

MiR-101 promotes nasopharyngeal carcinoma cell apoptosis through inhibiting Ras/Raf/MEK/ERK signaling pathway

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Abstract. – **OBJECTIVE:** Extra-cellular signal regulated kinase (ERK)/mitogen activated protein kinase (MAPK) signaling pathway is widely involved in cell proliferation and apoptosis. MAPK kinase 1 (MEK1) is the upstream protein kinase of ERK that can activate ERK/MAPK signaling pathway. microRNA-101 (MiR-101) down-regulation is found to be associated with nasopharyngeal carcinoma (NPC) pathogenesis. Bioinformatics analysis shows the complementary targeted relationship between miR-101 and the 3'-UTR of MEK1 mRNA. This study explores the role of miR-101 in regulating MEK1 expression, ERK/MAPK signaling pathway activation, and NPC pathogenesis.

MATERIALS AND METHODS: Dual luciferase assay confirmed the targeted relationship between miR-101 and MEK1. MiR-101 and MEK1 expressions were compared in inflammatory nasopharynx tissue and NPC tissue. MiR-101, MEK1 phosphorylated ERK 1/2 (p-ERK1/2), and survivin expressions in NP69, CNE-1, HONE1, and C666-2 cell lines were detected. NPC cell line C666-2 was cultured *in vitro* and divided into four groups, including NC, miR-101-NC and si-MEK1. Cell apoptosis was determined by flow cytometry. Cell proliferation was evaluated by EdU staining.

RESULTS: MiR-101 targeted and inhibited MEK1 expression. MiR-101 was significantly down-regulated while MEK1 was significantly elevated in NPC tissue compared with inflammatory nasopharynx tissue. MiR-101 was markedly declined, whereas p-ERK1/2, and survivin were apparently increased in CNE-1, HONE1, and C666-2 cells compared with NP69 cells. MiR-101 mimic and/or si-MEK1 transfection significantly reduced MEK1, p-ERK1/2, and survivin levels, inhibited cell proliferation, and enhanced cell apoptosis.

CONCLUSIONS: Down-regulation of miR-101 was related to NPC pathogenesis. MiR-101 elevation suppressed NPC cell proliferation and promoted apoptosis through targeted inhibiting MEK1 expression to alleviate ERK/MAPK signaling pathway and survivin expression.

Key Words: miR-101, MEK1, Ras/Raf/MEK/ERK signaling pathway, Apoptosis, Proliferation, Nasopharyngeal carcinoma.

Introduction

Nasopharyngeal carcinoma (NPC) is a kind of epithelial malignant tumor occurred in the top and side of nasopharynx. It is a common type of malignant tumor in South-East Asia and South China with an incidence at 40-60/100,000. The incidence of NPC has been top in head and neck malignant tumor. NPC may cause various symptoms, including nasal congestion, nose blood, diplopia, headache, and cranial nerve compression based on different severity degree, leading to serious threat to the quality of life and health^{2,3}. Therefore, in-depth study of NPC pathogenesis and exploration of the abnormal change of molecule expression are of great significance for NPC early diagnosis, therapeutic efficacy, and prognosis. Extra-cellular signal regulated kinase (ERK)/mitogen activated protein kinase (MAPK) signaling pathway that widely expresses in various tissues and cells, regulates multiple biological processes, including cell proliferation, cycle, apoptosis, migration, and invasion^{4,5}. ERK/MAPK signaling pathway excessive activation induces cell abnormal proliferation, apoptosis, and differentiation, which are closely related to tumor pathogenesis, progression, and metastasis⁶⁻⁸. MAPK kinase 1 (MEK1) phosphorylates the residue of Tyr/Thr on ERK protein to activate ERK/MAPK signaling pathway. MEK1 elevation plays a crucial promoting role in a variety of tumor occurrence and development⁹⁻¹¹. It was showed that MEK1 expression and function enhancement are related to NPC cell proliferation, apoptosis, and drug resistance¹²⁻¹⁴. microRNA (MiRNA) is a type of

endogenous single stranded non-coding RNA at the length of 22-25 nt discovered from eukaryote. It plays a degrading or inhibiting role on mRNA by binding with the 3'-UTR, thus participating in cell proliferation, differentiation, and migration. The role of miRNA in tumorigenesis receives more and more attention¹⁵⁻¹⁷. Numerous studies revealed that miR-101 significantly reduced in NPC tissue and cells, suggesting that miR-101 may play a tumor suppressor role in NPC occurrence and development^{18,19}. Bioinformatics analysis shows the complementary targeted relationship between miR-101 and the 3'-UTR of MEK1 mRNA. This study explores the role of miR-101 in regulating MEK1 expression, ERK/MAPK signaling pathway activation, and NPC pathogenesis.

Materials and Methods

Main Reagents and Materials

Human NPC cell line CNE-1, HONE1, and C666-1, and human nasopharynx epithelial cell line NP69 were purchased from Xinyu Biotech co., Ltd (Shanghai, China). Dulbecco's Modified Eagle Medium (DMEM) and Roswell Park Memorial Institute 1640 (RPMI-1640) medium were bought from Lonza Inc. (Allendale, NJ, USA). Fetal bovine serum (FBS), penicillin, and streptomycin were bought from Gibco BRL, Grand Island, NY, USA). MiniBEST Universal RNA Extraction Kit, PrimeScript RT reagent Kit, and SYBR Green were obtained from Takara (Dalian, China). Lipofectamine 2000 was purchased from Invitrogen Life Technologies (Carlsbad, CA, USA). MiR-101 mimic, miR-101 inhibitor, and miR-NC were bought from Genescript (Guangzhou, China). Mouse anti-human MEK1, p-ERK1/2, survivin, and β -actin primary antibodies were obtained from Abcam biotech. (Cambridge, MA, USA). Horse radish peroxidase (HRP) conjugated secondary antibody was derived from NeoBioscience (Shenzhen, China). Luciferase reporter gene vector pGLUC was bought from Ambion Inc. (Austin, TX, USA). Luciferase activity detection kit was purchased from Promega (Madison, MI, USA). Western blotting (WB), RIPA and bicinchoninic (BCA) protein assay kit, and BeyoECL Plus enhanced chemiluminescent (ECL) reagent were obtained from Beyotime (Haimen, China). Annexin V-FITC/PI cell apoptosis detection kit was derived from BD Biosciences (Franklin Lakes, NJ, USA). EdU cell proliferation detection kit was purchased from Molecular Probes (Eugene, OR, USA).

Clinical Information

A total of 65 NPC patients who received treatment in the 2nd Affiliated Hospital of Fujian Medical University between March 2016 and December 2016 were enrolled, including 29 males and 36 females with mean age at 52.3 ± 11.7 years old. All the subjects received surgery for the first time without preoperative radiotherapy or chemotherapy. Tumor tissue was resected and stored at -80°C . Another 40 cases of patients suffered from chronic nasopharyngitis were selected as control, including 18 males and 22 females with average age at 50.9 ± 10.8 years old (Table I). No significant difference was observed on age and gender between two groups (Table I, $p > 0.05$). The experimental protocol has been pre-approved by the Ethical Committee of the 2nd Affiliated Hospital of Fujian Medical University and written consents have been obtained from all patients and healthy volunteers.

Cell Culture

CNE-1, HONE1, C666-1, and NP69 cells were cultured in RPMI 1640 medium containing 10% FBS and 1% penicillin-streptomycin. The cells were passaged at 1:4.

Luciferase Reporter Gene Assay

PCR products containing the full-length of MEK1 gene 3'-UTR segment were cloned to pLUC after endonuclease SacI and XbaI cloning. Next, it was named as pLUC-MEK1-wt (or pLUC-MEK1-UTR-mut) and co-transfected to HEK293T cells using Lipofectamine 2000 together with miR-101 mimic (miR-101 inhibitor or miR-NC). The luciferase activity was detected according to the Stop&Glo Luciferase Assay manual after cultured for 48 h.

Cell Grouping and Transfection

C666-1 cells were cultured *in vitro* and divided into four groups, including miR-NC, miR-101 mimic, si-NC and si-MEK1 groups. MiR-NC or miR-101 mimic at 30 nmol/L and si-NC or si-MEK1 at 10 nmol/L were incubated with Lipofectamine at 10 μL at room temperature for 15-20 min. Then, they were added to the cells and incubated for 6 h. After changing the medium and incubation for 72 h, the cells were collected for the following experiments.

Quantitative Real-time PCR (qRT-PCR)

Total RNA was extracted using MiniBEST Universal RNA Extraction Kit and adopted for PCR reaction by PrimeScriptTM RT reagent Kit

(TaKaRa Bio. Inc., Otsu, Japan). The PCR reaction was composed of 95°C pre-denaturation for 15 min, followed by 40 cycles of 95°C for 15 s, 60°C for 30 s, and 72°C for 30 s. Real-time PCR was performed on Applied Biosystems QuantStudio 3 connect (PE Applied Biosystems, Foster City, CA, USA) to test the relative expression.

Western Blot

Total protein was extracted by RIPA from cells. A total of 40 µg protein was separated by sodium dodecyl sulfonate-polyacrylamide gel electrophoresis (SDS-PAGE) and transferred to membrane. Next, the membrane was blocked by 5% skim milk and incubated in primary antibody at 4°C overnight (MEK1, p-ERK1/2, survivin, and β-actin at 1:3000, 1:1000, 1:2000, and 1:10000, respectively). Then, the membrane was incubated in HRP labeled secondary antibody (1:30000) for 1 h after washed by phosphate buffer solution-tween 20 (PBST) for three times. At last, the protein expression was detected by BeyoECL Plus (Beyotime Biotech., Shanghai, China).

Cell Apoptosis Detection

The cells were digested by trypsin and resuspended in binding buffer. Next, the cells

were incubated in 5 µl Annexin V-FITC and 5 µl propidium iodide (PI). At last, the cells were tested on Beckman Coulter CytoFLEX flow cytometry to evaluate cell apoptosis.

Edu Staining

Cell proliferation was assessed by Click-iT® Edu Alexa Fluor 488 Flow Cytometry Assay Kits. The cells were added with 1 µM Edu solution for 2 h. After they were incubated for 48 h, the cells were digested and collected. After washed by PBS, fixed and permeabilized, the cells were incubated in reaction liquid containing Alexa Fluor 488 at room temperature avoiding light for 30 min. The cells were washed and tested by Beckman Coulter CytoFLEX flow cytometry (Beckman Coulter Inc., Brea, CA, USA).

Statistical Analysis

All data analyses were performed on SPSS (SPSS Inc., Chicago, IL, USA) software. The measurement data were depicted as mean ± standard deviation and compared by *t*-test. *p* < 0.05 was considered as statistical significance.

Results

MiR-101 Targeted Inhibited MEK1 Expression

Bioinformatics analysis showed the targeted binding site between miR-101 and 3'-UTR of MEK1 mRNA (Figure 1A). Dual luciferase assay revealed that miR-101 mimic transfection significantly declined, while miR-101 inhibitor transfection significantly elevated the relative luciferase activity of HEK293 cells, indicating the regulatory relationship between miR-101 and MEK1 mRNA (Figure 1B).

MiR-101 Down-regulated, While MEK1 Increased in NPC Tissue

qRT-PCR revealed that miR-101 significantly declined, whereas MEK1 mRNA obviously up-regulated in NPC tissue compared with inflammatory nasopharyngeal tissue (Figure 2A). MiR-101 was markedly correlated with tumor size and clinical staging. Western blot demonstrated that MEK1 protein apparently enhanced in NPC tissue compared with inflammatory nasopharyngeal tissue (Figure 2B).

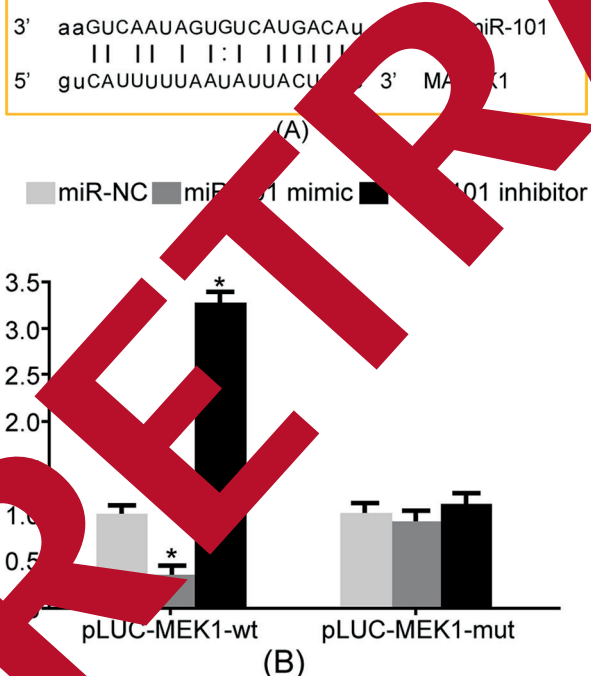


Figure 1. MiR-101 targeted inhibited MEK1 expression. (A) The binding site between miR-101 the 3'-UTR of MEK1 mRNA. (B) Dual luciferase assay. **p* < 0.05, compared with mimic NC.

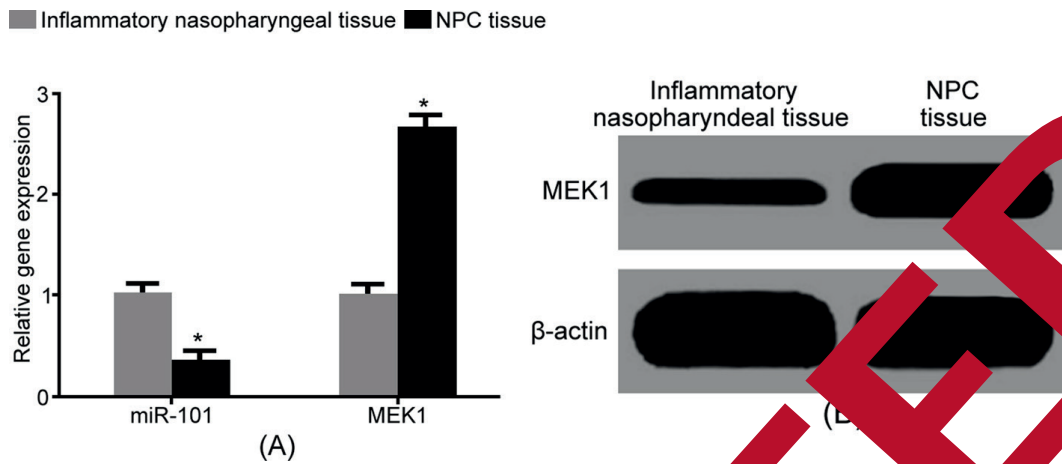


Figure 2. MiR-101 down-regulated, while MEK1 increased in NPC tissue. (A) qRT-PCR detection of miR-101 expression. (B) Western blot detection of MEK1 protein expression. * $p < 0.05$, compared with inflammatory nasopharyngeal tissue.

MiR-101 Reduced, Whereas MEK1 Enhanced in NPC Cells

qRT-PCR revealed that miR-101 significantly declined, whereas MEK1 mRNA obviously up-regulated in NPC cell lines CNE-1, HONE1, and C666-1 compared with NP69 cells (Figure 3A and B). Western blot demonstrated that MEK1 protein levels markedly increased in NPC cell lines CNE-1, HONE1, and C666-1 compared with NP69 cells (Figure 3C).

MiR-101 Over-expression in NPC Cells Down-regulation Suppressed NPC Cell Proliferation and Induced Cell Apoptosis

MiR-101 mimic and/or si-MiR-101 significantly reduced MEK1, ERK1/2, and survivin levels in C666-1 cells (Figure 4A and B), attenuated cell proliferation (Figure 4C), and enhanced cell apoptosis (Figure 4D).

Discussion

Globally, there are more than 100,000 patients diagnosed as NPC each year. More than 80% new cases come from South East Asia, China, and other Asian countries²¹. The morbidity peak is between 40-50 years old. It may cause nasal congestion, nose discharge, nasal fullness, hearing loss, diplopia, and facial numbness, and may induce cranial nerve because of peripheral tissue infiltration, which seriously threatens to life and health^{2,3}.

MAPK signaling pathway is an important signal transduction system that widely exists in eukaryotes. It regulates various target genes expression and activation mediated by intracellular receptor tyrosine kinase, G-protein coupled receptor, and cytokine receptor under the effect of cytokines, growth factors, neurotransmitter, and G-protein coupled receptor, thus participating in

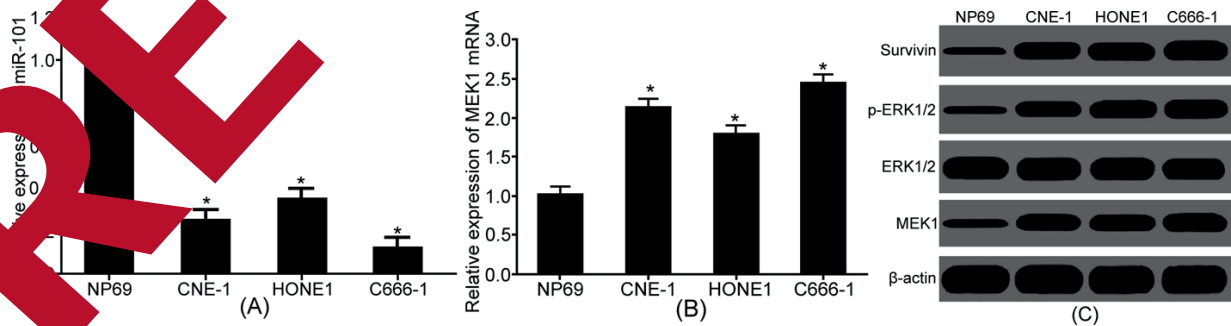


Figure 3. MiR-101 reduced, whereas MEK1 enhanced in NPC cells. (A) qRT-PCR detection of miR-101 expression. (B) qRT-PCR detection of MEK1 mRNA expression. (C) Western blot detection of protein expression. * $p < 0.05$, compared with NP69 cells.

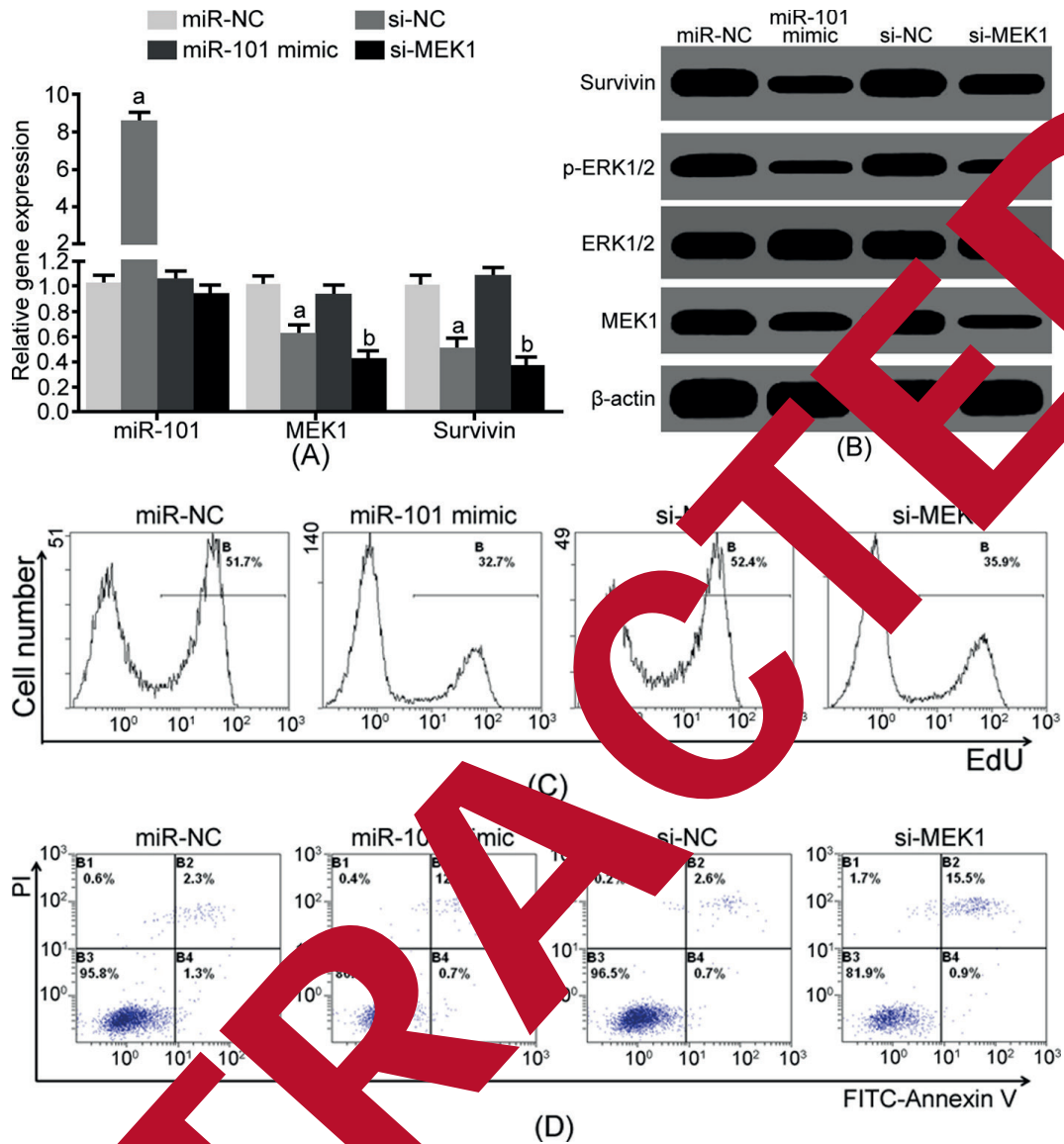


Figure 4. MiR-101 overexpression or MEK1 down-regulation suppressed NPC cell proliferation and induced cell apoptosis. (A) qRT-PCR detection of gene expression. (B) Western blot detection of protein expression. (C) EdU staining detection of cell proliferation. (D) Flow cytometry detection of cell apoptosis. ^a*p* < 0.05, compared with miR-NC; ^b*p* < 0.05, compared with si-NC.

affect cell survival, proliferation, migration, apoptosis, angiogenesis, and immune response [22,23]. ERK/MAPK signaling pathway excessive activation leads to cell abnormal proliferation, apoptosis, and differentiation, and promote malignant transformation. It is closely associated with tumors pathogenesis, progression, and metastasis, such as oral cancer⁶, lung cancer⁷, and esophageal cancer⁸. ERK/MAPK signaling pathway mainly includes small G protein Ras, Raf kinase, MEK, and ERK. ERK/MAPK signaling pathway conforms to the classic three-step enzymatic cascade, and presents the same

activation mode under different stimulus. As a member of MAP2K family, MEK1 phosphorylates the Tyr/Thr residue of ERK protein, thus activating ERK/MAPK signaling pathway. MEK1 expression and functional activity are related to a variety of cancers pathogenesis, progression, metastasis, and drug resistance, including pancreatic cancer²⁴, bile duct cancer²⁵, and hepatic cancer⁹. It was showed that MEK1 over-expression is related to NPC cell abnormal proliferation, apoptosis reduction, and drug resistance¹²⁻¹⁴. Multiple studies revealed that miR-101 expression reduced in NPC tissue and cell line, suggesting that miR-101 may play a tumor

Table I. MiR-101 expression in NPC tissue with different characteristics.

Group	Cases	miR-101 expression	p-value
Age			0.067
≤45 years old	25	1.62±0.25	
>45 years old	40	1.53±0.22	
Gender			0.182
Male	29	1.49±0.24	
Female	36	1.55±0.28	
T stage			<0.001
T1-T2	41	1.87±0.26	
T3-T4	24	1.23±0.21	
N stage			0.173
N0-N1	31	1.57±0.27	
N2-N3	34	1.63±0.24	
M stage			0.228
M0	13	1.61±0.23	
M1	52	1.55±0.26	
Clinical stage			<0.001
I-II	28	1.92±0.29	
III-IV	37	1.18±0.25	

suppressor role in the occurrence and development of NPC^{18,19}. Bioinformatics analysis shows the complementary targeted relationship between miR-101 and the 3'-UTR of MEK1 mRNA. This study explores the role of miR-101 in regulating MEK1 expression, ERK/MAPK signaling pathway activation, and NPC pathogenesis.

Dual luciferase assay revealed that miR-101 mimic transfection significantly reduced, while miR-101 inhibitor transfection elevated the relative luciferase activity of HONE1 cells, indicating the regulatory relationship between miR-101 and MEK1 mRNA. MiR-101 significantly declined in NPC tissue compared with inflammatory nasopharyngeal tissue. MiR-101 was markedly correlated with tumor size and clinical staging. It indicates that miR-101 down-regulation may play a role in increasing MEK1 expression and facilitating NPC tumorigenesis. Tang et al¹⁹ reported that miR-101 level declined in NPC tissue compared with normal nasopharynx epithelium, following which no metastasis. In this study, miR-101 expression was reduced in NPC tissue, which was in accordance with Tang et al¹⁹. MiR-101 markedly declined, while MEK1, p-ERK1/2, and survivin apparently enhanced in NPC cell lines CNE-1, HONE1, and C666-1 compared with NP69 cells, revealing that MEK1 elevation mediates ERK/MAPK signaling pathway and survivin expression, while miR-101 decrease is its promoting factor. Sun et al¹⁸ showed that miR-101 level significantly declined in NPC cell lines

CNE-1, CNE-2, 5-8F, and 6-10B compared with NP69 cells. Tang et al¹⁹ demonstrated that miR-101 down-regulated in NPC cell lines CNE-2, 5-8F, and C666-1 compared with NP69 cells. This study found that miR-101 abnormally down-regulated in NPC cells, which was similar to Sun et al¹⁸ and Tang et al¹⁹. Further analysis revealed that miR-101 mimic and/or si-MEK1 transfection significantly reduced MEK1, p-ERK1/2, and survivin levels, attenuated cell proliferation and enhanced cell apoptosis. Alajez²⁶ revealed that pre-miR-101 transfection upregulated miR-101 expression, attenuated C666-1 cell proliferation and survival, and enhanced radiotherapy sensitivity through targeting EZR. Tang observed that miR-101 mimic transfection weakened NPC cell migration and invasion *in vitro*, and inhibited NPC cell pulmonary metastasis in mice model via down-regulating TSG3 expression¹⁹. MEK1 inhibitor transfection enhanced NPC cell migration and invasion. Sun et al¹⁸ presented that miR-101 overexpression markedly alleviated NPC cell proliferation and survival, and reduced radiotherapy resistance in CNE-2 and 5-8F cells. It confirms that miR-101 declined malignant characteristics of NPC cells, which supported the above results. Wong et al²⁷ showed that NPC cell proliferation attenuated, while cell apoptosis enhanced after treated by AZD6244 to suppress MEK1 function and ERK/MAPK signaling pathway activity, indicating that MEK1 mediated ERK/MAPK signaling pathway is related to NPC pathogenesis. Yang et al¹⁴ presented that MEK1 suppression inhibited ERK1/2 phosphorylation and ERK/MAPK signaling pathway, arrested cell cycle, and attenuated cell proliferation in NPC cells. Cheung et al¹² exhibited that application of PD098059 blocked MEK1 activity, induced NPC cell apoptosis, and enhanced sensitivity to taxol. Wang et al²⁸ demonstrated that MEK1 siRNA or U0126 treatment markedly attenuated NPC cell migration and motility, whereas MEK1 overexpression enhanced cell invasion. It suggested that MEK1 mediated ERK/MAPK signaling pathway enhancement is a promoting factor of NPC pathogenesis, while reducing MEK1 expression may inhibit NPC. This study revealed that down-regulation of miR-101 increased MEK1, enhanced ERK/MAPK signaling pathway, elevated survivin expression, and promoted NPC pathogenesis. This study only used C666-1 cells to investigate the regulatory role of miR-101 on MEK1, ERK/MAPK signaling pathway, NPC cell proliferation and apoptosis. However, we did

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