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MicroRNA-185 inhibits cell proliferation and epithelial-mesenchymal transition in hepatocellular carcinoma by targeting Six2

S.-M. ZHU¹, C.-M. CHEN², Z.-Y. JIANG³, B. YUAN⁴, M. JI⁵, F.-H. WU⁶,



Corresponding Authors: Ji Min, MD; e-mail: jimin0518@126.com Fei-hua Wu, MD; e-mail: wufeihua1961@126.com Jian Jin, MD; e-mail: ausword@126.com croRNA-185 (miR-185) is one of the most commonly down-regulated miRNAs in several cancers including HCC, prostate carcinoma, breast cancer, and gastric cancer, suggesting that miR-185 may play an important role in carcinogenesis⁸⁻¹¹. In addition, the potential anti-cancer properties of miR-185 have been demonstrated through the regulation of critical cellular behaviors such as cell proliferation, apoptosis, cell migration, and invasion¹²⁻¹⁴.

Epithelial-mesenchymal transition (EMT) is a complex, reversible process that induces epithelial cells to transform to a mesenchymal phenotype¹⁵. Advances in research have highlighted the role of EMT in regulating the cellular process of liver carcinoma, but the molecular mechanisms regulating EMT remain poorly understood¹⁶.

In this current study, we focused on miR-185 in HCC and its relationship with EMT, and our results would complement the current knowledge of miR-185 in liver cancer.

Materials and Methods

Cell Culture

A normal liver cell line (LO2) and four an HCC cell lines (HepG2, HuH7, SNU-38 SNU-449) were purchased from the Amer Type Culture Collection (Man /A, US ified E Cells were cultured in Dulb gle's Medium (Invitroger arlsbad A, USA) bovine um (Sigsupplemented with 10% ma, St. Louis, MO, USA) a A). The cells tomycin (Sigma, S ouis, M 7°C in a h were maintained ied air atcarbon dio mosphere cont

Cell Proferation As

Cell diferation was m ed using an MTT -dimethylthiazol-2-, 1-2, 5-diphenyl-(3 - 1)nide) assay. Briefly, cells at a lium) te 0⁴ (cellstwell) were seeded in 96den of MTT (5 mg/ml) was well p Then 10 d incubated in dark at $37\Box$ ed to vel e was measured with a mih. Ab ate reader a wavelength of 490 nm (Bioules, CA, USA).

Cell Migration Assay

bell migration was measured using an *in vitro* and-healing assay. Cells were cultured in 6well plates overnight. Experimental wounds were made by dragging a rubber policeman (Fisher Scientific, Hampton, VA, USA) across the cell culture. The cultures were rinsed with PBS and placed in fresh quiescence medium. Three wounds were created for each specimer relative distance traveled by the cells us determined.

RNA Extraction and Real-Time

Total miRNA was extract from the ed cells using an RNeasy mi Kit (QIA Hilden, Germany) acc ng to the manufac er's protocol. cDNA nth ed from total verse tra RNA using a Taq n m. ription kit (Appli oiosysten. ster , CA, USA). miR-1 pression wa ed using ngMan Mik Assay Kit a miRNA rlsbad, CA, USA). PCR (Applied b. osystem reactions were perform n an ABI 7500 Real-Tir System (App **Biosystems**, Carls-CA, USA) with the following conditions: C, 10 min for 1 cycle, then 95°C, 15 sec, 1 min for ycles. The U6 small nuclear as used a loading control. The mRNA of S was measured using real-time exp PCR in 7500 Real-Time PCR System oplied Biosystems, Carlsbad, CA, USA), with as a control.

Plasmid Construction and Luciferase Activity Assay

For the fluorescent reporter assay, the following primers were used to amplify the 3'UTR of the Six2 gene: forward primer 5'- CTTGGTAC-CGAGCTCTCCTAGAGCTCTGTTCGCCT-3'; reverse primer 5'- -TGCTGGATATCTGCA CGAACATTCACATGAGGGCG-3'.

A plasmid containing the Six2 3'UTR and a fluorescent reporter was constructed. HepG2 cells were seeded into 48-well plates and co-transfected with a negative control, a miR-185 mimic or a miR-185 inhibitor. Luciferase activity was measured using a dual-luciferase reporter assay system (Promega, Madison, WI, USA). Firefly luciferase activity was normalized to renilla activity.

Western Blot

Cells were lysed using RIPA buffer and total cell extracts were subjected to sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE). For immunoblotting, the membranes were blocked with 5% non-fat milk in Tris-buffered saline then incubated with antibodies against Six2, E-cadherin, vimentin, and GAPDH (Santa Cruz, CA, USA). The membranes were washed thrice and incubated with horseradish peroxidase-conjugated secondary antibody. Protein bands were detected using an enhanced chemiluminescence (ECL) kit (Thermo Scientific, Waltham, MA, USA) and a ChemiGenius bioimaging system (Syngene, Frederick, MD, USA).

Statistical Analysis

All data are presented as mean \pm SD. For comparisons between two groups, statistical significance was determined using the Student's *t*-test. Comparisons between three or more groups were performed using Least-significant difference (LSD) following analysis of variance (ANOVA). A value of p < 0.05 was considered significant.

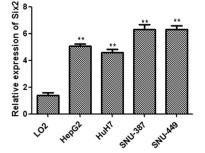
Results

Down-Regulation of miR-185 in HCC Cells

miR-185 is known to play a central role in various types of cancers including HCC. However, the biological function of miR-185 has not been fully elucidated. In the current study, we web-based target analysis tools like Targer camiRanda, and pictar to predict potential uset genes that may be regulated by miR-185. this analysis, Six2 was identified as a potential target of miR-185. We then went on to examine the expression of miR-185 and Six2 in four by man HCC cell lines (HepG2, HuH7, S and SNU-449) and a normal human li cell line (LO2) by real-time PCR. Our reg s revealed that miR-185 expression was si antly reduced in HCC cells compared with liver cells (Figure 1A). Converse we foun the mRNA and protein expr on of Six2 v are 1B nd C). Th creased in HCC cells results imply that mik ne of the upay six2 exp stream molecules ut m sion in liver cancer.

miR-185 ns ell Growth Migration in Ho

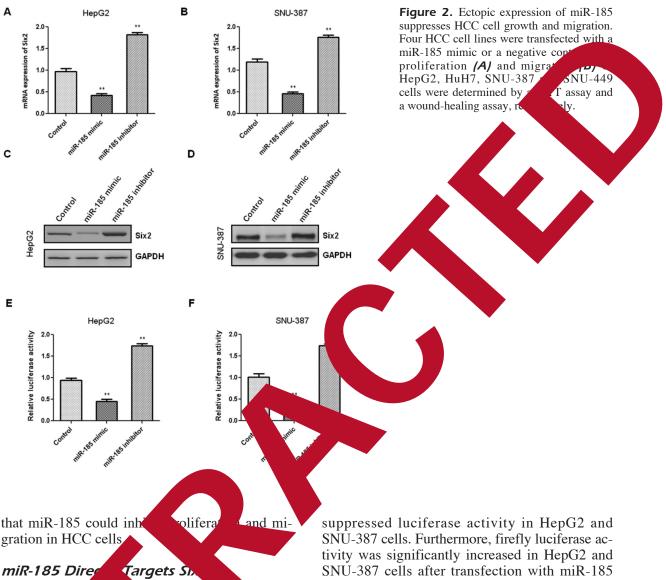
ical role of miR-185 To investigate the Il growth and ration, four HCC in ines were transiently transfected with either iR-185 mimicar a negative control. MTT asrmed to determine the effect were then p proliferation. Our results 0 2-185 on un_re ation of miR-185 significantly sho srowth of HCC cells (Figure 2A). suppres foreover, we found that ectopic expression of reduced the metastatic potential of is (Figure 2B). These results indicate



1. Negative correlation between miR-185 and Six2 expression in HCC cells. (A) The expression of miR-185 was determed in four HCC cell lines (HepG2, HuH7, SNU-387 and SNU-449) and a normal human liver cell line (LO2) by real-time PCR. (B and C) mRNA and protein levels of Six2 were assessed by real-time PCR and Western blot, respectively. *p < 0.05, **p < 0.01.

Six2

GAPDH



in HCC Cell To determ ix2 is directly regulate whe ed by mil 185, we tran d HepG2 and SNU-387 ce vith a miR-185 h or inhibitor, then d Six2 mRNA expression using real-time mea P Qur reg show that miR-185 up-regulatio y repressed Six2 mRNA expression control hereas inhibition of miRcompa ased Six2 levels in HepG2 in resu SNU-3 (Figure 3A and B). Western analysis and showed that over-expression of sulted in a markedly decrease in Six2 reas a reduction in miR-185 remarkbly increased Six2 protein levels in HepG2 and -387 cells (Figure 3C and D). We then pered fluorescent reporter assays to determine whether Six2 is a direct target of miR-185. As predicted, miR-185 over-expression significantly

inhibitor (Figure 3E and F). These results indicate that Six2 is a direct functional target of miR-185.

Determination of the Phenotypes of Four HCC Cell Lines

EMT progression is characterized by the loss of expression of epithelial cell junction proteins, such as E-cadherin, and a gain of mesenchymal marker expression, such as vimentin. In this study, we found that E-cadherin was primarily expressed in HepG2 and HuH7 cells, but was absent in SNU-387 and SNU-449 cells. In contrast, vimentin expression was higher in SNU-387 and SNU-449 cells than in HepG2 and HuH7 cells (Figure 4). These findings indicate that HepG2 and HuH7 cells exhibit an epithelial character while SNU-387 and SNU-449 cells have a mesenchymal phenotype.

Up-regulation of miR-185 Reverses EMT in HCC Cell lines

We used epithelial (HepG2 and HuH7) and mesenchymal (SNU-387 and SNU-449) cell lines to investigate the effect of miR-185 on EMT. We transfected a miR-185 mimic into HepG2 (epithelial) and SNU-387 (mesenchymal) cells and showed that ectopic miR-185 expression led to increased E-cadherin expression in the HepG2 epithelial cells (Figure 5A and B) and suppressed vimentin expression in SNU-387 mesenchymal cells (Figure 5C and D). Thus, these results demonstrate that up-regulation of miR-185 caused a reversal of EMT in HCC cell lines.

Discussion

To date, a large body of data has shown that miRNAs exhibit altered expression levels in multiple types of cancer, where they play key roles in tumor cell behavior through the regulation of target gene expression^{17,18}. Thus, the key to understanding miRNA function is to identify their functional targets. In this study, we demonstrated that miR-185 suppresses cell growth approgression in HCC cells by regulation $\delta tx2$ expression.

miR-185 is one of the most we ed miR-NAs in cancer biology. Several tigations^{13,19-21} have demonstrate nat mik cts as a tumor suppressor in stric cancer, and colorectal can cancer, prostate carcing However, although the essive effects r su its preci of miR-185 have en n role ed. in tumorigenesi s not bee eluc s encode tra Homeoboy 1 factors that act as velopment, of embryoni where they are invo in cell growth and differentiation²². It has also shown that homeobox gep an essential ro. umor initiation and ression²³. Six2, a member of the SIX family nomeobox gent, has been given much attene roles in cancer. Early studdue to its di

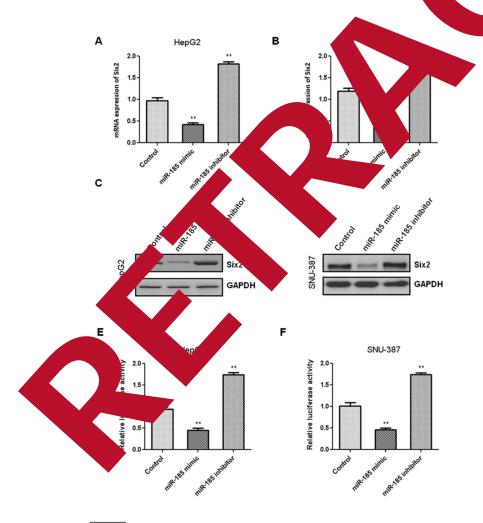


Figure 3. Six2 is a target of miR-185 in HCC cells. HepG2 and SNU-387 cells were transfected with a negative control, a miR-185 mimic or a miR-185 inhibitor. mRNA *(A* and *B)* and protein *(C* and *D)* expression of Six2 were determined by real-time PCR and Western blot, respectively. *(E* and *F)* Luciferase activity was assessed 48 h after transfection using a dual luciferase reporter assay, normalized to a renilla control. *p < 0.05, **p < 0.01.

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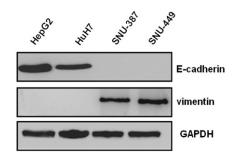


Figure 4. Expression of EMT-related markers in four HCC cell lines. Western blot analysis was performed using antibodies against E-cadherin (epithelial marker) and vimentin (mesenchymal marker) in HepG2, HuH7, SNU-387 and SNU-449 cell lines. GAPDH was used as an internal control.

ies^{24,25} suggested that Six2 regulates kidney development through the suppression of premature nephrogenesis and maintenance of mesenchymal progenitor populations. The pro-proliferative and pro-migratory effects of Six2 have been reported in the pathogenesis of several types of cancers, including breast cancer, nephroblastomas, and renal clear cell carcinomas^{26,27}. In the current repoobserved the concomitant down-regulation of miR-185 and up-regulation of *Six2* in HC ulls, suggesting that Six2 could be a potential of miR-185. As expected, transfection miR-185 significantly down-regulated the expression of *Six2*. Also, luciferase activity assays further confirmed the direct regulation of Six2 by miR-185 in HCC cells.

Functional researches have sug ed that miR-185 could induce cell cycle st and rer¹⁹, colpress cell proliferation in gastric orectal cancer²⁰, non-small-ce¹¹ lung 28 and glioma²⁹. Moreover, miR-1 nas also ported to be involved in astasis in col cancer and glioma^{29,30} sistent with these vious works, our result e that the ec ons nificant' topic expression mik upand mig ells. pressed cell gro n H hogenetic EMT is a n which Il polarity, cells lose atures such and gain Lesench properties such as increased motility³¹. Ac on of EMT enhances the e properties ncer cells. During F , the loss of cell-cell adhesion is a critical ion and metastases, and is ofin tumor inva ccompanie the down-regulation of Ein³². The thelial molecule E-cadherin с mpor role in epithelial polarization pla nor suppressor in many types of and act ncers³³. It has been suggested that decreased Eexpression is a major hallmark of EMT, closely associated with the malignant progression of HCC³⁴. A recent study in breast

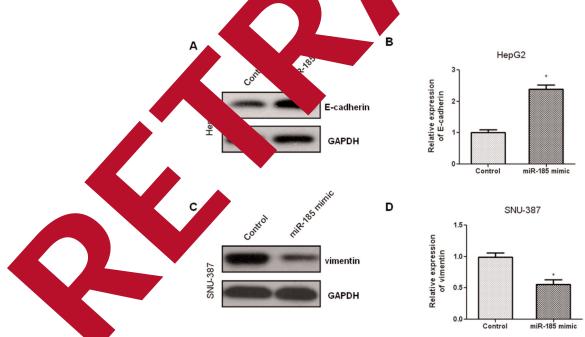


Figure 5. Up-regulation of miR-185 reversed EMT in HCC cell lines. HepG2 and SNU-387 cells were transfected with a miR-185 mimic or a negative control. 72 h after transfection, western blot was performed to determine the expression of E-cadherin (A and B) and vimentin (C and D) in HepG2 and SNU-387 cells, respectively. p < 0.05, p < 0.01.

cancer suggested that Six2 functions as a regulator of metastasis through the epigenetic regulation of E-cadherin, implicating a regulatory role for Six2 in EMT progression²⁶. In our paper, we found that up-regulation of miR-185 significantly increased the expression of E-cadherin in epithelial (HepG2) cells and reduced the expression of vimentin in mesenchymal (SNU-387) cells. Taken together, these results indicate that up-regulation of miR-185 could reverse EMT in liver cancer cells via the regulation of Six2.

Conclusions

We identified miR-185 as a potential regulator of EMT in HCC cells. In addition, the up-regulation of miR-185 or the inhibition of Six2 could be potential therapeutic strategies for the treatment of HCC.

Conflict of Interest

The Authors declare that they have no conflict of interests.

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