

Upregulation of long noncoding RNA DLX6-AS1 promotes cell growth and metastasis in esophageal squamous cell carcinoma *via* targeting miR-577

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Abstract. – **OBJECTIVE:** Esophageal squamous cell carcinoma (ESCC) is the most malignant type of esophageal cancer. Although significant advances have been made in ESCC diagnosis and therapy, its poor pathogenesis and prognosis remain a life-threatening problem. Meanwhile, long noncoding RNAs (lncRNAs) exert a pivotal function in tumorigenesis. In this research, we aimed to explore the association between the aberrant expression of lncRNA DLX6-AS1 and the development and metastasis of ESCC.

PATIENTS AND METHODS: DLX6-AS1 expression was monitored by quantitative real-time polymerase chain reaction (qRT-PCR) in ESCC specimens. Moreover, experiments were conducted to detect the effect of DLX6-AS1 on the cell proliferation and metastasis of ESCC. In addition, the underlying mechanism was further explored through luciferase assays and RNA immunoprecipitation assay (RIP).

RESULTS: DLX6-AS1 expression was significantly higher in ESCC specimens. Moreover, cell proliferation and metastasis of ESCC cells could be inhibited *via* reducing DLX6-AS1 expression. Besides, DLX6-AS1 was regarded as an oncogene in ESCC. Furthermore, DLX6-AS1 acted as a competing endogenous RNA *via* sponging miR-577 in ESCC.

CONCLUSIONS: In summary, DLX6-AS1 promotes development and metastasis of ESCC by sponging miR-577 and could be a potential therapeutic target.

Key Words: Long noncoding RNA, DLX6-AS1, Esophageal squamous cell carcinoma, MiR-577.

Introduction

Esophageal cancer (EC) is an ordinary malignant tumor that ranks 9th most common cancer worldwide and the fifth most frequent in China^{1,2}.

Esophageal squamous cell carcinoma (ESCC) accounts for almost 80% of EC cases in South-Eastern Asia^{3,4}. Due to the difficulty of prophylaxis and early diagnosis, most of esophageal squamous cell carcinoma (ESCC) patients develop a high rate of metastasis and recurrence with the overall 5-year survival rate below 10%⁵. It is vital in the treatment of malignant tumors to suppress ESCC proliferation and metastasis, which is the main and hot issue in the current study.

Non-coding RNAs (ncRNAs) account for approximately 70% of total transcribed RNAs. Long noncoding RNAs (lncRNAs), as a subtype of ncRNAs, are longer than 200 nucleotides. Recently, evidence has proved that lncRNAs serve as important regulators in the progression of malignant tumors. LncRNA 91H functions as an oncogene in breast cancer by up-regulating the expression of H19/IGF2 which increases aggressive phenotype of breast cancer cells⁶. LncRNA SNHG7 promotes tumor proliferation in osteosarcoma by targeting miR-34a⁷. Through interacting with miR-124, lncRNA XIST serves as an oncogene which promotes cell growth, migration, and invasion in bladder cancer⁸. Moreover, few researches also uncover the role of lncRNAs in ESCC progression. By regulating the stability of DNMT1 and depressing the expression of tumor suppressors, lncRNA LUCAT1 promotes esophageal squamous cell carcinoma formation and cell metastasis⁹. LncRNA RUNX1-IT1 acts as a tumor suppressor in esophageal squamous cell carcinoma by the inhibition of cell migration and cell proliferation¹⁰; however, the underlying mechanism of how lncRNAs function in ESCC has not been fully understood.

LncRNA DLX6-AS1 is a novel lncRNA which has been found aberrantly expressed in some malignant tumors and functions as an oncogene.

Whereas, its role in ESCC has not been studied. In this work, we found out that lncRNA DLX6-AS1 was aberrantly expressed in ESCC tissues. Moreover, the proliferation and metastasis of ESCC cells were inhibited by reducing DLX6-AS1 expression. The underlying mechanism of how DLX6-AS1 functions in ESCC development was further explored.

Patients and Methods

Cell Lines and Clinical Samples

55 ESCC tissues were enrolled from patients receiving resection surgery at the Second Hospital of Dalian Medical University. All tissues were kept at -80°C until further use. No preoperative chemotherapy or radiation was performed prior to surgery. The protocol of the study was approved by the Ethics Committee of the Second Hospital of Dalian Medical University and was performed as Declaration of Helsinki Principles required. Written informed consent was achieved before the operation.

Cell Culture

Four human ESCC cell lines (TE-1, TE-2, TE-3, EC-1, and Eca-109), and one normal human esophageal epithelial-1 cell line (HEEC) were offered by the American Type Culture Collection (ATCC; Rockefeller, MD, USA). Culture medium was Roswell Park Memorial Institute-1640 (RPMI-1640; HyClone, Logan, UT, USA) containing 10% fetal bovine serum (FBS; Gibco, Logan, UT, USA) and 1% penicillin-streptomycin at 37°C with 5% CO_2 .

Cell Transfection

We purchased lentiviral expressing short-hairpin RNA (shRNA) against DLX6-AS1 from GenePharm (Shanghai, China), which was then used for transfection of Eca-109 cells using Lipofectamine 2000 (Invitrogen, Carlsbad, CA, USA). 24 h later, cells were harvested for further experiments.

Extraction and Quantitative Real-time-Polymerase Chain Reaction (qRT-PCR)

Following the manufacturer's protocol, RNeasy lysis reagent (Invitrogen, Carlsbad, CA) was used to extract the total RNA from ESCC tissues and cells. Reverse transcription was conducted using the Reverse Transcription Kit (Ta-

KaRa Biotechnology Co., Ltd., Dalian, China). The $2^{-\Delta\Delta\text{Ct}}$ method was utilized to calculate relative expression. The primers using qRT-PCR are the following: DLX6-AS1 primers forward 5'-AGTTTCTCTCTAGATTGCC-3', reverse 5'-ATTGACATGTTAGTGCCCT-3', GAPDH primers forward 5'-CCAATATCAAGGCGGG-CAATGCTGG-3' and reverse 5'-TGATGGATGGACTGTGGTCATT-3'.

Cell Proliferation Assay

Following the manufacturer's protocol, 4×10^3 transfected cells were plated in 96-well plates and cell proliferation was assessed by the Cell Proliferation Assay Kit I (MTT; Roche, Basel, Switzerland) at 24 h, 48 h, and 72 h. The absorbance at 490 nm was assessed with an enzyme-linked immunosorbent assay (ELISA) reader system (Multiskan Ascent, LabSystems, Helsinki, Finland).

EdU Incorporation Assay

EdU (Merkel, Mannheim, Germany) was used to detect cell proliferation of transfected cells. Briefly, 6×10^3 Eca-109 cells were seeded in 96-well plate. 48 h later, the culture medium was added with 50 μM EdU for 2 h, which were then fixed with 4% formaldehyde for 30 min and incubated with 2 mg/mL Glycine for 10 min. After cells were permeabilized by 0.5% Triton X-100 for 20 min, they were added with staining liquid and Hoechst 33342. The percentage of EdU positive cells was monitored using a fluorescence microscope (Nikon Eclipse Ti Microscope, Tokyo, Japan).

Wound Healing Assay

Cells, transferred into 6-well plates, were cultured in RPMI-1640 medium overnight. Once scratched with a plastic tip, cells were cultured in serum-free RPMI-1640. 48 h later, wound closure was viewed. Each assay was repeated three times independently.

Transwell Assay

After transfection, cells (5×10^4) in 200 μL serum-free RPMI-1640 were added to the top chamber of an 8 μm pore size insert (Corning, Corning, NY, USA) with or without 50 μg Matrigel (BD; Bedford, MA, USA). RPMI-1640 and FBS were added to the lower chamber. 48 h later, the top surface of chambers was treated by methanol for 30 min after wiped by a cotton swab.

Then, they were stained in crystal violet for 20 min. Three fields were used to count the data for migration and invasion membrane.

Luciferase Assays

DLX6-AS1 3'-UTR was cloned into the pGL3 vector (Promega, Madison, WI, USA) as wild-type (WT) 3'-UTR. Next, site-directed mutagenesis of the miR-577 binding site in DLX6-AS1 3'-UTR was performed by quick-change site-directed mutagenesis kit (Stratagene, La Jolla, CA, USA) as mutant (MUT) 3'-UTR. Following was a transfection of WT-3'-UTR or MUT-3'-UTR and negative control or miR-577 for 48 h. Then, the luciferase assay was conducted on the dual luciferase reporter assay system (Promega, Madison, WI, USA).

RNA Immunoprecipitation (RIP) Assay

Magna RIP RNA-Binding Protein Immunoprecipitation Kit (Millipore, Billerica, MA, USA) was conducted according to the manuscript. Briefly, treated Eca-109 cells were lysed in the RIP lysis buffer, which was previously added with protease inhibitor and RNase inhibitor. Then, they were incubated with the RIP buffer containing magnetic beads coated with Ago2 antibodies (Millipore, Billerica, MA, USA) for 2 h at 4°C. Immuglobin G (IgG) was identified as negative control (input group). QRT-PCR was used to monitor coprecipitated

Statistical Analysis

All these results were analyzed with SPSS Product and Service Solutions, Version 19.0 (IBM Inc., Chicago, IL, USA). Independent *t*-test was

performed for comparison between the two groups (standard deviation). The statistical significance was set at $p < 0.05$.

Results

Upregulation of DLX6-AS1 in ESCC Tissues and Cells

To identify the role of DLX6-AS1 in the tumorigenesis of ESCC, we detected DLX6-AS1 expression levels in 55 pairs of ESCC and adjacent tissues by quantitative Real-time Polymerase Chain Reaction (qRT-PCR). DLX6-AS1 was markedly upregulated in ESCC tissues (Figure 1A). Meanwhile, we detected the expression level of DLX6-AS1 in ESCC cells and normal human esophageal epithelial cell (HEEC). The results showed that DLX6-AS1 expression level in ESCC cells was higher than that of HEEC (Figure 1B).

Knockdown of DLX6-AS1 Led to the Inhibition of Cell Proliferation in ESCC Cells

To further determine whether DLX6-AS1 was involved with the occurrence of ESCC, we further explored the role of DLX6-AS1 in ESCC cells. Eca-109 cell line was selected for transfection of DLX6-AS1 shRNA due to its highest level of DLX6-AS1 among four ESCC cell lines. QRT-PCR results showed that DLX6-AS1 shRNA group had a better transfection efficiency compared with control group (Figure 2A). In this work, the growth ability of Eca-109 cells was detected by methyl thiazolyl tetrazolium

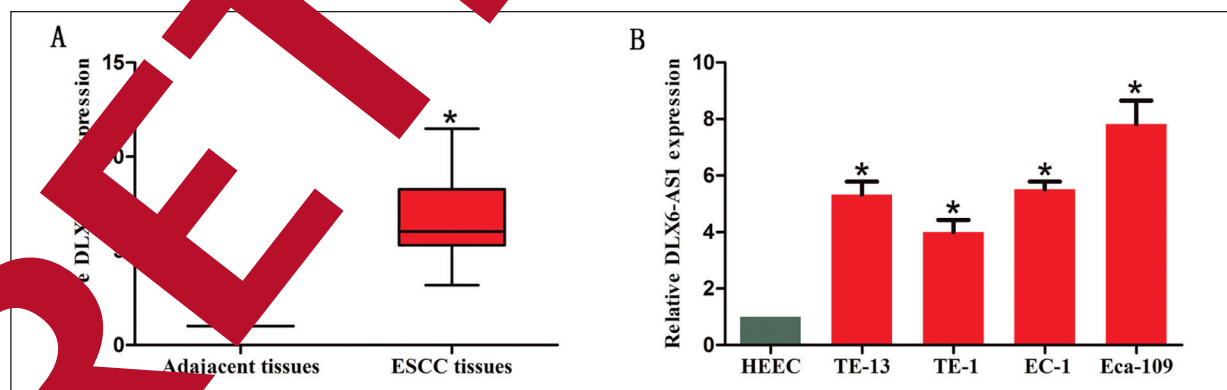


Figure 1. DLX6-AS1 expression in ESCC tissues and cells. **A**, DLX6-AS1 expression was significantly increased in the ESCC tissues compared with adjacent tissues. **B**, Expression levels of DLX6-AS1 were determined in the human ESCC cell lines and normal human esophageal epithelial-1 cell (HEEC) by qRT-PCR. GAPDH was used as an internal control. Data are presented as the mean \pm standard error of the mean. * $p < 0.05$.

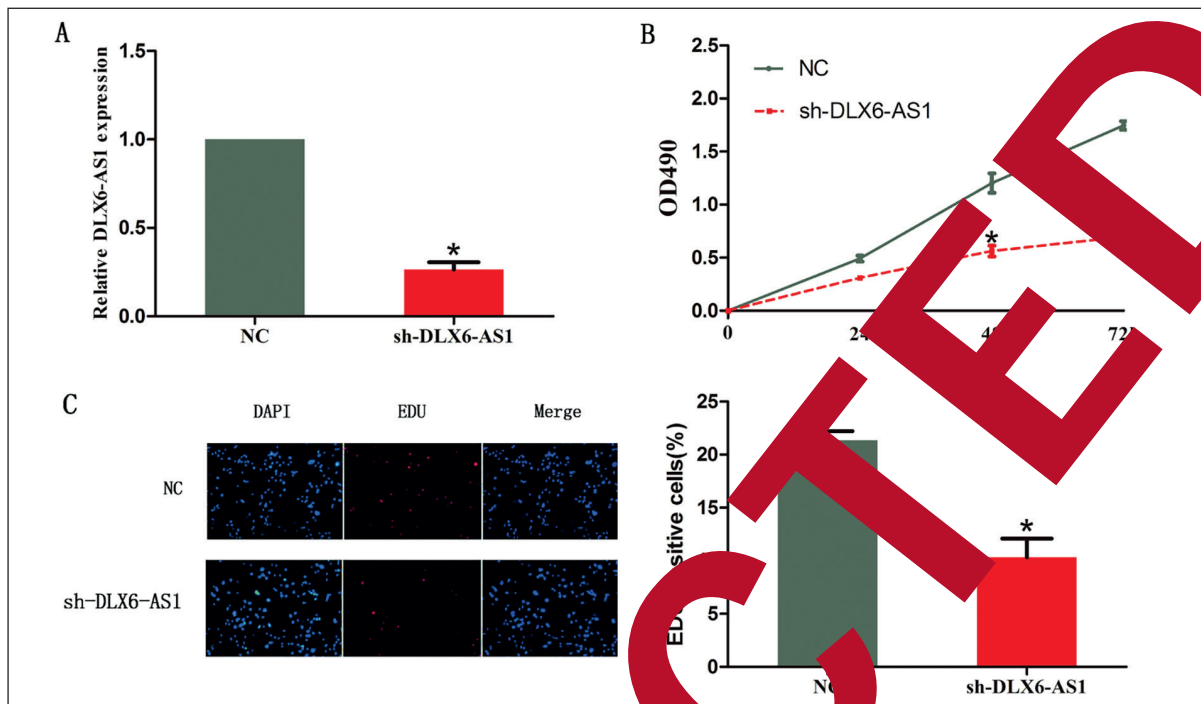


Figure 2. Knockdown of DLX6-AS1 inhibited ESCC cell proliferation. **A**, DLX6-AS1 expression in Eca-109 cells transfected with DLX6-AS1 shRNA (sh-DLX6-AS1) and negative control (NC) was determined by qRT-PCR. GAPDH was used as an internal control. **B**, MTT assay showed that knockdown of DLX6-AS1 significantly enhanced cell growth ability in Eca-109 cells. **C**, EdU incorporation assay also showed that EdU positive cells were reduced after knockdown of DLX6-AS1 in Eca-109 cells (magnification: 40 \times). The results represent the average of three independent experiments (standard error of the mean). * $p < 0.05$.

(MTT) assay after knockdown of DLX6-AS1. Results disclosed that knockdown of DLX6-AS1 considerably inhibited the growth ability of Eca-109 cells (Figure 2). Furthermore, EdU incorporation assay also indicated that EdU positive cells were reduced after knockdown of DLX6-AS1 in Eca-109 cells (Figure 2C).

Knockdown of DLX6-AS1 Led to the Inhibition of Cell Migration and Invasion in ESCC Cells

We further identified whether knockdown of DLX6-AS1 affected the metastasis of ESCC. Results of wound healing assay showed that knockdown of DLX6-AS1 inhibited the migrated length of ESCC cells (Figure 3A). Transwell assay revealed that the number of migrated and invaded cells was significantly decreased after DLX6-AS1 was knocked down in ESCC cells (Figures 3B and 3C).

miR-577 is a Direct Target of DLX6-AS1 in ESCC

Potential microRNAs of DLX6-AS1 was predicted by DIANA LncBASE Predicted v.2.

MiR-577, containing a binding area of DLX6-AS1, was chosen for our further study (Figure 4A). Then qRT-PCR results showed that the expression level of miR-577 in Eca-109 cells was significantly higher in DLX6-AS1 shRNA (sh-DLX6-AS1) group when compared with the miR-577 level in control group (Figure 4B). Furthermore, the luciferase assay revealed that co-transfection of DLX6-AS1-WT and miR-577 largely decreased the luciferase activity, while co-transfection of DLX6-AS1-MUT and miR-577 had no effect on the luciferase activity either (Figure 4C). Furthermore, the RIP assay showed that DLX6-AS1 and miR-577 were significantly enriched in Ago2-containing beads compared to input group (Figure 4D).

Discussion

Several studies¹¹⁻¹³ have suggested that lncRNAs affect carcinogenesis of ESCC by regulation of various cellular processes. LncRNA DLX6 antisense RNA 1 (DLX6-AS1), a newly discovered lncRNA, has caught more atten-

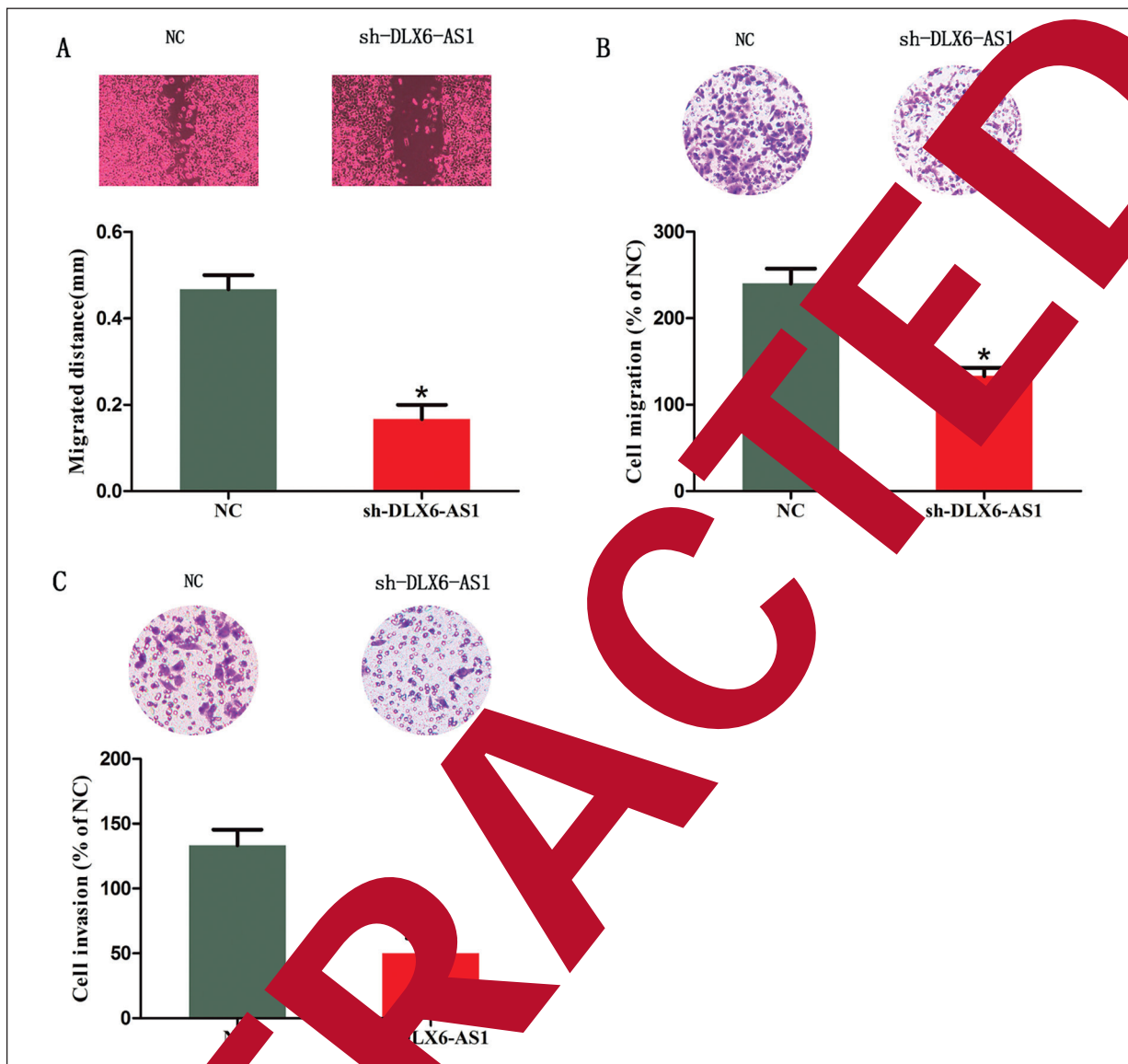


Figure 3. Knockdown of DLX6-AS1 represses ESCC cell migration and invasion. **A**, Wound healing assay showed that knockdown of DLX6-AS1 significantly reduced cell migration in Eca-109 cells (magnification 40×). **B**, Transwell assay showed that number of migrated cells was significantly decreased *via* knockdown of DLX6-AS1 in Eca-109 cells (magnification 40×). **C**, Transwell assay showed that number of invaded cells was significantly decreased *via* knockdown of DLX6-AS1 in Eca-109 cells (magnification 40×). The results represent the average of three independent experiments (mean ± standard error of the mean). * $p < 0.05$.

tion for metastasis in malignant development of cancer. Likewise, DLX6-AS1 promotes invasion of pancreatic cancer *in vitro*¹⁴. DLX6-AS1 induces osteosarcoma progressing by targeting miR-129-5p¹⁵. DLX6-AS1 promotes cell proliferation and metastasis by regulating E2F1 in glioma¹⁶. DLX6-AS1 induces the aggressiveness of renal cell carcinoma *via* targeting miR-26a¹⁷. In the current work, we

first observed that DLX6-AS1 was markedly upregulated in ESCC patients. Mechanistically, the knockdown of DLX6-AS1 inhibited cell proliferation, migration, and invasion in ESCC cells.

Latest studies¹⁸ reveal that lncRNAs function in tumor progression by binding to miRNAs. MiR-18a targeted by lncRNA CASC2 inhibits proliferation and metastasis in esophageal squa-

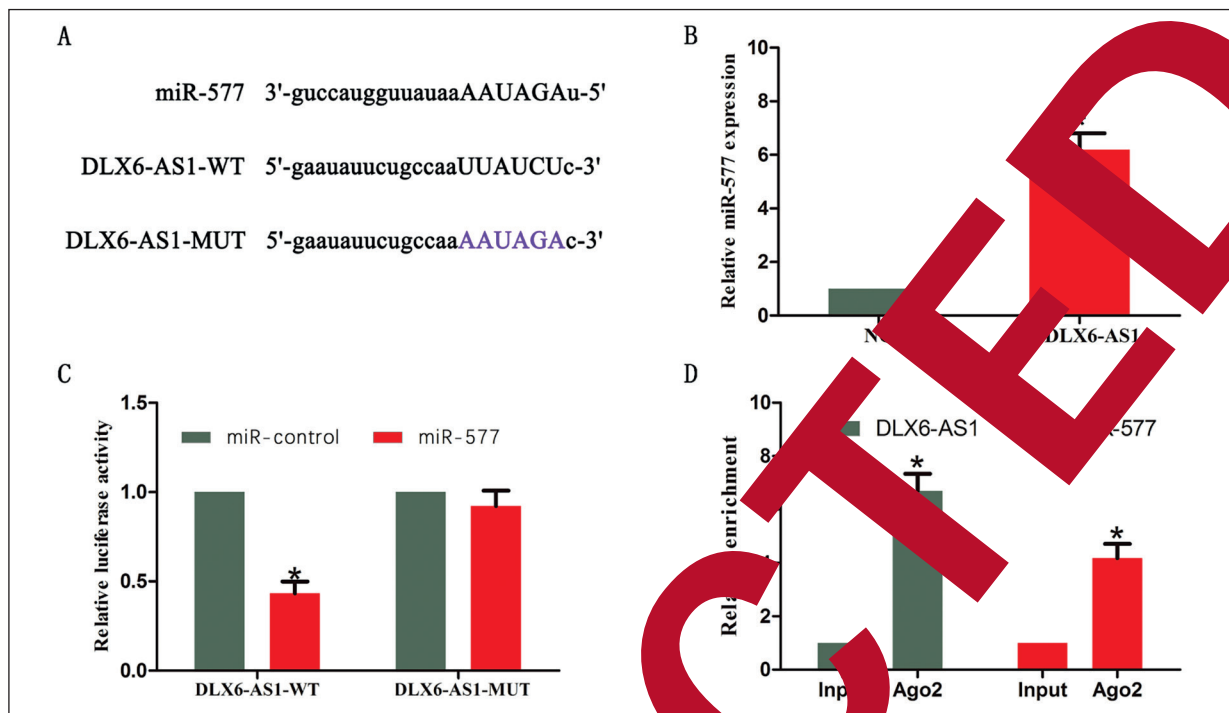


Figure 4. Interaction between DLX6-AS1 and miR-577. **A**, Binding site of miR-577 on DLX6-AS1. **B**, QRT-PCR results showed that miR-577 expression was increased in DLX6-AS1 shRNA (DLX6-AS1) group compared with negative control (NC) group. **C**, Co-transfection of miR-577 and DLX6-AS1-WT strongly decreased the luciferase activity, while co-transfection of miR-577 and DLX6-AS1-MUT did not change the luciferase activity. **D**, RIP assay identified that DLX6-AS1 and miR-577 were significantly enriched in Ago2-containing beads compared with input group. The results represent the average of three independent experiments. Data are presented as the mean \pm standard error of the mean. * $p < 0.05$.

mouse cell carcinoma. lncRNA miR-577, as miR-211 sponge, was reported to suppress the cell growth of ESCC¹⁹. The interaction between lncRNA XIST and miR-577 was demonstrated in the progression of nasopharyngeal carcinoma²⁰. Through bioinformatics analysis, miR-577 was predicted as the potential binding target RNA of DLX6-AS1. MiR-577 is a tumor suppressor in many carcinomas, participates in diverse biological processes of malignant tumors. MiR-577 enhances cell invasion via targeting Rab25 in breast cancer²¹. MiR-577 is associated with prognosis of glioblastoma patients²². Yuan et al²³ showed that abnormal expression of miR-577 and TSGA10 is associated with prognosis and development of ESCC.

In the present study, the interaction between miR-577 and DLX6-AS1 was first discovered. The results disclosed that the expression level of miR-577 was upregulated by knockdown of DLX6-AS1. Furthermore, miR-577 could directly bind to DLX6-AS1 through a luciferase assay, while miR-577 was significantly enriched by the DLX6-AS1 RIP assay.

Conclusions

Collectively, we revealed that DLX6-AS1 could promote ESCC proliferation and metastasis by sponging miR-577 and might act as a candidate target for therapy of ESCC.

Conflict of Interest

The Authors declare that they have no conflict of interests.

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