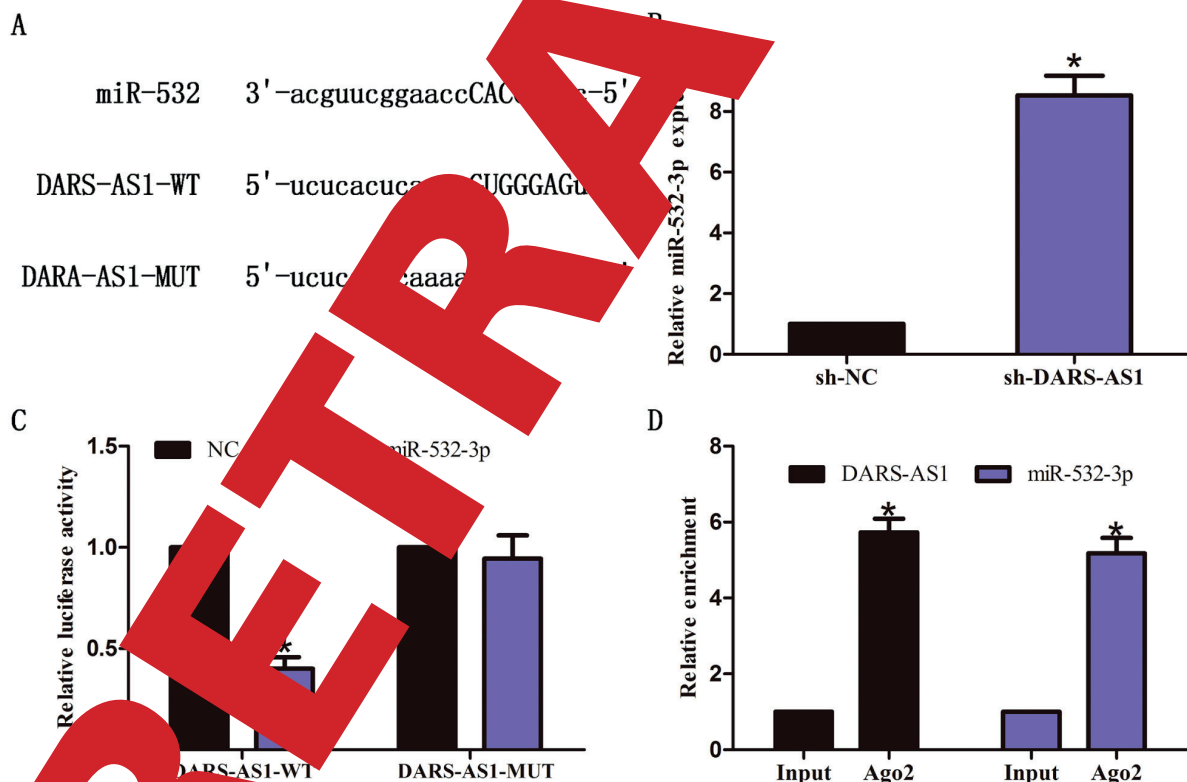


Figure 4. Relationship between DARS-AS1 and miR-532-3p. **A**, The binding sites of miR-532-3p on DARS-AS1. **B**, miR-532-3p expression was increased in sh-DARS-AS1 group compared with negative control shRNA (sh-NC) group. **C**, Co-transfection of miR-532-3p and DARS-AS1-WT strongly decreased the luciferase activity, while co-transfection of negative control and DARS-AS1-WT did not change the luciferase activity. **D**, RIP assay identified that DARS-AS1 and miR-532-3p were significantly enriched in Ago2-containing beads compared to input group. The results represent the average of three independent experiments. Data are presented as the mean \pm standard error of the mean. * $p < 0.05$.

532-3p could induce cell apoptosis of lung adenocarcinoma cancer¹⁵. Recently, high expression of miR-532-5p is associated with better prognosis in ovarian cancer¹⁶. Besides, miR-532-3p was reported to repress cell growth and invaded ability in ovarian cancer¹⁷.

In the present study, miR-532-3p could be directly targeted by DARS-AS1 through a luciferase assay. Moreover, miR-532-3p expression could be upregulated through knockdown of DARS-AS1. Furthermore, miR-532-3p was significantly enriched by DARS-AS1 through RIP assay. All these results showed that DARS-AS1 could work as a miR-532-3p sponge in ovarian cancer.

Conclusions

We demonstrated that DARS-AS1 could enhance ovarian cancer cell proliferation and metastasis through sponging miR-532-3p. These findings implied that lncRNA DARS-AS1 could act as a prospective therapeutic target for ovarian cancer.

Conflict of Interest

The Authors declare that they have no conflict of interest.

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